

## Supplementary Information

### Testing with $\delta^{44/40}\text{Ca}$ and $\delta^{88/86}\text{Sr}$ for ocean acidification during the early Toarcian.

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### 4.1. Sample preservation

Coherent trends of multiple palaeo-proxies through the studied interval, discussed briefly below, attest to the good preservation of the samples. The Peniche belemnites analyzed here are those used to prove the synchronicity of ammonite correlations between Peniche and Yorkshire (McArthur *et al.* 2020); the concordance of  $^{87}\text{Sr}/^{86}\text{Sr}$  data between Peniche and Yorkshire (*ibid.*) provides compelling evidence of good preservation of analyzed samples. Discussions of preservation were presented in those papers. A more detailed study of Yorkshire samples in thin-section was made by McArthur *et al.* (2007) and a more comprehensive account of belemnite preservation can be found in Saelen *et al.* (1989) and Podlaha *et al.* (1998).

We assessed the degree of preservation/alteration of our samples by visual inspection of whole samples with the naked eye and with a hand lens, and inspection of fragmented samples by eye under the binocular microscope. For belemnites in hand specimen, the diagnostic features of good preservation are clarity of calcite and a brown colour in the range light to dark. The concentric rings typically seen in transverse section, and often termed ‘growth rings’, are faint or absent in well preserved belemnites. These bands are, in fact, rings caused by diagenetic alteration (Saelen *et al.* 1989). Alteration in belemnites is patchy, so even where alteration rings are abundant, the fragmented sample usually yields unaltered material. In fragmented samples picked for analysis, the diagnostic features of good preservation are a radial fabric, transparency, and little or no colour (Fig. S1). For brachiopods, one diagnostic feature of good preservation, arising from the decay of binding organic-matter, is a readiness of the fragmented sample to flake into thin sheets, and a readiness of those sheets to disintegrate into lath-like crystallites of uncoloured, clear, calcite (Fig. S1). Our samples show these features.

Our primary means of evaluating sample quality has been visual inspection under the microscope. Nevertheless, we note that the concentrations of Ca, Sr, Mg, and Na, and values of  $\delta^{18}\text{O}$ , in our samples (Table 1) are within the range of well-preserved biogenic calcites reported in

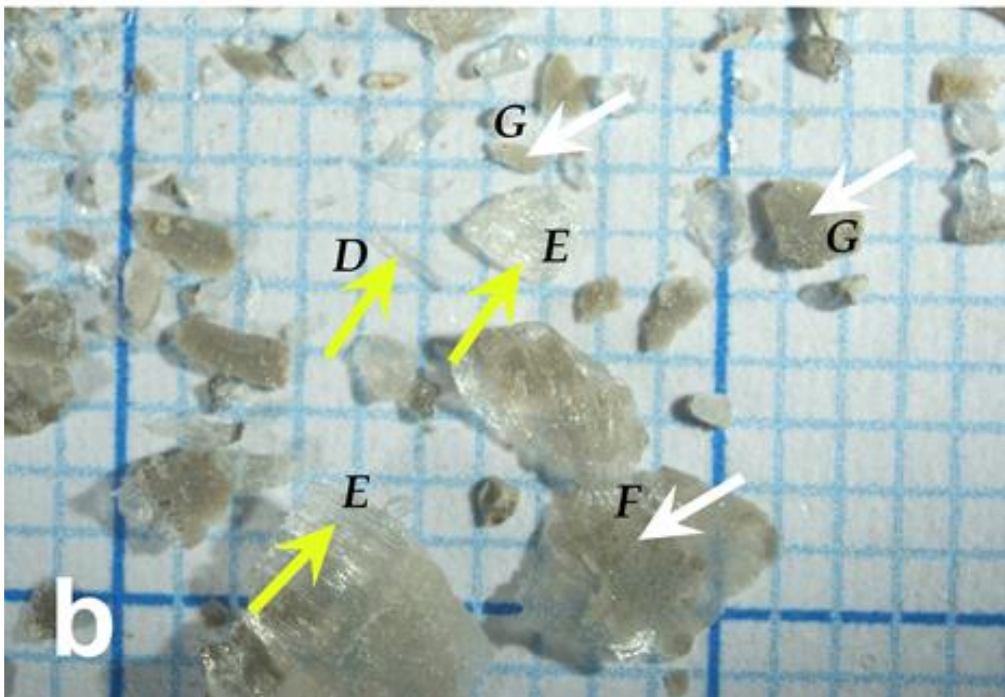
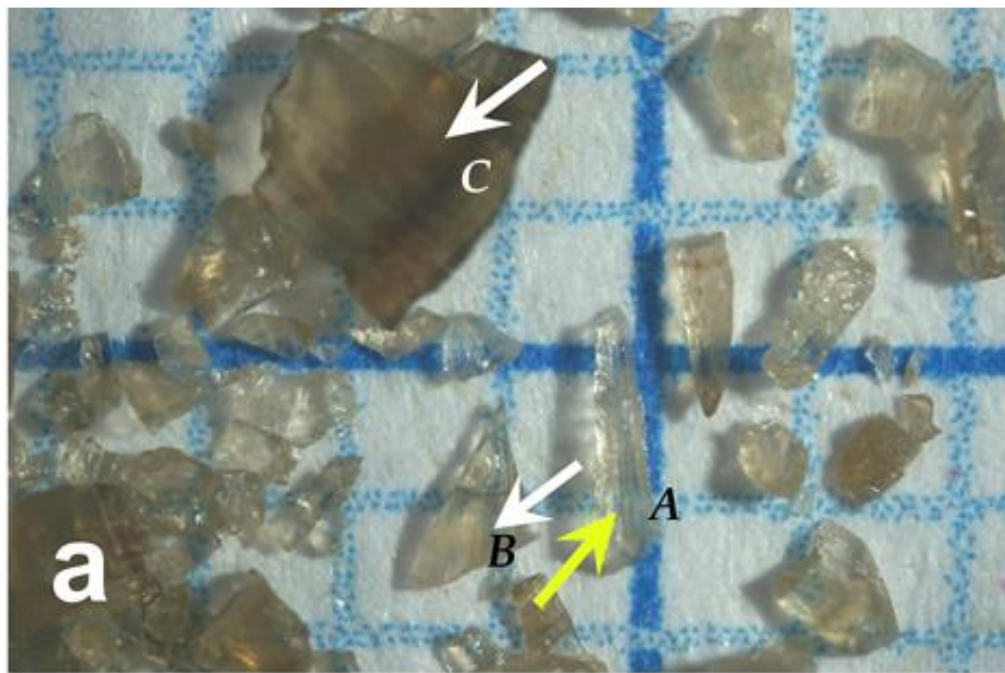


Fig. S1. Photomicrographs of fragments of a) a Peniche belemnite and b) brachiopod 9.65 m. White arrows point to altered fragments, yellow arrows point to unaltered fragments. Squares are 1 mm on a side. A, a fragment of pristine belemnite showing clear calcite with a strong radial fabric (oriented N-S); B, a triangular fragment in which pristine calcite (upper 30%) is separated from slightly altered calcite (lower 70%) by a black band that is interpreted to be organic matter. C, altered, cloudy, calcite from close to the belemnite exterior; useless for most chemical study. D, pristine crystallite of clear, pristine, calcite. E, foliated sheets of bundled crystallites of clear, pristine, calcite that flake to give thinner sheets and individual crystallites. F, foliated sheets of calcite in which the uppermost sheet is partly altered and cloudy. The upper sheet was easily flaked off from the underlying pristine calcite. G, altered calcite, the right-hand fragment being more altered than the left-hand fragment; both are useless for most chemical study.

the literature (*e.g.* Brand and Veizer 1980, 1981; Podlaha *et al.* 1998); the Ba concentrations are  $\leq 17$  ppm with only 3 samples  $> 6$  ppm; these low concentrations indicate good preservation in the sub-samples analyzed. Concentrations of Mn are  $< 20$  ppm excepting one sample containing 43 ppm and Fe concentrations exceed 100 ppm in only two samples; these higher Fe and Mn concentrations can be attributed to oxides deposited on crystal surfaces rather than incorporation into the calcite structure on alteration.

### Correlation of C and O isotopes in samples

The positive correlation between base- $\delta^{44/40}\text{Ca}_{\text{cal}}$  and temperature (Fig. 3a) might be interpreted as suggesting that isotope fractionation of oxygen into belemnites is controlled by kinetic isotope fractionation, not equilibrium isotope fractionation (McConnaughey 1989; Watkins *et al.* 2013; Daëron *et al.* 2019). Values of  $\delta^{18}\text{O}$  do not correlate with  $\delta^{13}\text{C}$  (Fig. S2, below) in any of our taxa, or within the combined group as a whole, as might occur were kinetic isotope-effects influencing oxygen isotope compositions (see also Uchikawa and Zeebe 2012). Furthermore, using paired clumped-isotopes, Bajnai *et al.* (2020) report equilibrium fractionation of oxygen isotopes into a belemnite (and by analogy, all belemnites?). In view of the above, we discount kinetic isotope fractionation as a significant factor in contributing to the scatter in the  $\delta^{18}\text{O}$  data plotted in Fig. 3a.

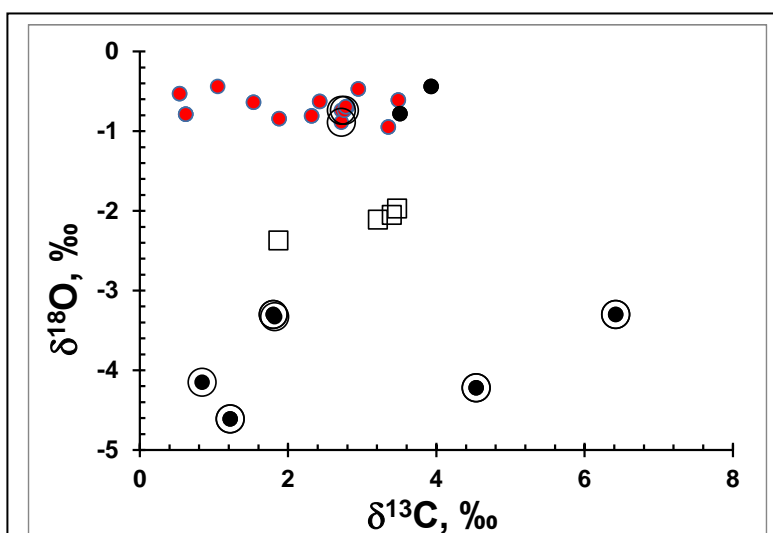


Fig. S2. Cross-plot of  $\delta^{13}\text{C}$  v  $\delta^{18}\text{O}$  for belemnites and brachiopods from Peniche, showing no relation between or within taxonomic groups

## Analytical Methods.

### *Ca isotope methodology on Triton plus TIMS at Cambridge.*

Samples of a few hundred micrograms were dissolved in 6M HNO<sub>3</sub> to make solutions with a Ca concentration of 1 µg/µl. Aliquots of 4 µl Ca were spiked with a <sup>42</sup>Ca – <sup>48</sup>Ca double spike, brought to dryness and then redissolved in 1 µl of 2M nitric acid for loading with H<sub>3</sub>PO<sub>4</sub> as an activator on a double Re-filament assembly.

Measurements were done in two steps: <sup>40</sup>Ca to <sup>44</sup>Ca, and <sup>42</sup>Ca to <sup>48</sup>Ca. Masses of <sup>40</sup>Ca, <sup>42</sup>Ca, <sup>43</sup>Ca and <sup>44</sup>Ca were measured in the first sequence and <sup>42</sup>Ca, <sup>44</sup>Ca and <sup>48</sup>Ca in the second sequence (Table S1). Cup configurations are given in Table S2. The signal from <sup>41</sup>K was below 1mV so no correction for <sup>40</sup>K was required. The <sup>40</sup>Ca signal was constrained to a 0.5 – 1 nA window using a 10<sup>11</sup> Ω resistor on collector L3. Each sample run consists of 10 blocks of 20 cycles each (for more detail of the the running protocol see Bradbury, 2018). Replicate measurements of NIST SRM-915b standard yield an average of 0.76 ± 0.12 ‰ (2sd, n = 29) on δ<sup>44/40</sup>Ca, relative to SRM-915a.

Table S1. Cup configuration for Ca isotope measurements on the Triton plus TIMS at Cambridge.

CUPS	L4	L3	L2	L1	C	H1	H2	H3	H4
STEP 1		<sup>40</sup> Ca	<sup>41</sup> K		<sup>42</sup> Ca	<sup>43</sup> Ca	<sup>44</sup> Ca		
STEP 2	<sup>42</sup> Ca			<sup>44</sup> Ca					<sup>48</sup> Ca

### *Ca isotopic analysis on IsotopX Phoenix-X62 TIMS at Royal Holloway*

Sample were dissolved in 6M HNO<sub>3</sub> to make solutions with a Ca concentration of 1 µg/µl. Aliquots of 1 – 2 µl were loaded onto a single Re-filament in nitrate with TaF<sub>5</sub>- H<sub>3</sub>PO<sub>4</sub> emitter with the aid of parafilm dams to minimize sample spread. A <sup>43</sup>Ca – <sup>46</sup>Ca double spike was used to correct for mass fractionation. Isotopic abundances of Ca were measured in a static mode in one sequence shown in Table S2. Neither <sup>87</sup>Sr<sup>2+</sup> nor Ti ions were observed in our samples. Running the Sr standard SRM987 whilst monitoring masses 40 – 44 detected no interference from Sr. Interference on <sup>40</sup>Ca from <sup>40</sup>K was monitored by measuring mass 41 and using the <sup>40</sup>K/<sup>41</sup>K ratio of 0.0017384 for correction, but the <sup>40</sup>K correction was only 1 – 2 ppm.

With a Ca load of 1 – 2 µg, the <sup>40</sup>Ca signal was constrained to a 1.5 – 2 nA window using a 10<sup>10</sup>Ω resistor on collector L7. Each sample run contains 20 blocks, 10 cycles each, with 10 s integration time for each peak. Baselines were measured at half masses with 10 s integration time before and after each peak. The measured isotopic ratios of unspiked Ca isotopic standard HPSnew were then normalized to a <sup>42</sup>Ca/<sup>44</sup>Ca of 0.30886 (Jorg *et al.* 2012). Analyses of unspiked HPS<sub>new</sub> yielded mean values of 46.134855 ± 0.00096 (2se) for <sup>40</sup>Ca/<sup>44</sup>Ca, 0.064519 ± 0.00001 (2se) for <sup>43</sup>Ca/<sup>44</sup>Ca, and 0.001528 ± 0.00001 (2se) for <sup>46</sup>Ca/<sup>44</sup>Ca. and a mean δ<sup>44/40</sup>Ca value of 0.71 ± 0.20 ‰ (2 s.d. n = 11) relative to SRM-915a, consistent with its published values (Reynard et al., 2010 and Li et al., 2016).

Table S2. Cup configuration for Ca isotope measurements on the Phoenix at RHUL

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CUPS	L7	L4	L3	L2	AX	H1	H2	H3	H4
MASS	<sup>40</sup> Ca		<sup>41</sup> K	<sup>42</sup> Ca	<sup>43</sup> Ca	<sup>87</sup> Sr <sup>2+</sup>	<sup>44</sup> Ca		<sup>46</sup> Ca

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140 The <sup>43</sup>Ca – <sup>46</sup>Ca double spike (DS) solution was made from stocks of OakRidge <sup>43</sup>Ca and <sup>46</sup>Ca  
141 spikes. Its calibrated isotopic compositions are listed in Table S3 and the concentration of each  
142 isotope in the DS was 1ng/g. Routinely, 1 µg of Ca in a sample was mixed with 10 µl of <sup>43</sup>Ca-<sup>46</sup>Ca  
143 DS. Measured Ca-isotopic ratios of sample-spike mixtures were corrected offline for mass  
144 fractionation using a Matlab model (see Li *et al.*, 2016)

145

146 Table S3: Ca isotopic ratios and abundance of <sup>43</sup>Ca–<sup>46</sup>Ca double spike (DS) solution; the isotopic  
147 abundances were calculated from the masses of each spike in the DS solution and their certified  
148 values of abundance.

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Isotopic ratios	43-46Ca DS	Isotopic abundance* (%)	43-46Ca DS
<sup>40</sup> Ca/ <sup>44</sup> Ca	9.256967 ± 0.000668	<sup>40</sup> Ca	45.4753
<sup>42</sup> Ca/ <sup>44</sup> Ca	0.136588 ± 0.000005	<sup>42</sup> Ca	0.7754
<sup>43</sup> Ca/ <sup>44</sup> Ca	4.328696 ± 0.000075	<sup>43</sup> Ca	25.4655
<sup>46</sup> Ca/ <sup>44</sup> Ca	4.470016 ± 0.000151	<sup>44</sup> Ca	5.5057
		<sup>46</sup> Ca	21.5758
		<sup>48</sup> Ca	1.1860

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### S3. Sr isotope method

Samples were dissolved in sub-boiled 8 M HNO<sub>3</sub>, purified using Sr-Spec resin, and loaded as nitrate on a single Re filament with TaF<sub>5</sub> - H<sub>3</sub>PO<sub>4</sub> emitter. An <sup>87</sup>Sr - <sup>84</sup>Sr double spike solution was used to correct for mass fractionation and to determine the true Sr isotope ratios of samples. For isotopic measurement, the machine was run in multi-dynamic mode with correction for <sup>87</sup>Rb. Spiked and unspiked samples were prepared and run separately for δ<sup>88/86</sup>Sr and <sup>87</sup>Sr/<sup>86</sup>Sr analysis. A <sup>87</sup>Sr – <sup>84</sup>Sr spike solution was prepared from two single isotopically enriched Sr-carbonates (<sup>87</sup>Sr-spike, <sup>84</sup>Sr-spike) from Oak Ridge National Laboratory (ORNL). The two carbonates were mixed to contain 43.5 % of <sup>87</sup>Sr spike solution and 56.5 % of <sup>84</sup>Sr spike solution, giving a <sup>87</sup>Sr/<sup>84</sup>Sr ratio (0.87344) close to the optimum value of 0.8139 from the double spike inversion theory of Rudge *et al.* (2009) for similar ORNL <sup>87</sup>Sr – <sup>84</sup>Sr double spikes. The purpose was to optimize the effect of double spike and minimize the propagated errors on mass-fractionation corrected Sr isotope ratios.

The true Sr isotopic composition of the double spike solution was determined from the certified isotopic ratios of Sr standard SRM-987 and the measured ratios of <sup>87</sup>Sr – <sup>84</sup>Sr DS and SRM-987 – DS mixtures, by treating the <sup>87</sup>Sr – <sup>84</sup>Sr DS as the ‘unknown’ and the SRM-987 standard as the ‘spike’. Data correction closely followed the Pb DS algorithm developed by Thirlwall (2000). The defined isotopic composition and isotope abundance (calculated) of the <sup>87</sup>Sr – <sup>84</sup>Sr spike solution are shown in Table S4.

Table S4: Sr isotopic ratios and abundance of <sup>84</sup>-<sup>87</sup>Sr double spike (DS) solution

Isotopic ratios	84-87Sr DS	Isotopic abundance* (%)	84-87Sr DS
<sup>84</sup> Sr/ <sup>86</sup> Sr	15.80030	<sup>84</sup> Sr	46.47
<sup>87</sup> Sr/ <sup>86</sup> Sr	13.80063	<sup>86</sup> Sr	2.45
<sup>88</sup> Sr/ <sup>86</sup> Sr	3.83797	<sup>87</sup> Sr	40.58
<sup>87</sup> Sr/ <sup>84</sup> Sr	0.87344	<sup>88</sup> Sr	10.50



# Sensitivity Analysis for Variations in Salinity and $\delta^{18}\text{O}$ .

The profiles of  $\delta^{44/40}\text{Ca}$  against stratigraphic level (Fig. 5) show a positive excursion in Interval 2. The excursion occurs in raw data and in data corrected for temperature using a value of  $\delta^{18}\text{O}_{\text{sw}} = -1\text{‰}$ . Here we show in Fig. S3 the effect of varying  $\delta^{18}\text{O}_{\text{sw}}$  between  $+1\text{‰}$  and  $-2\text{‰}$  for seawater in Yorkshire whilst maintaining a value of  $-1\text{‰}$  for Peniche. Other values can be interpolated or extrapolated from these values. Whatever values are chosen, the positive excursion in  $\delta^{44/40}\text{Ca}$  remains in Interval 2.

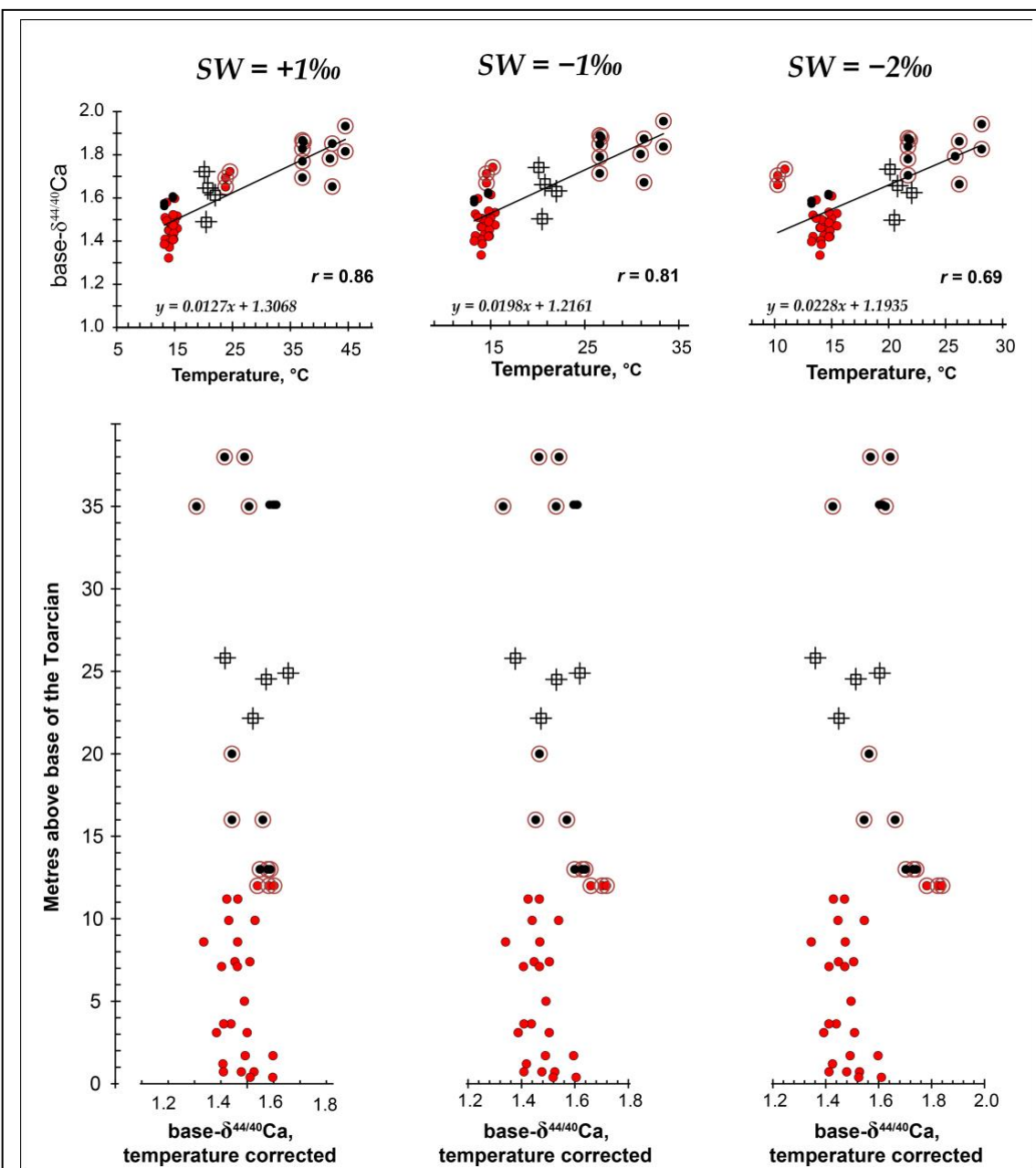
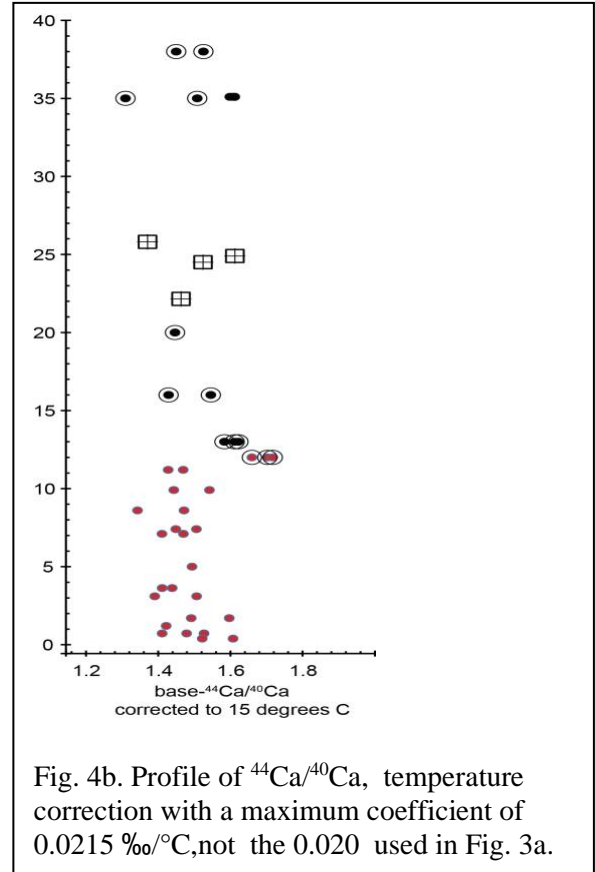
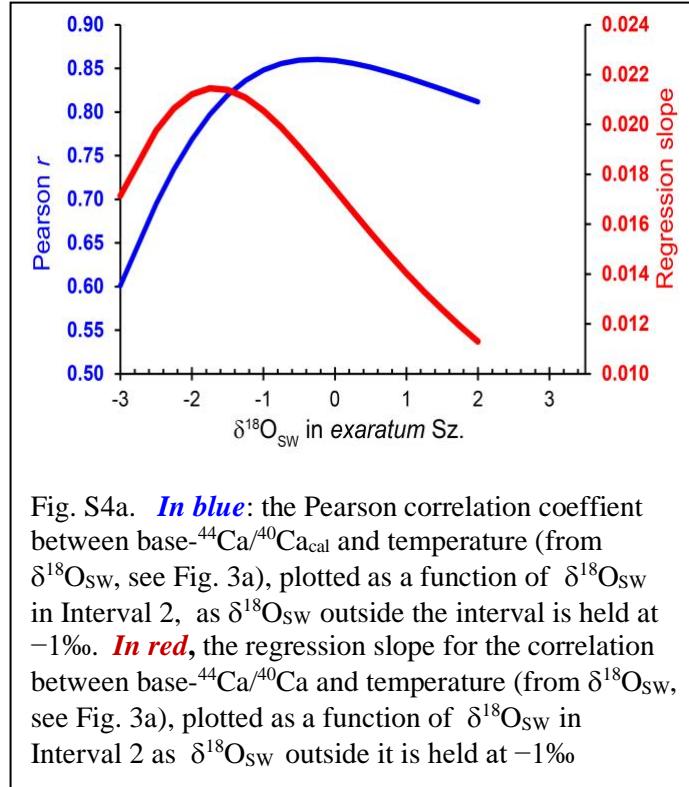


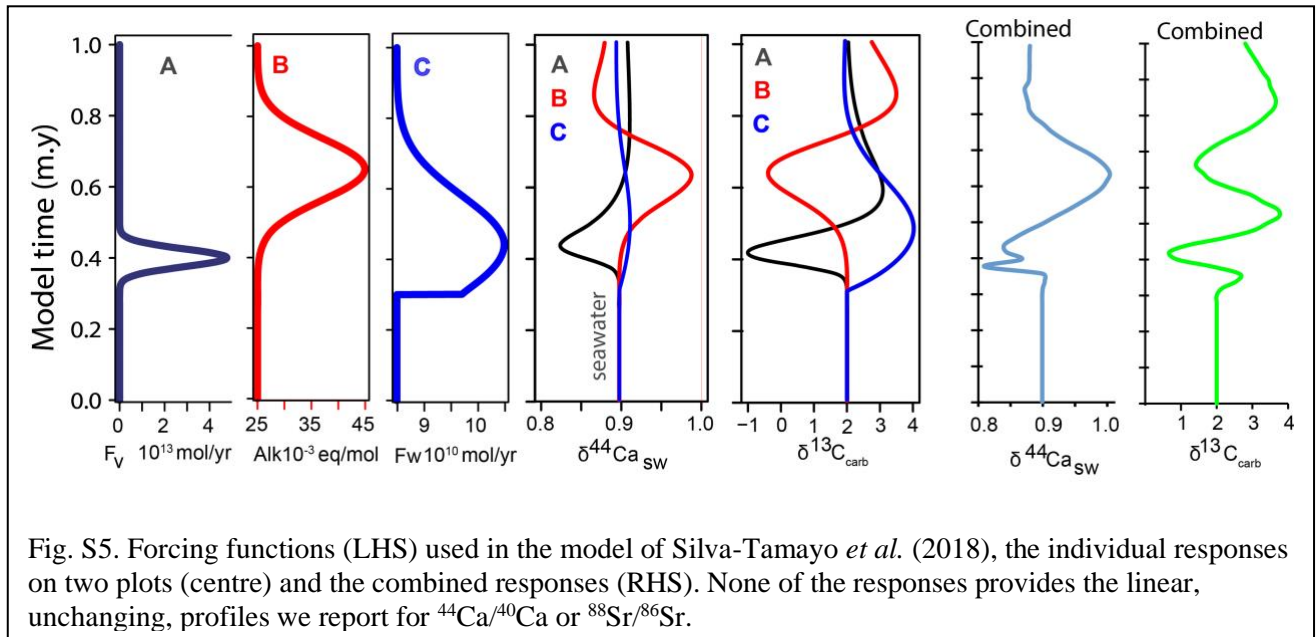
Fig. S3. Effect of differing values of  $\delta^{18}\text{O}_{\text{sw}}$  on the temperature dependence of  $\delta^{44/40}\text{Ca}$  (as shown in Fig. 3a (upper three diagrams) and on the stratigraphic profile of  $\delta^{44/40}\text{Ca}$  as shown in Fig. 6 (lower three diagrams). Symbols as on previous figures.

Shown in Fig. S4a is a summary of the effect of varying  $\delta^{18}\text{O}_{\text{sw}}$  between +2‰ and -3‰ for seawater in the *exaratum* Sz. whilst maintaining a value of -1‰ outside it. The maximum slope of the regression line is 0.0215. The maximum Pearson correlation coefficient is 0.86. Whatever values between -3‰ and +2‰ are chosen for  $\delta^{18}\text{O}_{\text{sw}}$  in the *exaratum* Sz., the positive excursion in  $\delta^{44/40}\text{Ca}$  remains in Interval 2 (Fig. S4b).



### Combined models of Silva Tamayo et al. (2015)

In Fig. S5 we show the isotopic profiles that result from combining the models of Silva-Tamayo *et al.* (2015). The combined profiles do not match our measured profiles.





276 **Data of Müller *et al.* (2020), M20 hereinafter.**

277 Muller et al. (2020) report profiles through Peniche of  $\delta^{11}\text{B}$  and interpret them as showing ocean  
278 acidification in correlative equivalent of the *exaratum* Sz. Here we examine the robustness of the  
279 data on which that conclusion is based.

280  
281 **Sample selection**

282 A detailed curve of  $\delta^{13}\text{C}$  in bulk sediment through the Peniche section was provided by Hesselbo *et al.*  
283 (2007), who showed that belemnite  $\delta^{13}\text{C}$  closely tracked bulk-sediment  $\delta^{13}\text{C}$  but was offset to  
284 higher values by 0 to 1.5‰. Similar tracking and offsets were found by McArthur *et al.* (2020)  
285 for belemnites, by  
286 Suan *et al.* 2008 for  
287 brachiopods. Of the  
288 brachiopods of M20,  
289 all but P48, at 17.42 m  
290 above datum, track  
291 bulk carbonate. Sample  
292 P48 (arrowed in Fig.  
293 S6) has a  $\delta^{13}\text{C}$  value  
294 of +3.18‰, which is  
295 too for its stratigraphic  
296 position. The value is  
297 3.42‰ heavier than  
298 sample P49, at 17.52 m,  
299 but was not analysed  
300 for  $\delta^{11}\text{B}$ . The  
301 stratigraphic position  
302 of P48 is incorrect, so  
303 its value of  $\delta^{11}\text{B}$  should be discounted.

304  
305  
306 **Sample Preservation: Sr-isotopes.**

307 The excessively high values of  $^{87}\text{Sr}/^{86}\text{Sr}$   
308 of all but two samples of M20 analysed  
309 for  $^{87}\text{Sr}/^{86}\text{Sr}$ , relative to expected values  
310 (Fig. S7), shows that the Sr-isotope  
311 system in almost all samples of M20  
312 analysed for  $^{87}\text{Sr}/^{86}\text{Sr}$  has been disturbed.  
313 It is unlikely that the Sr-isotope  
314 system has been disturbed whilst the B-isotope  
315 system has not, so the  $\delta^{11}\text{B}$  values are  
316 suspect.

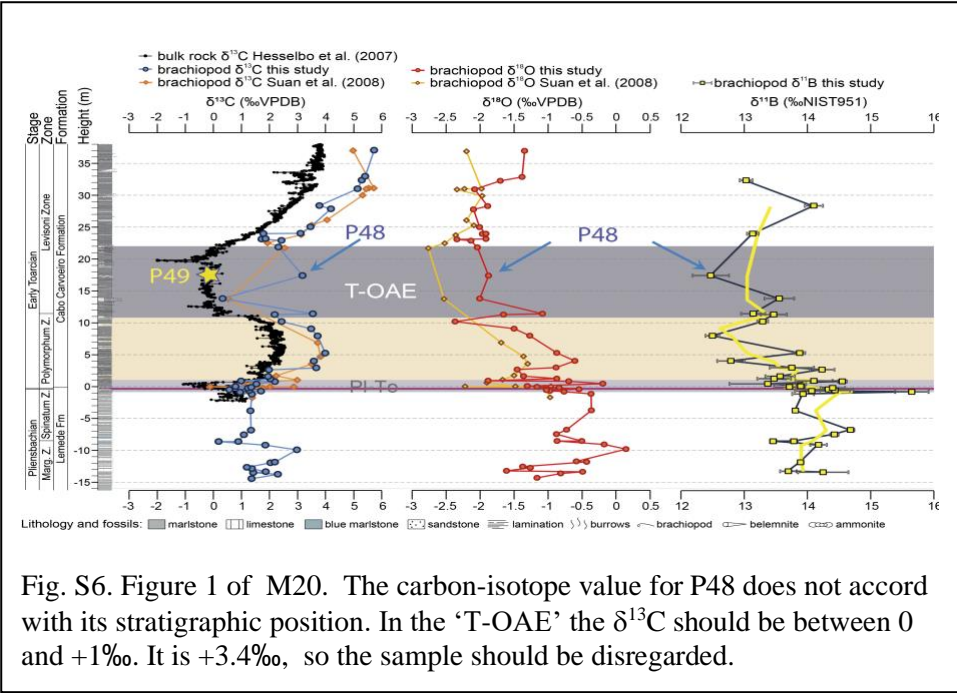


Fig. S6. Figure 1 of M20. The carbon-isotope value for P48 does not accord with its stratigraphic position. In the 'T-OAE' the  $\delta^{13}\text{C}$  should be between 0 and +1‰. It is +3.4‰, so the sample should be disregarded.

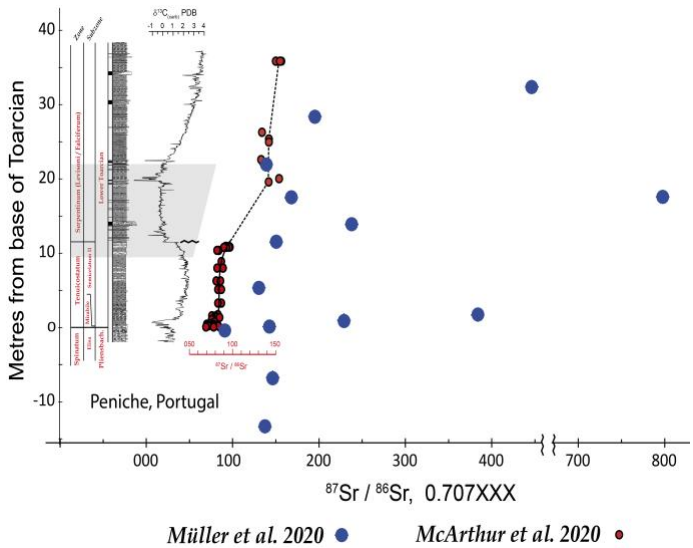


Fig. S7. Comparison of  $^{87}\text{Sr}/^{86}\text{Sr}$  profiles for Peniche from M20 and from McArthur *et al.* (2020); the former are higher by up to 0.000 650 and so plainly altered.

### Sample preservation: aluminium concentrations

The samples of M20 contain up to 4.1 % of Al (Supplementary Information of M20) and correlate with concentrations of Mg (Fig. S8). The correlation shows the presence of a contaminant phase with a Mg/Al mass-ratio of 0.75. The Mg/Al mass ratio of 0.75 is within the wide range reported for diagenetic palygorskite. The presence of such a contaminant phase (or others that are unidentified) provides a ready explanation of why  $^{87}\text{Sr}/^{86}\text{Sr}$  values of many samples are so high – they are reflecting not just Sr in biogenic calcite but also Sr in contaminant Al-rich phases.

It is therefore no surprise that concentrations of Al correlate with  $\delta^{11}\text{B}$  (Fig. S9).

To find useable data in M20, there are two ways to filter out samples that appear altered. Firstly, by discarding samples with aberrant  $^{87}\text{Sr}/^{86}\text{Sr}$ . Secondly, by discarding all samples with Al concentrations  $> 1000$ . Doing both is, perhaps preferable and leaves no samples between 0 and 22 m in the section, making it impossible to identify any trend in the critical Interval 2 (Fig.

S10).

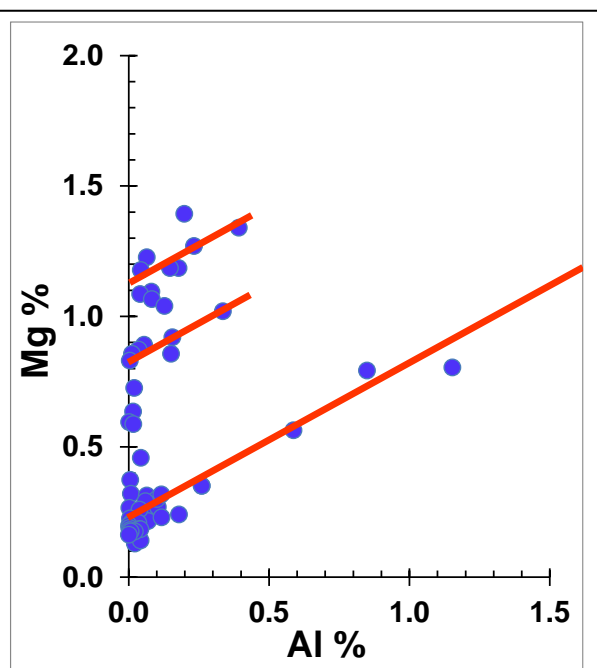


Fig. S8. Al v Mg in samples analysed by M20 for  $\delta^{11}\text{B}$ . Red squares are samples for which Mg and Al correlated strongly. Two samples not shown contain 2.1 % and 4.1 % Al and fall on the lowest red correlation line but were not used by M20 for pH-reconstruction.

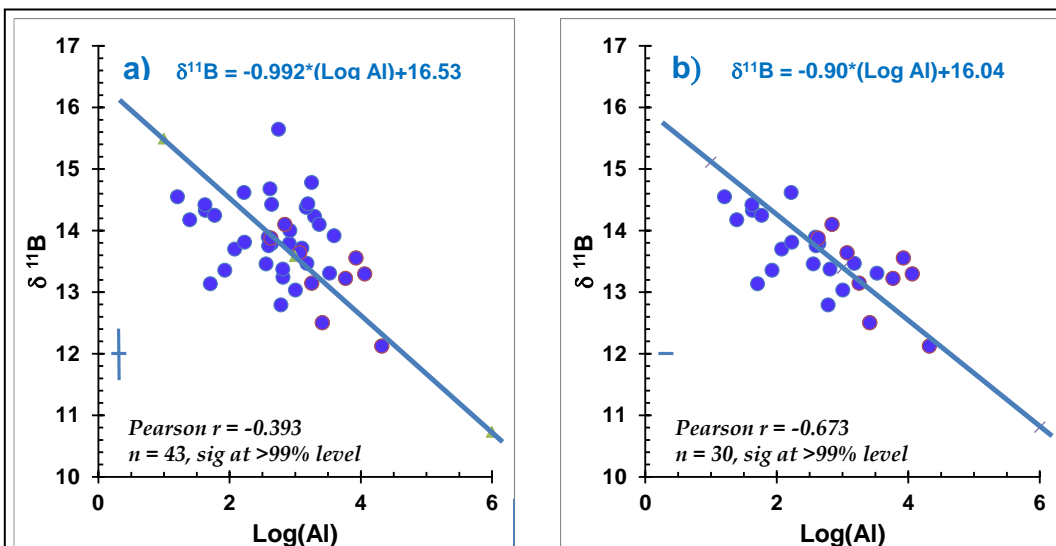
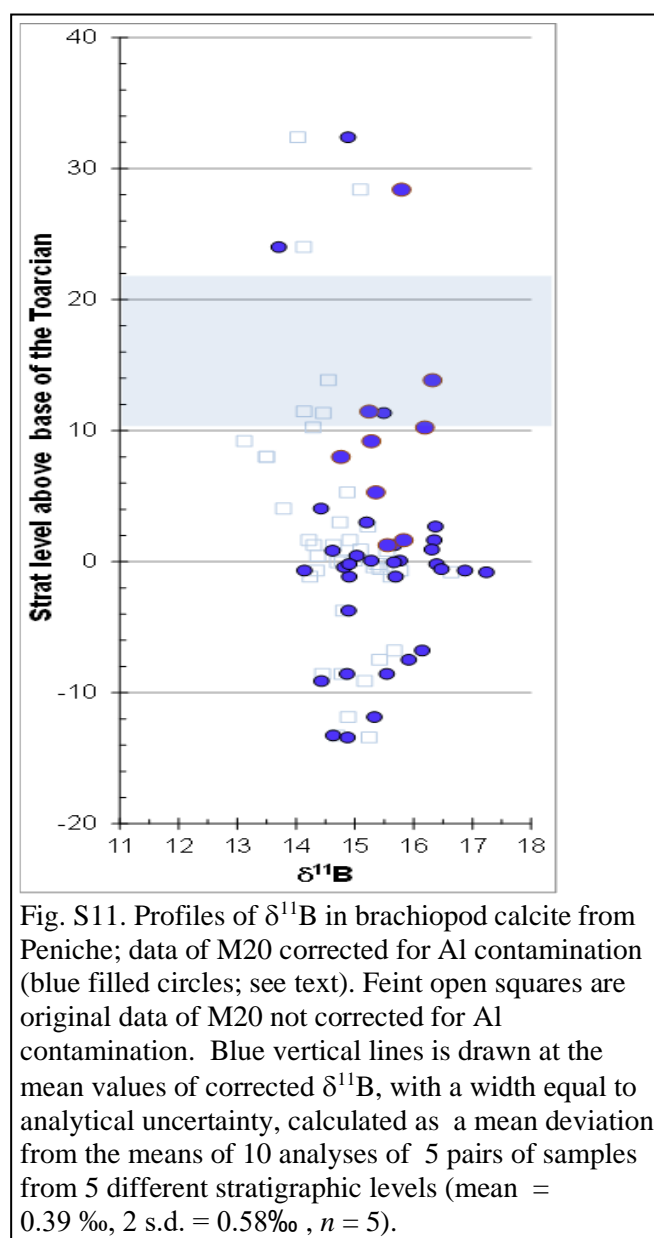
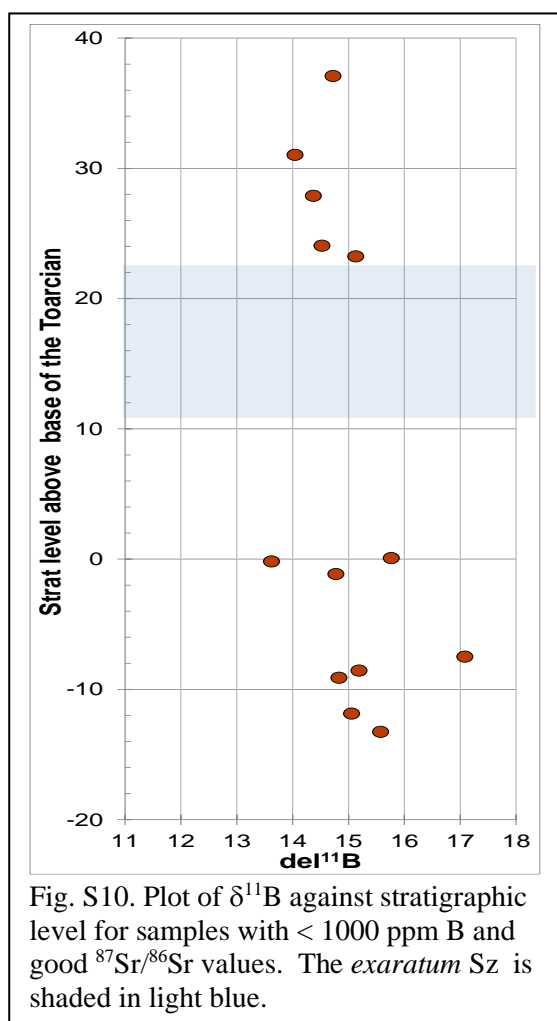


Fig. S9. Relation of Al to  $\delta^{11}\text{B}$  in the data of M20. a) all data; b) excluding *Liospiriferina*



An alternative to discarding altered samples is to correct for the effect of Al-contamination on  $\delta^{11}\text{B}$  using the correlation between Al concentrations and  $\delta^{11}\text{B}$  (Fig. S9a). In practice, some caution is needed in doing so. Firstly, the correlation line in Fig. S9a extrapolates to a value of around  $11.5\text{‰}$  for  $\delta^{11}\text{B}$  at  $10^5 \mu\text{g/g}$  of Al, which is approximating 100% Al-bearing contaminant. Marine clays typically have negative  $\delta^{11}\text{B}$ . The slope of the correlation line is therefore less than expected were the contamination to be due to the presence of detrital clay, a consideration that strengthens our view that the contamination reflects precipitation of Al in diagenetic palygorskite.

Whatever the contaminant phase(s), its influence on samples will depend on the relative concentrations of B in contaminant and sample and their respective  $\delta^{11}\text{B}$ . Alteration will have more effect on samples with low concentrations of B (e.g. *Soaresirhynchia*, 11 to  $36 \mu\text{g/g}$ ) than on samples with high concentrations (*Liosiriferina*; 743 to  $1585 \mu\text{g/g}$ ). Indeed, it might be that the concentrations of B in *Liospiriferina* are too high to be much altered by diagenesis.

To accommodate this observation, and refine the possible influence of alteration on  $\delta^{11}\text{B}$ , we show in Fig. S9b samples without *Liospiriferina*, an exclusion that improves the correlation of Al with

386  $\delta^{11}\text{B}$ . Nevertheless, we show in Fig. S11 the stratigraphic profile of  $\delta^{11}\text{B}$  corrected using the  
 387 relationship shown in Fig. S9a; there appears to be no unequivocal change in  $\delta^{11}\text{B}$  through the  
 388 interval studied so, we conclude, there is no evidence for ocean acidification in the  $\delta^{11}\text{B}$   
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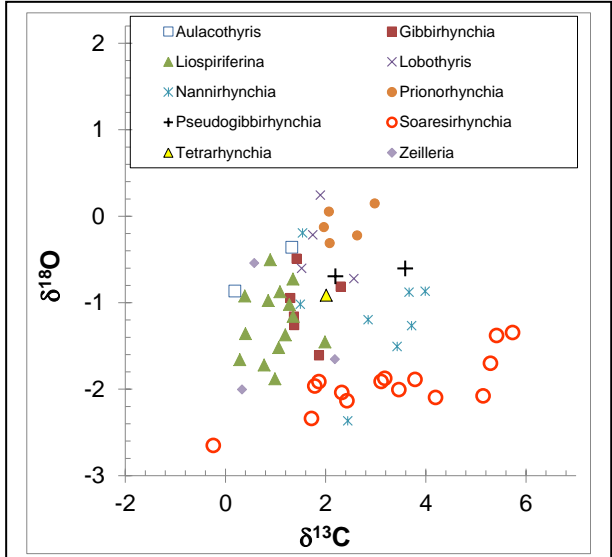
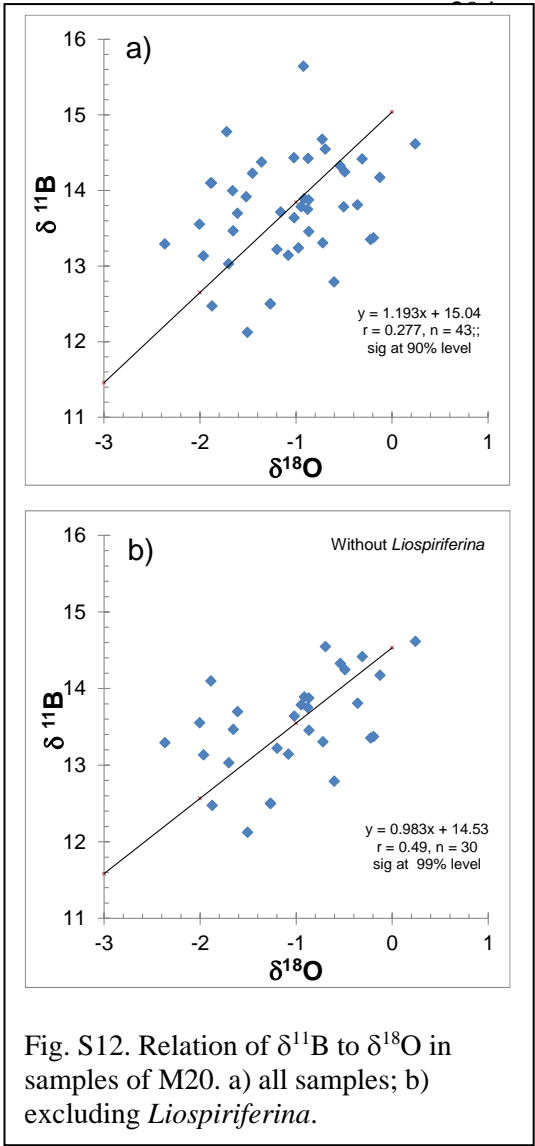
390 **Sample preservation: relation to  $\delta^{18}\text{O}$**

391 M20 note that, through the section, “The changes in  $\delta^{11}\text{B}$  closely follow the  $\delta^{18}\text{O}$  record ( $r = 0.75$ ,  $p$   
 392  $<0.0001$ ).....”. A correlation is confirmed by a cross-plot of  $\delta^{18}\text{O}$  and  $\delta^{11}\text{B}$  (Fig. S12), but the  
 393 correlation is barely significant (90% level) when all

samples are included (Fig. S10a). It is significant at the 99% level when *Liospiriferina* is omitted from the plot (Fig. S12b).

Penman *et al.* (2013) noted a positive relation in brachiopods between  $\delta^{11}\text{B}$  and  $\delta^{18}\text{O}$ , with  $\delta^{11}\text{B}/\delta^{18}\text{O}$  between 1.4 and 2.4. In Fig. S12 the values are 1.2 and 1.0. The relationships suggest that the incorporation of B isotopes into brachiopod calcite is controlled, in part, by vital effects, and/or the temperature and/or the rate of calcification.

Penman *et al.* (2013) also noted that in brachiopods “intra-shell variability up to 7‰ was observed in  $\delta^{13}\text{C}$  and  $\delta^{18}\text{O}$  possibly because of kinetic isotope fractionation effects (Carpenter and Lohmann, 1995; Auclair *et al.*, 2003)...”. Only for M20’s brachiopod species *Soaresirhynchia* do  $\delta^{13}\text{C}$  and  $\delta^{18}\text{O}$  correlate well (orange open circles on Fig. S13), suggesting either alteration, or a kinetic effect on  $\delta^{13}\text{C}$  and  $\delta^{18}\text{O}$  for this species. A rate effect on  $\delta^{11}\text{B}$  may therefore affect *Soaresirhynchia*.



417 ***Species bias.***

418 Penman *et al.* (2013), in summarising previous work on  $\delta^{11}\text{B}$  brachiopods, noted that “*Boron*  
419 *isotope data from various modern species of brachiopods exhibit a large (15–23‰) range*  
420 *(Hemming and Hanson, 1992; Lécuyer et al., 2002; Joachimski et al., 2005; Simon et al., 2006),*  
421 *which is too great to be attributed solely to regional pH variations.*”.

422 Such observations undermine attempts to use brachiopods for pH reconstruction. We speculate that  
423 the large spread in the data of M20 for any particular stratigraphic level arises both from alteration  
424 and from species differences in values of  $\delta^{11}\text{B}$ . For example, samples P22 and P23, both recovered  
425 from a position 71 cm below the Pl-Toa boundary, have values that differ by 1.43 ‰  
426 (*Liospiriferina rostrata* = 14.78 ‰; *Prionorhynchia serrata* = 13.35 ‰).

428 ***Intra-species variation***

429 Jurikova *et al.* 2019, noted that “*...boron incorporation into the innermost shell layers (of*  
430 *brachiopods) is primarily driven by calcifying fluid pH, whereas the outermost calcite layers are to*  
431 *some extent influenced by ambient seawater pH.*”. Such an observation accords with the notion that  
432 the outer (primary) shell of brachiopods is deposited out of equilibrium with ambient seawater  
433 whilst the inner (secondary) shell approaches isotopic equilibrium (Carpenter and Lohmann 1995).  
434 This view is confirmed by Bajnai *et al.* (2018) for brachiopods. Non-equilibrium precipitation of  
435 calcite is seen in the range of 3.5 ‰ range in  $\delta^{11}\text{B}$  across individual brachiopod shells reported by  
436 Penman *et al.* (2013) and in the correlation of values of  $\delta^{11}\text{B}$  with  $\delta^{18}\text{O}$  in brachiopod shells (*ibid.*).  
437 It is not certain that such influences are absent in the subsamples of M20.

438  
439  
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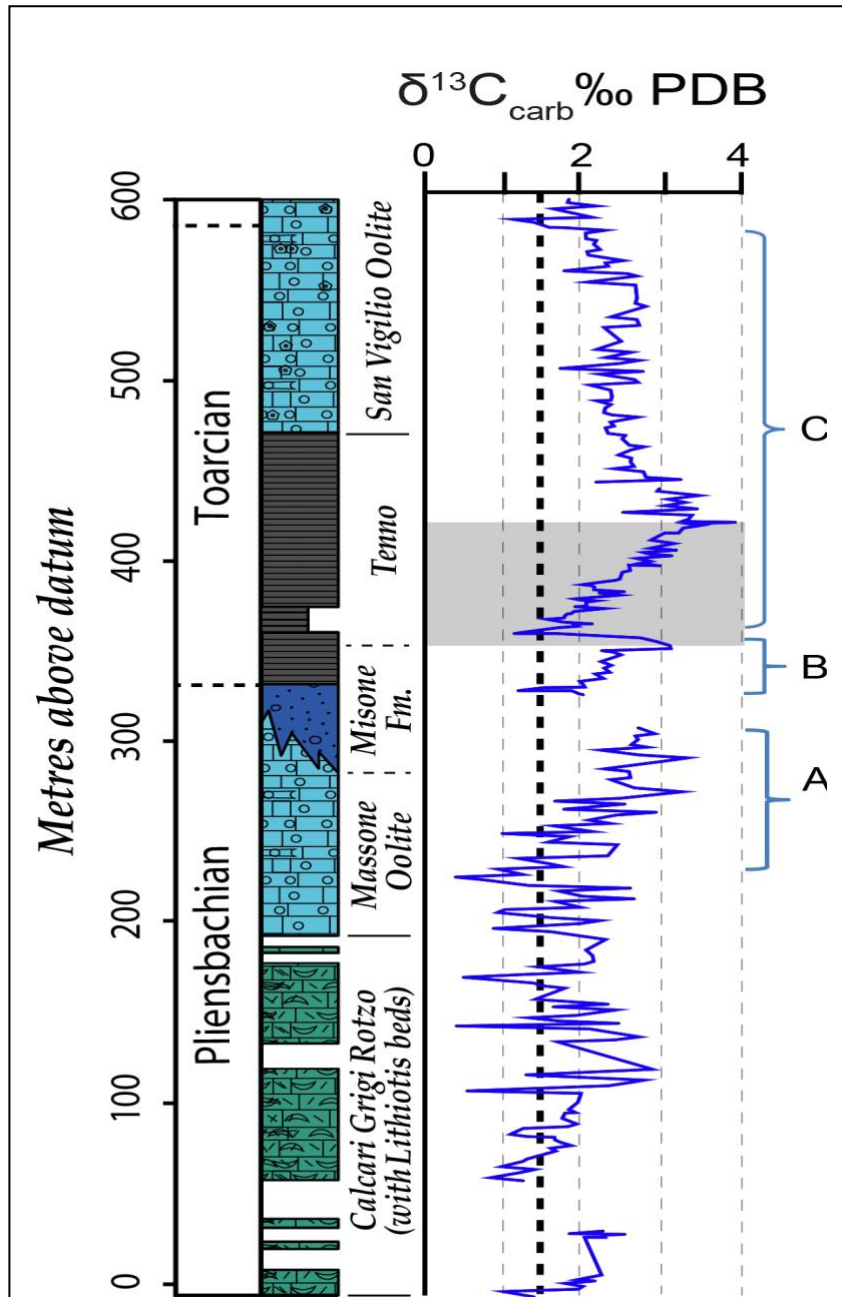


Fig. S14. Trend of  $\delta^{13}\text{C}_{\text{carb}}$  through the Trento Platform, after Ettinger *et al.* (2021). Grey area denotes the putative T-OAE as it is identified by those authors. Between A and B, and between B and C, values of  $\delta^{13}\text{C}_{\text{carb}}$  return to around +1‰ to +2‰, which are interpreted as typical (background) values for early Mesozoic times. Heavy dashed line drawn at +1.5 ‰.

Positive excursions are labeled A, B, C. Positive excursion B is the *Tenuicostatum* Zone positive excursion noted by many authors. Positive excursion C is truncated in many C-isotope profiles. The failure to define the C-isotope stratigraphy over the full stratigraphic profile shown here has led many to the erroneous interpretation that the return to backgrounds are negative isotope excursions.



482 **Completeness of the Section at Peniche and Age Model of M20.**

483 The Peniche Section is the GSSP for the Toarcian. The rules for adoption of a GSSP are set out one  
484 <https://stratigraphy.org/gssps/> (accessed on 10/09 2020). The Peniche section contravenes rules 4, 7  
485 and 8, which are :

- 486
- 487 **4. The horizon in which the marker appears should have minerals that can be**  
488 **radiometrically dated.**
- 489 **7. The outcrop has to have an adequate thickness.**
- 490 **8. Sedimentation has to be continuous without any changes in facies**  
491

492 The Pl-To boundary interval at Peniche is marked by a hardground (Rocha *et al.* 2016). The  
493 boundary shows extreme condensation and the Tenuicostatum Zone is an interval of varying  
494 sedimentation rate (McArthur *et al.* 2020a). These facts compromise the age model developed for  
495 the interval by Müller *et al.* (2020), which is based on a cyclostratigraphic analysis of Peniche that  
496 ignores these observations. To emphasise this point, we reproduce the following from McArthur *et al.*  
497 *al.* (2020), together with Fig. 4 of those authors shown here as Fig. S12.

498 At Peniche, values of  $^{87}\text{Sr}/^{86}\text{Sr}$  increase by 0.000 012 over the first 3 metres of section (Fig.  
499 3) and then remain at 0.707 085 over the next 2.5 m (to 5.5 m). Upsection from 5.5 m to 11.1 m, the  
500 rate of increase in  $^{87}\text{Sr}/^{86}\text{Sr}$  increases as level increases. It follows that the sedimentation rate  
501 through the Tenuicostatum Chronozone was lowest in the basal 3 m of the section, highest between  
502 3 and 5.5 m, where  $^{87}\text{Sr}/^{86}\text{Sr}$  remains constant, and decreased upward from 5.5 m to the base of  
503 the Serpentinum Chronozone at 11.1 m. Of the increase in  $^{87}\text{Sr}/^{86}\text{Sr}$  of 0.000 023 though this  
504 chronozone, half occurs in the basal 3 m of the section, so half of Tenuicostatum time is compressed  
505 (non-linearly) into this basal 3 m of section. Most of the rest of Tenuicostatum time is non-linearly  
506 recorded in the sediments between 5.5 and 11.1 m. These observations of varying sedimentation  
507 rates will affect any cyclostratigraphic interpretation of the section (Suan *et al.* 2008b, Huang and  
508 Hesselbo 2014, cf. Ruebsam *et al.* 2014) ...

509 That the sedimentation rate was slowest in the basal 3 m of the section is no surprise: Elmi  
510 (2007) noted that Bed 15 is a “condensed interval” and that characteristics of the 5 beds in Bed  
511 15a–15e “indicate a low sedimentation rate” and also that these beds “are capped by a hard  
512 ground (top surface of  
513 level 15e in Mouterde  
514 1955...)”. Our Sr-isotope  
515 data show that this  
516 condensation extends into  
517 the overlying 3 m of  
518 sediment with  
519 condensation intensity  
520 decreasing upwards.  
521 Condensation is confirmed  
522 by the relative thicknesses  
523 of the Mirabile and  
524 Semicelatum II  
525 Subchronozones at  
526 Peniche compared to elsewhere (Fig. 4). .....  
527

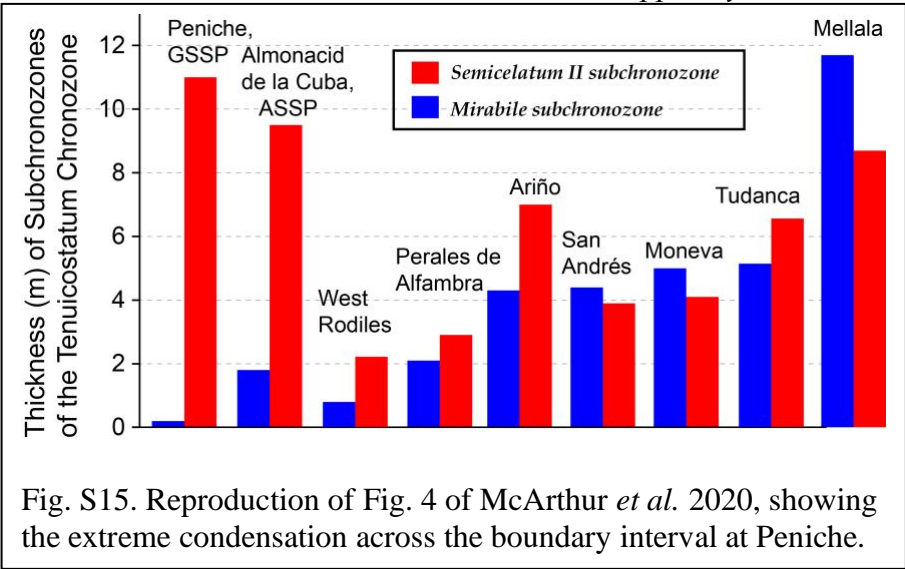


Fig. S15. Reproduction of Fig. 4 of McArthur *et al.* 2020, showing the extreme condensation across the boundary interval at Peniche.

To further illustrate the point that the age model of Muller et al. (2019) is inaccurate, we note that it is based on the cyclostratigraphic analysis of the Peniche section by Huang and Hesselbo (2014) who, in undertaking that analysis, made inadequate allowance for changes in sedimentation rate through the interval. According to Huang and Hesselbo (2014), “a duration of ~58 kyr was estimated for the negative  $\delta^{13}\text{C}_{\text{carb}}$  excursion at the Pliensbachian/Toarcian boundary from the 405-kyr tuned series (Fig. 3D), although we note ... a degree of stratigraphic condensation within about a meter on either side of the boundary (Elmi, 2006), meaning that the estimate should be considered a minimum.”.

Three problems arise with this cyclostratigraphical estimate. Firstly, the data were ‘detrended’ and this procedure introduces spurious spectral peaks in cyclostratigraphic analysis (Vaughan et al. 2015). Secondly, the minimum isn't at the boundary, but some 0.7 m above it in a highly condensed section in which stratal thickness does not equate to time. Thirdly, the negative  $\delta^{13}\text{C}_{\text{carb}}$  excursion includes the mirabile Subzone (0–0.2 m above the boundary), and some 0.68 m of the overlying, condensed, part of the semicelatum II Subzone, which is 11 m thick. The negative excursion in  $\delta^{13}\text{C}_{\text{carb}}$  therefore occupies all of mirabile time and a minimum of 0.68/11 of semicelatum II time (and probably more, given the condensation noted here and by Huang and Hesselbo, 2014). Given the relative thicknesses (and so roughly, durations) of these two subzones elsewhere than Peniche (Fig. 4 of McArthur et al. 2020) the duration of the negative excursion in  $\delta^{13}\text{C}_{\text{carb}}$  cannot be, as they state “~58 ka”. This estimate has a knock-on effect on estimates of the duration of other parts of the section which, in turn, cannot be correct.

## Additional References

- Bajnai D., Guo W., Spötl C., Coplen T.B., Methner K., Löffler N., Krsnik E., Gischler E., Hansen M., Henkel D., Price G.D., Raddatz J., Scholz D. and Fiebig J. 2020. Dual clumped isotope thermometry resolves kinetic biases in carbonate formation temperatures. *Nature Communications*, 11:4005, <https://doi.org/10.1038/s41467-020-17501-0>.
- Bajnai D., Fiebig J., Tomašových A., Garcia S.M., Rollion-Bard C., Raddatz J., Löffler N., Primo-Ramos C. and Brand U. 2018. Assessing kinetic fractionation in brachiopod calcite using clumped isotopes. *Scientific Reports*, 8:533. DOI:10.1038/s41598-017-17353-7
- Brand U. and Veizer J. 1980. Chemical diagenesis of a multicomponent carbonate system: 1. Trace elements. *J. Sediment. Petrol.* 50, 1219–1236.
- Brand U. and Veizer J. 1981. Chemical diagenesis of a multicomponent carbonate system: 2. Stable isotopes. *J. Sediment. Petrol.* 51, 987–997.
- Carpenter S.J. and Lohmann K.C. 1995.  $\delta^{18}\text{O}$  and  $\delta^{13}\text{C}$  values of modern brachiopod shells. *Geochimica et Cosmochimica Acta*, 59 (18) 3749–3764.
- Jurikova H., Liebetrau V., Gutjahr M., Rollion-Bard C., Huc M.Y., Krause S., Henkel D., Hiebenthal C., Schmidt M., Laudien J. and Eisenhauer A. 2019. Boron isotope systematics of cultured brachiopods: Response to acidification, vital effects and implications for palaeo-pH reconstruction. *Geochimica et Cosmochimica Acta* 248, 370–386.
- Penman D.E., Hönisch B., Rasbury E.T., Hemming N.G., Spero H.J. (2013). Boron, carbon, and oxygen isotopic composition of brachiopod shells: Intra-shell variability, controls, and potential as a paleo-pH recorder. *Chemical Geology*, 340, 32–39.

573 Pittet, B., Suan, G., Lenoir, F., Duarte, L. V., Mattioli, L. V., 2014. Carbon isotope evidence for  
574 sedimentary discontinuities in the lower Toarcian of the Lusitanian Basin (Portugal): Sea level  
575 change at the onset of the Oceanic Anoxic Event. *Sed. Geol.* 303, 1–14.

576 Podlaha, O.G., Mutterlose, J., and Veizer, J. 1998. Preservation of  $\delta^{18}\text{O}$  and  $\delta^{13}\text{C}$  in belemnite rostra  
577 from the Jurassic/Early Cretaceous successions. *American Journal of Science*, **298**, 324–347.

578 Saelen, G. 1989. Diagenesis and construction of the belemnite rostrum. *Palaeontology*, **34(4)**, 765–  
579 798.

580 Vaughan S., Bailey R.J. and Smith D.G. 2015. Cyclostratigraphy: data filtering as a source of  
581 spurious spectral peaks. *In* Smith D.G., Bailey R.J., Burgess P.M. & Fraser A.J. (eds) *Strata and*  
582 *Time: Probing the Gaps in Our Understanding*. Geological Society, London, Special Publication,  
583 404, 151–156.