1	Prevalence of Nosema microsporidians in commercial bumblebees (Bombus
2	terrestris) is not related to the intensity of their use at the landscape scale
3	Alejandro Trillo <sup>1</sup> , Mark J. F. Brown <sup>2</sup> and Montserrat Vilà <sup>1</sup>
4	Estación Biológica de Doñana (EBD-CSIC), Avda. Américo Vespucio 26, Isla de la
5	Cartuja, E-41092, Sevilla, Spain
6	<sup>2</sup> School of Biological Sciences, Royal Holloway University of London, Egham TW20
7	0EX, UK
8	
9	Corresponding author: A. Trillo
10	atrilloig@gmail.com
11	

**Abstract** – The use of commercial bumblebees to aid crop pollination may result in overcrowding of agricultural landscapes by pollinators. Consequently, transmission of parasites between pollinators via shared flowers may be substantial. Here we assessed the initial infection status of commercial Bombus terrestris colonies, and then explored spatial and seasonal influences on changes in parasite prevalence across a landscape where bumblebee colonies are intensively used to pollinate berry crops in SW Spain. Colonies were placed inside strawberry greenhouse crops and in woodlands adjacent and distant to crops in winter and in spring, as representative periods of high and low use of colonies, respectively. Worker bumblebees were collected from colonies upon arrival from a producer and 30 days after being placed in the field. The abdomen of each bumblebee was morphologically inspected for a range of internal parasites. Upon arrival 71% of the colonies were infected by spores of *Nosema*. Three bumblebees from two colonies harbored A. bombi spores at the end of their placement in woodlands adjacent to crops. Nosema colony prevalence did not change significantly either among sites or between seasons. We found no evidence for the density of commercial B. terrestris impacting Nosema epidemiology in those commercial colonies, but our results highlight the potential risk for parasites to be transmitted from commercial bumblebees to native pollinators.

12

13

14

15

16

17

18

19

20

21

22

23

24

25

26

27

28

29

30

agricultural landscape / Apicystis bombi / Fragaria × ananassa / parasite

#### 1. INTRODUCTION

31

32

33

34

35

36

37

38

39

40

41

42

43

44

45

46

47

48

49

50

51

52

53

In the last half century there has been an increase in the expansion of pollinatordependent crops (Aizen et al., 2008) that has required a parallel demand for commercially produced bees (Potts et al., 2016). Bumblebees (*Bombus* sp.) started to be commercially produced in Europe in the late 1980s, to replace the costly mechanicalpollination of tomatoes (Solanum lycopersicum) (Ravestijn and Sande, 1991; Velthuis and van Doorn, 2006). Quickly, bumblebee breeding techniques advanced and colonies were mass-produced and transported worldwide, where they currently pollinate over 20 different pollinator-dependent crops. Over two million bumblebee colonies are produced annually (Graystock, Blane, et al., 2016). The use of commercial pollinators such as bumblebees to aid crop pollination is not free of environmental risks. For instance, queens of commercial bumblebees have become established in many parts of the world (Matsumura et al., 2004; Morales et al., 2013), and there is empirical evidence showing competition for nest sites with other native bumblebee queens in the lab (Ono, 1997) and in the field (Inoue et al., 2008). In addition, commercial bumblebees may compete for food with other native pollinators (Matsumura et al., 2004; Morales et al., 2013), as well as promote the spread of parasites via shared flowers (Colla et al., 2006; Meeus et al., 2011; Schmid-Hempel et al., 2014). Several bee parasite species have been found in commercial bumblebee colonies. In 1999, Goka et al. (2000) found for the first time the presence of a parasite, Locustacarus buchneri, in commercially produced Bombus terrestris colonies upon arrival in Japan from an overseas supplier. The presence of this parasite has been linked to shorter

lifespan (Otterstatter and Whidden, 2004) and changes in behavior of bumblebees (Otterstatter et al., 2005). Further studies have reported that commercial bumblebee colonies frequently have a range of bumblebee parasites (Graystock et al., 2013a; Murray et al., 2013) and even honeybee parasites (Graystock et al., 2013a), with the latter probably via the consumption of honeybee pollen by reared bumblebees (Goulson and Hughes, 2015). Importantly, the use of commercial pollinators in crops produces high densities of pollinators not only in the agricultural fields or greenhouses, but in adjacent natural areas as well (Ishii et al., 2008; González-Varo and Vilà, 2017; Trillo et al., 2018). Presumably, in those areas, the rate of parasite transmission among pollinators will rise, because high densities of hosts provide ideal conditions for the spread of parasites (Arneberg et al., 1998). In fact, several studies have shown, through the collection of free-flying bumblebees, high prevalence of parasites in sites adjacent to greenhouses where commercial bumblebees are used compared with sites distant to those greenhouses (Colla et al., 2006; Murray et al., 2013) or in greenhouses absent of such commercial bumblebees (Graystock et al., 2014), although there is also evidence against this (Whitehorn et al., 2013). To partially reduce the impact of commercial bumblebees on native pollinator populations and because healthy bumblebees may perform better, as is seen with honeybees (Geslin et al., 2017), producers are under pressure to produce parasite-free bumblebee colonies. In this study, we first examined whether commercially produced B. terrestris colonies, used to pollinate berry crops in Huelva (SW Spain), carried parasites upon arrival from a producer. We morphologically searched for five common internal bee parasites: larvae of the family Conopidae and Braconidae, L. buchneri, Apicystis bombi and parasites of the genus Nosema, which all potentially affect bumblebee health.

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

70

71

72

73

74

75

76

For instance, larvae of parasitic flies lead to bee death (Schmid-Hempel and Schmid-Hempel, 1996), L. buchneri likely reduces lifespan of individual infected host (Otterstatter and Whidden, 2004), A. bombi is linked to deterioration of the fat body (Graystock, Meeus, et al., 2016; Macfarlane et al., 1995) and most *Nosema* species reduce worker survival and colony size (Otti and Schmid-Hempel, 2007; Rutrecht and Brown, 2009; Graystock et al., 2013a). We then experimentally tested spatial and seasonal influences on changes in the prevalence of these parasites across a landscape where bumblebee colonies are intensively used. Importantly, L. buchneri, A. bombi and Nosema are likely to be transmitted among pollinators via shared flowers (Durrer and Schmid-Hempel, 1994; Goka et al., 2006; Graystock et al., 2015). Colonies were placed inside strawberry crops and in woodlands adjacent and distant to those crops in January (winter) and again in April (spring), as representative periods of high and low use of bumblebee colonies in berry crops, respectively. We expected parasite prevalence to be highest with high densities of commercial bumblebees in the landscape, that is, 1) higher levels of prevalence at sites inside and adjacent to greenhouse crops than distant, and 2) higher levels in winter than in spring because of the greater use of colonies in winter.

78

79

80

81

82

83

84

85

86

87

88

89

90

91

92

93

#### 2. MATERIALS AND METHODS

# 2.1. Study system

The study was conducted in the Guadalquivir Valley in the province of Huelva (SW Spain). In this region there are large intensively cultivated areas of berries (9,500 ha), especially strawberries (~70% of the total area devoted to berry crops) (Freshuelva, 2015). Strawberries are cultivated in semi-open polytunnel greenhouses with open sides from November to May. In order to aid crop pollination, farmers use commercial bumblebees (*Bombus terrestris*). Notably, bumblebee colonies are especially used at the beginning of the flowering period (i.e. in winter; personal observations) due to major revenues and worse weather conditions than in spring. The most common remaining natural habitat patches across berry crops are woodlands composed of a rich flora of entomophilous Mediterranean shrubs and herbs, which provide flowers throughout the strawberry cultivation period (Herrera, 1988).

# 2.2. Experimental design

In 2015, we purchased 48 *B. t. terrestris* colonies from Koppert Biological Systems, one of the main producers in Europe and specifically in this region. Each colony consisted of a plastic box within a cardboard container, with syrup solution provided *ad libitum*.

Each colony included a queen and ~100 workers.

First, to quantify colony parasite prevalence, at the arrival of the colonies (period 'before'), we collected 10 workers from each colony. Each worker was frozen in an individual clean vial at -20 °C for later analyses.

Second, to investigate changes in colony parasite prevalence across the landscape, we placed two colonies each in four strawberry crops ('inside') and in eight woodlands, four adjacent to the selected strawberry crops (~50 m; 'adjacent') and four without berry crops in the surrounding 2 km radius landscape ('distant') (Fig. 1). We chose a 2 km buffer radius because most bumblebee foraging flights do not exceed this distance (Osborne et al., 2008). The surrounding landscape for inside and adjacent plots had a high berry crop cover (overall mean  $\pm$  SE = 48  $\pm$  5.6%; see Table S1). Inside/adjacent plots and distant plots are representative of contrasting landscapes in terms of commercial bumblebee colony density. The density is high and absent in those landscapes, respectively. In fact, commercial bumblebees are frequently observed in landscapes with berry crop cover, rather than when berry crop cover is absent in the landscapes (Trillo et al., 2019). The average (± SE) distance between adjacent and distant woodland plots was  $5903 \pm 1038$  m (range = 3.1-11.4 km). This distance meets independence criteria to avoid spatial pseudoreplication between non-paired plots. Third, to investigate seasonal change effects, the experiment was conducted in January ('winter') and repeated in April ('spring'), as representative periods of high and low use of bumblebee colonies in strawberry crops, respectively. Here, the climate is typically Mediterranean with mild winters and warm springs (AEMET, 2015). These two seasons also differ in wild floral resources. The flowering peak is in spring when the floral richness and density are almost triple that in winter (Trillo et al., 2019). Wild pollinator species occur as flowering plant species thrive (Herrera, 1988). In each season, we placed two colonies of bumblebees in the center of each plot. Bumblebees were allowed to forage for 30 days. In strawberry crops the two colonies were hung between four separate greenhouses. The distance between the two colonies

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

within a plot was  $\sim$ 20 m. In woodland plots the two colonies were hidden in wooden boxes to avoid predation. At the end of the experiment, we collected 10 bumblebee workers per colony returning to it (period 'after') over two days using aerial nets. Bumblebees were kept in individual clean vials with ice until arrival at the lab where they were stored at -20 °C.

# 2.3. Parasite screening

The abdomen of each bumblebee was dissected and inspected under a magnifying lens for larvae of parasitic conopid flies (Conopidae, Diptera) and braconid wasps of the genus *Syntretus* (Braconidae, Hymenoptera), and the air sacs were specifically inspected for the tracheal mite *Locustacarus buchneri* (Podapolipidae) (Yoneda et al., 2008).

Then, a piece (0.2 cm × 0.2 cm, approx.) of the fat body was dissected out from each bumblebee and mounted on a slide (note that the gut was not included for these analyses). By screening only the fat body, we were able to confirm that we were detecting true infections, not just passage through the gut by vectored spores. We completely screened each slide at ×400 magnification for the presence of spores of the neogregarine *Apicystis bombi* (Lipotrophidae) and microsporidians of the genus *Nosema* (Nosematidae). We estimated parasite prevalence (presence or absence) instead of individual infection levels (abundance) because the latter is influenced by many confounding factors that drive infection intensity (Rutrecht and Brown, 2009).

#### 2.4. Statistical analyses

Only *Nosema* infections (Table S2) were statistically analysed, because the remaining parasites showed no or very low prevalence in the colonies (see results). *Nosema* prevalence was calculated estimating the percentage of bumblebees infected taking into

account the 10 individuals collected per colony. A linear mixed model (LMM; Gaussian error distribution based on homogeneity in the residuals) was used to analyse whether changes in *Nosema* prevalence were related to our experimental setting. The difference in *Nosema* prevalence in the colonies before and after being placed in the field was used as the response variable. Season (winter/spring), plot type (inside, adjacent and distant), and their interaction were included as fixed factors in the model, while study plot was included as a random factor to account for the paired design between inside and adjacent plots and the re-sampled plots in winter and in spring (see Table S3 for the R code). All statistical analyses were conducted in R (v.3.1.3, R Core Team, 2014). We used the package *lmerTest* (Kuznetsova et al., 2013) for the LMM and Satterthwaite's approximations for F- and p- values.

#### 3. RESULTS

175

176

177

178

179

180

181

182

183

184

185

186

187

188

189

190

191

192

193

In total, over the two seasons we screened 919 bumblebee workers. We missed one colony and several individuals from other colonies due to low colony activity. On average ( $\pm$  SE) we collected 19.55  $\pm$  0.13 (range = 16–20) bumblebees per colony. None of the bumblebees were infected by larvae of parasitic conopid flies (Conopidae, Diptera), braconid wasps of the genus Syntretus (Braconidae, Hymenoptera), or the tracheal mite, Locustacarus buchneri. The prevalence of Apicystis bombi was extremely low; only three bumblebees harbored spores in their fat body, and these were collected from two colonies at the end of their placement in adjacent woodlands. In contrast, spores of *Nosema* were found in 58.3% (14 out of 24 colonies) of colonies in winter and in 83.3% (20 out of 24) in spring at the start of each experimental block, that is, upon arrival from the producer prior to their placement in the field. The average *Nosema* prevalence per colony in the before period was  $14.0 \pm 3.4\%$  (mean  $\pm$  SE, hereafter) in winter, and  $19.7 \pm 3.2\%$  in spring. The average *Nosema* prevalence in the after period was  $10.2 \pm 2.3\%$  in winter and  $26.4 \pm 6.6\%$  in spring. Neither the season (F<sub>1</sub>)  $_{35} = 2.88$ , p < 0.10) nor the distance ( $F_{2,19} = 0.25$ , p < 0.79) or their interaction ( $F_{2,35} = 0.50$ , p < 0.61) had a significant effect on changes in *Nosema* colony prevalence between periods (Fig. 2A and 2B).

#### 4. DISCUSSION

194

195

196

197

198

199

200

201

202

203

204

205

206

207

208

209

210

211

212

213

214

215

216

217

The use of commercial bumblebees has been linked to the decline of several native pollinator species (Cameron et al., 2011; Morales et al., 2013; Schmid-Hempel et al., 2014). Among the mechanisms behind this decline, parasite spillover from commercial to native pollinator populations may play a substantial role (Meeus et al., 2011). Mass commercial breeding programs may facilitate the probability of parasite transmission among hosts, as companies usually handle high densities of bumblebees in their facilities. In parallel, the provision of ad libitum food may facilitate the reproduction of infected hosts (Brown et al., 2000). Furthermore, even in the case that commercial bumblebees are parasite-free, they may act as reservoirs for parasites in the field, through a spill-back mechanism, leading to an increase in parasite prevalence (Stout and Morales, 2009; Meeus et al., 2011). Upon arrival, we found no evidence for the presence of larvae of parasitic conopid flies (Conopidae, Diptera) and braconid wasps of the genus Syntretus (Braconidae, Hymenoptera), or the tracheal mite, Locustacarus buchneri, in the screened Bombus terrestris colonies. Although the presence of larvae of parasitic insects has never been reported in commercial bumblebees, the tracheal mite, L. buchneri, was highly prevalent at the end of the 20th century (Goka et al., 2000) spilling over to native bumblebees (Goka et al., 2006). However, it seems that producers have largely eliminated this parasite from commercial bumblebee colonies (Goka et al., 2006; Murray et al., 2013; although see Sachman-Ruiz et al., 2015). In addition, neither these parasitoids nor the tracheal mite, L. buchneri, were observed in bumblebees from the colonies after being placed in the field for a month. One explanation for this is that parasitoids of bumblebees might be at low abundance in our study sites, because native

bumblebees (B. terrestris lusitanicus) are rare (Magrach et al., 2017; Trillo et al., 2019), as they are at the limit of their distributional range (Goulson, 2010). In fact, in this region, the density of commercial bumblebees is around four times greater than that of native bumblebees (Trillo et al., 2019). Another possible and complementary explanation for this low prevalence might be that when bumblebees are parasitized, they desert their colony (Schmid-Hempel and Müller, 1991). In addition, even though L. buchneri may be present in native bumblebees (although we note that there is no information in Spain; Jabal-Uriel et al. 2017) it might be very difficult to detect parasite spillover from native to managed bumblebees because native bumblebees are at very low abundance, as described above. Similarly, there was no evidence for the presence of the neogregarine *Apicystis bombi* (Lipotrophidae) in the screened colonies upon arrival. However, three bumblebees were found to be infected after having been placed in the field. In other regions, the parasite A. bombi has been detected infecting commercial bumblebee colonies, although in a low number of colonies (Graystock et al., 2013b; Murray et al., 2013; although again see Sachman-Ruiz et al., 2015). Native bumblebees can host A. bombi (Jabal-Uriel et al., 2017), but, as noted above, they are rare in our study region (Magrach et al., 2017; Trillo et al., 2019). In contrast, thousands of commercial colonies from at least three producers (Koppert, Biobest and Agrobio, personal observation) are used on an annual basis. Therefore, it is more likely that other commercial bumblebees infected by A. bombi transmitted the parasite to the bumblebee colonies we screened, rather than native bumblebees, or, more parsimoniously, our initial screen failed to detect it in arriving colonies.

218

219

220

221

222

223

224

225

226

227

228

229

230

231

232

233

234

235

236

237

238

239

In contrast, we found commercially produced bumblebee colonies to be heavily infected with parasites of the genus *Nosema* upon arrival from the producer. Other studies have also reported similar levels of prevalence with around three quarters of commercial colonies infected (Graystock et al., 2013a; Murray et al., 2013). Unfortunately, our methodology did not allow us to distinguish between the bumblebee parasite N. bombi and the honeybee parasite N. ceranae. Both can infect bumblebees (Graystock et al., 2013a; Fürst et al., 2014). Unexpectedly, our results showed no significant variation in *Nosema* infection rate at a colony level over time, as in a previous study that monitored wild bumblebees (Goulson et al., 2018), even in landscapes where commercial bumblebees were intensively used to pollinate crops. Even in parasite-free landscapes, one would expect that if commercial colonies are infected by a parasite, it spreads within the colony across time due to the high density of hosts and low genetic variability (Schmid-Hempel, 1998). We propose two potential explanations. On the one hand, bumblebees, in line with other social insects, have evolved social immune systems that combine prophylactic and activated responses to avoid, control or eliminate parasite infections (reviewed by Cremer et al., 2007). Both colony and individual (i.e. immunocompetence, reviewed by Schmid-Hempel 2005) defense mechanisms might be involved in maintaining roughly constant *Nosema* prevalence over time. On the other hand, it has been experimentally demonstrated that *Nosema*, specifically *N. bombi*, relies more on transmission through the larval stage than through transmission among adults (Rutrecht et al., 2007). If we consider that colonies were placed in the field for a month period and that the total development of a bumblebee from larvae to adult is about 4-5 weeks (Alford, 1975),

241

242

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

this could explain why we failed to detect an increase in prevalence. Imhoof and

Schmid-Hempel (1999) showed an average delay to *Nosema* infection in commercial colonies placed in the field of ~30 days.

Our study showed, for the first time in Spain, that commercially produced bumblebee colonies can be infected by *Nosema* parasites prior to their deployment in the field.

These parasites may reduce lifespan and have detrimental effects on bumblebee behavior (Otti and Schmid-Hempel, 2007; Rutrecht and Brown, 2009; Graystock et al., 2013a). Because commercial bumblebees placed in semi-open greenhouses frequently forage in natural areas (Foulis and Goulson, 2014), they have the potential to spread the parasites into native pollinator populations (Colla et al., 2006; Murray et al., 2013).

Despite the fact that there is some regulation about commercial bee colony health (e.g. for Europe see 92/65/EEC in European Commission 1992), this regulation does not cover all parasites. This implies that commercial colonies can be highly infected by parasites such as *Nosema*, as our study show. Therefore, there is a need for the enforcement of more stringent protocols to preserve the health of commercial and native pollinators.

#### **ACKNOWLEDGEMENTS**

We thank J. Angelidou, C. Apostolidou, A. Montero-Castaño, D. Ragel and E. Tsiripli for field assistance and the farmers for letting us work on their lands. We thank J. Bagi and E.J. Bailes for lab support. We would also like to thank the editor and the anonymous reviewers for comments that significantly improved the manuscript. Funding was provided by the Spanish Ministry of Economy and Competitiveness project FLORMAS ('Influence of mass flowering crops on pollinator biodiversity, project no CGL2012-33801') and by the Biodiversa-FACCE project ECODEAL

('Enhancing biodiversity-based ecosystem services to crops through optimised densities 288 of green infrastructure in agricultural landscapes, project no PCIN-2014-048'). AT was 289 supported by a Severo-Ochoa predoctoral fellowship (SVP-2013-067592) and by a 290 291 Short Term Scientific Mission from the COST Action (FA1307:35075) for international 292 mobility. **AUTHOR CONTRIBUTIONS** 293 AT and MV conceived this research and designed experiments; MJFB participated in 294 295 the design and interpretation of the data; AT performed experiments, analyses and wrote 296 the first draft of the manuscript; MJFB and MV edited and contributed to the writing of the manuscript. All authors read and approved the final manuscript. 297 COMPLIANCE WITH ETHICAL STANDARDS 298 Conflict of interest. The authors declare that they have no potential conflict of interest 299 300 in relation to the study in this paper. 301 302 REFERENCES AEMET (2015). Valores climatológicos normales. Huelva, Ronda Este. 303 http://www.aemet.es/es/ (Accessed 01 September 2015). 304 Aizen, M. A., Garibaldi, L. A., Cunningham, S. A., Klein, A. M. (2008). Long-term 305 306 global trends in crop yield and production reveal no current pollination shortage but increasing pollinator dependency. Curr. Biol. 18 (20), 1572–1575. 307

Alford, D. V. (1975). Bumblebees. Davis-Poynter, London.

- 309 Arneberg, P., Skorping, A., Grenfell, B., Read, A. F. (1998). Host densities as 310 determinants of abundance in parasite communities. Proc. R. Soc. B Biol. Sci. 265 (1403), 1283–1289. 311 Brown, M. J. F., Loosli, R., Schmid-Hempel, P. (2000). Condition-dependent 312 expression of virulence in a trypanosome infecting bumblebees. Oikos 91 (3), 421– 313 427. 314 Cameron, S. A., Lozier, J. D., Strange, J. P., Koch, J. B., Cordes, N., Solter, L. F., 315 316 Griswold, T. L. (2011). Patterns of widespread decline in North American bumble 317 bees. Proc. Natl. Acad. Sci. U. S. A. 108 (2), 662–667. 318 Colla, S. R., Otterstatter, M. C., Gegear, R. J., Thomson, J. D. (2006). Plight of the bumble bee: pathogen spillover from commercial to wild populations. Biol. 319 320 Conserv. 129 (4), 461–467. 321 Cremer, S., Armitage, S. A. O., Schmid-Hempel, P. (2007). Social immunity. Curr. 322 Biol. **17** (16), 693–702. 323
- Durrer, S., Schmid-Hempel, P. (1994). Shared use of flowers leads to horizontal pathogen transmission. Proc. R. Soc. B Biol. Sci. **258** (1353), 299–302.
- European Commission (1992). Council Directive 92/65/EEC. No. L 268/54.
- Foulis, E. S. J., Goulson, D. (2014). Commercial bumble bees on soft fruit farms collect pollen mainly from wildflowers rather than the target crops. J. Apic. Res. **53** (3), 404–407.
- Freshuelva (2015). http://www.freshuelva.es/ (Accessed 13 November 2015).

- Fürst, M. A., McMahon, D. P., Osborne, J. L., Paxton, R. J., Brown, M. J. F. (2014).
- Disease associations between honeybees and bumblebees as a threat to wild
- pollinators. Nature **506** (7488), 364–366.
- Geslin, B., Aizen, M. A., Garcia, N., Pereira, A.-J., Vaissière, B. E., Garibaldi, L. A.
- 334 (2017). The impact of honey bee colony quality on crop yield and farmers' profit
- in apples and pears. Agric. Ecosyst. Environ. **248**, 153–161.
- Goka, K., Okabe, K., Yoneda, M. (2006). Worldwide migration of parasitic mites as a
- result of bumblebee commercialization. Popul. Ecol. **48** (4), 285–291.
- Goka, K., Okabe, K., Niwa, S., Yoneda, M. (2000). Parasitic mite infestation in
- introduced colonies of European bumblebees, *Bombus terrestris*. Japanese J. Appl.
- 340 Entomol. Zool. **44** (1), 47–50.
- González-Varo, J. P., Vilà, M. (2017). Spillover of managed honeybees from mass-
- flowering crops into natural habitats. Biol. Conserv. **212**, 376–382.
- Goulson, D. (2010). Bumblebees: behaviour, ecology, and conservation. Second ed.
- Oxford University Press, Oxford, UK.
- Goulson, D., Hughes, W. O. H. (2015). Mitigating the anthropogenic spread of bee
- parasites to protect wild pollinators. Biol. Conserv. **191**, 10–19.
- Goulson, D., O'Connor, S., Park, K. J. (2018). The impacts of predators and parasites
- on wild bumblebee colonies. Ecol. Entomol. **43** (2), 168–181.
- Graystock, P., Goulson, D., Hughes, W. O. H. (2014). The relationship between
- managed bees and the prevalence of parasites in bumblebees. PeerJ 2, e522.

- Graystock, P., Goulson, D., Hughes, W. O. H. (2015). Parasites in bloom: flowers aid
- dispersal and transmission of pollinator parasites within and between bee species.
- 353 Proc. R. Soc. B Biol. Sci. **282** (1813), 20151371.
- Graystock, P., Yates, K., Darvill, B., Goulson, D., Hughes, W. O. H. (2013a). Emerging
- dangers: deadly effects of an emergent parasite in a new pollinator host. J.
- 356 Invertebr. Pathol. **114** (2), 114–119.
- Graystock, P., Blane, E. J., McFrederick, Q. S., Goulson, D., Hughes, W. O. H. (2016).
- Do managed bees drive parasite spread and emergence in wild bees? Int. J.
- 359 Parasitol. Parasites Wildl. **5** (1), 64–75.
- Graystock, P., Meeus, I., Smagghes, G., Goulson, D., Hughes, W. O. H. (2016). The
- effects of single and mixed infections of *Apicystis bombi* and deformed wing virus
- in *Bombus terrestris*. Parasitology, **143** (3), 358–365.
- Graystock, P., Yates, K., Evison, S. E. F., Darvill, B., Goulson, D., Hughes, W. O. H.
- 364 (2013b). The Trojan hives: pollinator pathogens, imported and distributed in
- bumblebee colonies. J. Appl. Ecol. **50** (5), 1207–1215.
- Herrera, J. (1988). Pollination relationships in southern Spanish Mediterranean
- shrublands. J. Ecol. **76** (1), 274–287.
- Imhoof, B., Schmid-Hempel, P. (1999). Colony success of the bumble bee, *Bombus*
- 369 terrestris, in relation to infections by two protozoan parasites, Crithidia bombi and
- 370 *Nosema bombi*. Insectes Soc. **46** (3), 233–238.
- Inoue, M. N., Yokoyama, J., Washitani, I. (2008). Displacement of Japanese native
- bumblebees by the recently introduced *Bombus terrestris* (L.) (Hymenoptera:

- 373 Apidae). J. Insect Conserv. **12** (2), 135–146.
- Ishii, H. S., Kadoya, T., Kikuchi, R., Suda, S. I., Washitani, I. (2008). Habitat and
- flower resource partitioning by an exotic and three native bumble bees in central
- 376 Hokkaido, Japan. Biol. Conserv. **141** (10), 2597–2607.
- Jabal-Uriel, C., Martín-Hernández, R., Ornosa, C., Higes, M., Berriatúa, E., De la Rúa,
- P. (2017). First data on the prevalence and distribution of pathogens in bumblebees
- 379 (Bombus terrestris and Bombus pascuorum) from Spain. Spanish J. Agric. Res. 15
- 380 (1), 1–6.
- Kuznetsova, A., Brockhoff, P. B., Christensen, R. H. B. (2013). lmerTest: Test for
- random and fixed effects for linear mixed effect models (lmer objects of lme4
- package). R package version 2.0-30.
- Macfarlane, R. P., Lipa, J. J., Liu, H. J. (1995). Bumble bee pathogens and internal
- enemies. Bee World, **76** (3), 130–148.
- Magrach, A., González-Varo, J. P., Boiffier, M., Vilà, M., Bartomeus, I. (2017).
- Honeybee spillover reshuffles pollinator diets and affects plant reproductive
- 388 success. Nat. Ecol. Evol. **1** (9), 1299–1307.
- Matsumura, C., Yokoyama, J., Washitani, I. (2004). Invasion status and potential
- ecological impacts of an invasive alien bumblebee, *Bombus terrestris* L.
- 391 (Hymenoptera: Apidae) naturalized in Southern Hokkaido, Japan. Glob. Environ.
- 392 Res. **8** (1), 51–66.
- Meeus, I., Brown, M. J. F., De Graaf, D. C., Smagghe, G. (2011). Effects of invasive
- parasites on bumble bee declines. Conserv. Biol. **25** (4), 662–671.

- Morales, C. L., Arbetman, M. P., Cameron, S. A., Aizen, M. A. (2013). Rapid
- ecological replacement of a native bumble bee by invasive species. Front. Ecol.
- 397 Environ. **11** (10), 529–534.
- Murray, T. E., Coffey, M. F., Kehoe, E., Horgan, F. G. (2013). Pathogen prevalence in
- commercially reared bumble bees and evidence of spillover in conspecific
- 400 populations. Biol. Conserv. **159**, 269–276.
- Ono, M. (1997). Ecological implications of introduced *Bombus terrestris*, and
- significance of domestication of Japanese native bumblebees (*Bombus* spp.). Proc.
- Int. Work. Biol. Invasions Ecosyst. by Pests Benef. Org. NIAES, Minist. Agric.
- 404 For. Fish. Japan, Tsukuba, pp. 244–252.
- Osborne, J. L., Martin, A. P., Carreck, N. L., Swain, J. L., Knight, M. E., Goulson, D.,
- Hale, R. J., Sanderson, R. A. (2008). Bumblebee flight distances in relation to the
- forage landscape. J. Anim. Ecol. **77** (2), 406–415.
- Otterstatter, M. C., Whidden, T. L. (2004). Patterns of parasitism by tracheal mites
- (Locustacarus buchneri) in natural bumble bee populations. Apidologie **35** (4),
- 410 351–357.
- Otterstatter, M. C., Gegear, R. J., Colla, S. R., Thomson, J. D. (2005). Effects of
- parasitic mites and protozoa on the flower constancy and foraging rate of bumble
- bees. Behav. Ecol. Sociobiol. **58** (4), 383–389.
- Otti, O., Schmid-Hempel, P. (2007). *Nosema bombi*: a pollinator parasite with
- detrimental fitness effects. J. Invertebr. Pathol. **96** (2), 118–124.
- Potts, S. G., Imperatriz-Fonseca, V., Ngo, H. T., Aizen, M. A., Biesmeijer, J. C., et al.

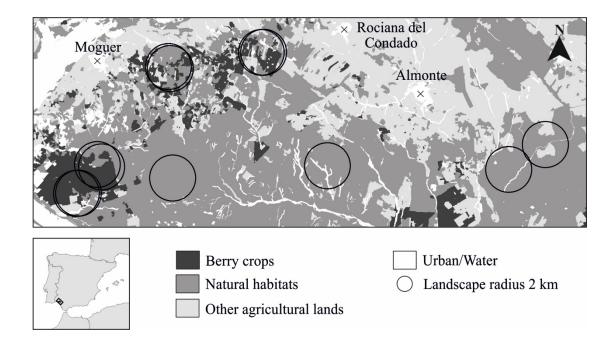
- 417 (2016). Safeguarding pollinators and their values to human well-being. Nature, **540**
- 418 (7632), 220–229.
- R Core Team. (2014). R: A language and environment for statistical computing.
- Vienna, Austria: R Foundation for Statistical Computing.
- van Ravestijn, W., van der Sande, J. (1991). Use of bumblebees for the pollination of
- glasshouse tomatoes. Acta Hortic. **288**, 204–212.
- Rutrecht, S. T., Brown, M. J. F. (2009). Differential virulence in a multiple-host parasite
- of bumble bees: resolving the paradox of parasite survival? Oikos 118 (6), 941–
- 425 949.
- Rutrecht, S. T., Klee, J., Brown, M. J. F. (2007). Horizontal transmission success of
- Nosema bombi to its adult bumble bee hosts: effects of dosage, spore source and
- 428 host age. Parasitology **134** (12), 1719–1726.
- Sachman-Ruiz, B., Narváez-Padilla, V., Reynaud, E. (2015). Commercial *Bombus*
- *impatiens* as reservoirs of emerging infectious diseases in central México. Biol.
- 431 Invasions **17** (7), 2043–2053.
- Schmid-Hempel, P. (1998). Parasites in social insects. Princeton University Press.
- Schmid-Hempel, P. (2005). Evolutionary ecology of insect immune defenses. Annu.
- 434 Rev. Entomol. **50**, 529–551.
- Schmid-Hempel, R., Müller, C. B. (1991). Do parasitized bumblebees forage for their
- 436 colony? Anim. Behav. **41** (5), 910–912.
- Schmid-Hempel, R., Schmid-Hempel, P. (1996). Larval development of two parasitic

- flies (Conopidae) in the common host *Bombus pascuorum*. Ecol. Entomol. **21** (1),
- 439 63–70.
- Schmid-Hempel, R., Eckhardt, M., Goulson, D., Heinzmann, D., Lange, C., Plischuk,
- S., Escudero, L. R., Salathé, R., Scriven, J. J., Schmid-Hempel, P. (2014). The
- invasion of southern South America by imported bumblebees and associated
- parasites. J. Anim. Ecol. **83** (4), 823–837.
- Stout, J. C., Morales, C. L. (2009). Ecological impacts of invasive alien species on bees.
- 445 Apidologie **40** (3), 388–409.
- Trillo, A., Herrera, J. M., Vilà, M. (2018). Managed bumble bees increase flower
- visitation but not fruit weight in polytunnel strawberry crops. Basic Appl. Ecol.,
- **30**, 32–40.
- Trillo, A., Montero-Castaño, A., González-Varo, J. P., González-Moreno, P., Ortiz-
- Sánchez, F. J., Vilà, M. (2019). Contrasting occurrence patterns of managed and
- 451 native bumblebees in natural habitats across a greenhouse landscape gradient.
- 452 Agric. Ecosyst. Environ., **272**, 230–236.
- Velthuis, H. H. W., van Doorn, A. (2006). A century of advances in bumblebee
- domestication and the economic and environmental aspects of its
- commercialization for pollination. Apidologie, **37**(4), 421–451.
- Whitehorn, P. R., Tinsley, M. C., Brown, M. J. F., Goulson, D. (2013). Investigating the
- impact of deploying commercial *Bombus terrestris* for crop pollination on
- pathogen dynamics in wild bumble bees. J. Apic. Res. **52** (3), 149–157.
- 459 Yoneda, M., Furuta, H., Kanbe, Y., Tsuchida, K., Okabe, K., Goka, K. (2008).

Commercial colonies of *Bombus terrestris* (Hymenoptera: Apidae) are reservoirs of the tracheal mite *Locustacarus buchneri* (Acari: Podapolipidae). Appl. Entomol. Zool. **43** (1), 73–76.

464	FIGURES CAPTIONS
465	Fig. 1 Geographical distribution of plots inside, adjacent and distant to berry crops
466	located in the province of Huelva (SW Spain). Names denote towns.
467	Fig. 2 Mean (+SE) change in <i>Nosema</i> prevalence in commercially produced bumblebee
468	colonies before and after being placed in plots inside, adjacent (~50 m) and distant (>2
469	km) to berry crops in winter (A) and in spring (B). Differences were not significant
470	

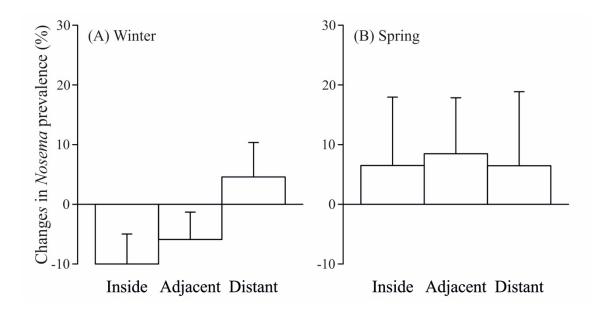
# 471 Fig. 1



**Fig. 1** Geographical distribution of plots inside, adjacent and distant to berry crops located in the province of Huelva (SW Spain). In total, 12 plots were selected and two commercial bumblebee colonies were used per plot in winter and again in spring.

Names and crosses denote towns.

# Fig. 2



**Fig. 2** Mean (+SE) change in *Nosema* prevalence in commercially produced bumblebee colonies before and after being placed in plots inside, adjacent (~50 m) and distant (>2 km) to berry crops in winter (A) and in spring (B). Differences were not significant