- 1 Earthworms affect plant growth and resistance against herbivores: a
- 2 meta-analysis

3

- 4 Zhenggao Xiao^{1,2}, Xie Wang³, Julia Koricheva⁴, Alan Kergunteuil², Renée-Claire Le
- 5 Bayon², Mangiang Liu^{1,5*}, Feng Hu^{1,5} and Sergio Rasmann²

6

- ¹Soil ecology lab, College of Resources and Environmental Sciences, Nanjing
- 8 Agricultural University, Nanjing 210095, China
- ⁹ Functional ecology lab, Institute of Biology, University of Neuchâtel, Neuchatel 2000,
- 10 Switzerland
- ³Soil and Fertilizer Research Institute, Sichuan Academy of Agricultural Sciences,
- 12 Chengdu 610066, China
- ⁴School of Biological Sciences, Royal Holloway University of London, Egham, Surrey,
- 14 TW20 0EX, UK
- ⁵Jiangsu Collaborative Innovation Center for Solid Organic Waste Resource
- 16 Utilization, Nanjing 210014, China

17

- *Corresponding author:
- 19 Email: liumq@njau.edu.cn or manqiang-liu@163.com
- 20 Tel.: +86 25 84395104; Fax: +86 25 84395210

21

22 Running headline: Earthworm effects on plant resistance

Summary

23

24 1. Subterranean detritivores such as earthworms can increase soil nutrient availability 25 through their burrowing and casting activities. A number of recent studies have 26 explored whether these changes caused by earthworms may in turn affect plant 27 performance and resistance to herbivores, but no formal synthesis of this literature has 28 been conducted to date. 29 **2.** We here formally tested for the effects of earthworms on plant growth, resistance 30 and chemical defence against insect herbivores by performing a meta-analysis of the 31 existing literature up to 2016. We also explored ecological factors that might explain 32 among-studies variation in the magnitude of the earthworm effects on plant growth 33 and resistance. 34 **3.** We found that earthworm presence increases plant growth (by 20 %) and nitrogen 35 content (by 11 %). Overall, earthworms did not affect plant resistance against 36 chewing herbivores (caterpillars, slugs and rootworms), and even led to a 22 % 37 decrease in plant resistance against phloem-feeding herbivores (aphids). However, 38 earthworm presence increased production of chemical defences by 31% when plants 39 where attacked by cell-feeders (thrips), and resulted in an 81 % increase in resistance 40 against thrips. The magnitude of earthworm effects was stronger when earthworm 41 inoculations consisted of a mix of species and ecological types, and when densities of 42 earthworms were high. 43 **4.** These results suggest that earthworm presence is an important factor underlying 44 natural variation in plant defences against herbivores, and call for a better integration

- of the soil fauna in the studies of plant-herbivore interaction, both for applied and
- 46 fundamental research.

47

- 48 **Key-words**: Endogeic earthworms, detritivore diversity, herbivore-feeding guilds,
- 49 plant nutrients, plant resistance, chemical defences, plant growth-defence trade-off,
- 50 meta-analysis.

Introduction

52

53 In response to the constant threat imposed by herbivores, plants have evolved a broad 54 range of defensive strategies, including mechanical and chemical barriers that reduce 55 herbivore performance (Schoonhoven, Van Loon & Dicke 2005; Agrawal 2007; 56 Johnson 2011). The effect of defensive traits on herbivore performance and fitness is 57 termed plant resistance (Karban & Baldwin 2007), while the ability of the plants to recover from tissue loss is termed tolerance (Strauss & Agrawal 1999; Tiffin 2000; 58 59 Núñez-Farfán, Fornoni & Valverde 2007). Understanding the factors driving variation 60 in plant anti-herbivore strategies remains a core question in ecology (Walling 2000), 61 and advances in this area could be used to inform crop protection (Lyon, Newton & 62 Walters 2014). 63 It is generally assumed that plant ability to defend itself is costly, and thus it 64 should trade off with other life history traits such as growth and reproduction (Coley, 65 Bryant & Chapin 1985; Herms & Mattson 1992; Koricheva 2002). Nonetheless, the 66 consequences of differences in allocation between growth and defences against 67 herbivores vary depending on environmental conditions, such as variation in soil 68 nutrients (Coley, Bryant & Chapin 1985; Fine et al. 2006). 69 For optimal plant growth, soil nutrients must be available in sufficient and 70 balanced amounts (Aerts & Chapin 1999). While soils generally contain a relatively 71 large stock of nutrients, these reserves are usually present in the forms of complexated 72 organic compounds, rendering nutrients inaccessible for plants. The turnover and 73 release of nutrients from soil organic matter (SOM) depend on the rate of

74 decomposition and mineralization of elements through biogeochemical processes 75 (Seastedt 1984; Prescott 2005). Therefore, the efficiency of SOM decomposition and 76 mineralization will influence the magnitude of soil nutrient availability, in turn 77 affecting plant growth and performance (Ladha et al. 2004; Yoshitake, Soutome & 78 Koizumi 2014). Among the highly diverse soil fauna, the invertebrates of the meso-79 and macrofauna are the key organisms participating in SOM turnover and nutrient 80 release (Bardgett & Chan 1999; Edwards 2004; Bhadauria & Saxena 2010), owing to 81 their critical role in breaking down detrital inputs and priming detritus for microbial 82 decomposition (Seastedt 1984; Prescott 2005). 83 Earthworms are among the most important detritivores within soil food webs and 84 are commonly considered as ecosystem engineers (Edwards 2004; Blouin et al. 2013; 85 Cunha et al. 2016). Through their burrowing and casting activities, earthworms 86 improve soil nutrient availability via greater mineralization and/or humification of soil 87 organic matter, modifications of soil porosity and aggregation, and the stimulation of 88 soil microflora (Scheu 2003; Brown, Edwards & Brussaard 2004; van Groenigen et al. 89 2014; Bertrand et al. 2015; Cunha et al. 2016). In addition to these proven 90 growth-promoting effects (e.g. van Groenigen et al. 2014), recent studies have 91 highlighted that earthworms can also benefit plants by increasing their ability to resist 92 herbivore attacks (Wurst et al. 2008; Lohmann, Scheu & Muller 2009; Wurst 2013; 93 Trouve et al. 2014). 94 The mechanisms of earthworm-mediated plant resistance include, for example, an

increase in plant tolerance to herbivores by stimulating plant biomass production

during herbivore attack (Blouin et al. 2005; Wurst et al. 2008). Additionally, earthworms can alter plant resistance by influencing the expression of stress-responsive genes, and subsequently, the production of toxic secondary metabolites (Blouin et al. 2005; Lohmann, Scheu & Muller 2009; Jana et al. 2010). Nonetheless, earthworm effects on plant resistance against herbivores range from negative to positive (e.g. Scheu, Theenhaus & Jones 1999; Johnson et al. 2011; Loranger-Merciris et al. 2012). For instance, the endogeic earthworm *Aporrectodea* caliginosa had a negative effect on the aphid Rhopalosiphum padi (Ke & Scheu 2008), while the anecic earthworm *Lumbricus terrestris* had a positive effect on the same aphid species (Eisenhauer & Scheu 2008). Moreover, the positive impact of earthworms on plant growth could interact with defence allocation (Coley, Bryant & Chapin 1985; Herms & Mattson 1992; Koricheva 2002). Therefore, earthworm effects on plant resistance against herbivores seem to be highly context dependent (Wurst 2010; 2013), but are there general trends that emerge from the literature? We here performed a meta-analysis to formally quantify the effects of earthworms on plant growth and resistance against herbivores, and to identify ecological factors, such as earthworm ecological types and diversity in the soil and herbivore feeding guilds, driving variation in the magnitude of earthworm effects among studies. Earthworm species are classified into three major ecological types (anecic, epigeic and endogeic), which have distinct burrowing patterns. Epigeic earthworms live in litter or topsoil layers where they forage primarily on plant residues. Anecic earthworms live in permanent deep vertical burrows, and endogeic earthworms live in

96

97

98

99

100

101

102

103

104

105

106

107

108

109

110

111

112

113

114

115

116

the soil and forage on soil organic matter (Bouché 1977). Distinct burrowing patterns and food preferences, as well as variation in earthworm density and species richness have been shown to differentially affect soil nutrient mobilization and plant nutrient uptake (Bossuyt, Six & Hendrix 2006; Curry & Schmidt 2007; Spurgeon et al. 2013; Andriuzzi et al. 2016). We therefore hypothesized that the combination of different earthworm ecological types should result in better resource acquisition via niche partitioning, and therefore favour plant growth and nutrient content more than a single earthworm type (Newington et al 2004). In addition, earthworms could modify plant eco-physiological status, in turn affecting the ability of plants to respond to herbivore attack. For instance, Arabidopsis thaliana plants growing in the presence of A. caliginosa showed that enhanced expression of genes involved in phytohormone signalling (e.g. auxin, ethylene, jasmonic acids or salicylic acid), known to respond to biotic and abiotic stresses (Puga-Freitas et al. 2012; Puga-Freitas et al. 2016). Generally, plants activate the jasmonic acid (JA)-dependent signalling pathways in response to tissue-chewing herbivores such as caterpillars and cell-content-feeding herbivores such as thrips (Howe & Jander 2008), whereas salicylic acid (SA)-dependent defences are activated in response to phloem-feeders such as aphids (Stam et al. 2014; Onkokesung et al. 2016). We therefore hypothesized that earthworms could enhance plant resistance against a variety of herbivore types by simultaneously activating several

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

138

phytohormonal pathways.

Finally, given that selection for increased yield in domesticated crops often leads to reduced levels of resistance to herbivores as compared to wild relatives (Rosenthal & Welter 1995; Rosenthal & Dirzo 1997; Whitehead, Turcotte & Poveda 2017), we postulated that the magnitude of earthworm effects on plant growth would be stronger for wild plants, whereas the effects of earthworms on resistance to herbivores would be stronger for crop plants.

We specifically asked the four following questions: 1) Do earthworms increase plant growth and nutrient content? 2) Do earthworms increase plant resistance and defences against herbivores? 3) Which ecological factors (plant type; herbivore feeding guild; earthworm ecological type, earthworm density and species richness) lead to variation in earthworm-mediated plant resistance/defence? 4) Is there a trade-off between earthworm-mediated plant growth and resistance/ defence under herbivore attack? We predicted that: 1) earthworm presence increases plant growth and nutrient content, 2) earthworm presence reduces plant resistance due to increased plant nutritional quality, 3) earthworm effects on plant defences are context dependent, and 4) earthworms have opposing effects on plant growth and resistance.

Materials and methods

DATA COLLECTION

The data set was compiled by conducting keyword searches in the ISI Web of Science up to December 2016 using combinations of relevant terms ("earthworm", "decomposer invertebrate", "ecosystem engineers", "plant growth or tolerance",

"herbivore or herbivory or insect or nematode", "defence or defense or resistance"). Additional searches using the same keywords were conducted in the Google Scholar and reference lists of individual papers were screened to finally obtain a list of studies that met all the following inclusion criteria: 1) plants were subjected to at least two treatments: an earthworm inoculation treatment and control treatment without earthworm; 2) plants in both treatments were under herbivore attack; 3) Concerning plant growth, the study included at least one parameter of plant growth (e.g. aboveground biomass, belowground biomass or total biomass) was measured; concerning plant resistance, the study included at least one measured parameter of plant resistance (i.e. herbivore performance parameters such as growth rate, mass, fecundity, development time, consumption, oviposition preference, density, or the degree of plant damage), and/or plant chemical defences (i.e. secondary metabolite production); and 4) the data included means, some measure of variance, and at least three independent replicates of each treatment. In total, the search yielded 20 papers published between 1999 and 2016 that met our criteria (See Appendix S1 in Supporting Information). However, meta-analyses exclusively based on published studies may produce biased results since the probability of the study to be published could depend upon the statistical significance, magnitude, and/or direction of research findings (Koricheva, Gange & Jones 2009). It has been recommended, therefore, whenever possible, to include unpublished studies and grey literature (e.g. dissertations) in a meta-analysis (Møller & Jennions 2001). By searching in Google using the same keywords as in Web of Science and by contacting individual

161

162

163

164

165

166

167

168

169

170

171

172

173

174

175

176

177

178

179

180

181

researchers, we obtained one published PhD thesis (Kadir 2014), in which the effects of 18 different earthworm combinations on *Brassica rapa* growth and resistance were tested. Finally, we also included two own unpublished studies (Xiao et al., unpublished data shown on Fig. S1). Overall, this grey literature based-dataset includes work done on tomato and corn plants, and represents 15%, 13%, 4%, and 48% of the total sample size for growth, nutrient, resistance, and defence-related effect sizes, respectively (Appendix S1, Fig. S1). To test whether inclusion of our own unpublished datapoints affected the results of the analysis, we performed sensitivity analyses by excluding these data and reanalysing the overall effects for all major variables (see Table S1). Overall, we found no significant differences in results (Table S1 *versus* Tables S2-S5), therefore we report the results of analyses including the unpublished data.

In total, our full searches yielded 79, 64, and 23 datapoints for plant responses in terms of growth, resistance, and defence, respectively (Appendix S1). When available, we also included data that measured earthworm effects on plant nutritional elemental composition (i.e. total carbon, nitrogen and phosphorous concentration), as a measure of how earthworms might modify plant nutrient content (n = 65 datapoints, Appendix S1).

Finally, because of our initial search constraints, earthworm effects on plant growth were assessed when plants were infested with herbivores. We thus aimed at confirming that earthworm effects on plant growth we observed were not masked by the presence of herbivores feeding on the plants. In addition, when available, we

collected a subset of datapoints on plant growth parameter when plants were left herbivore-free, but only if these datapoints came from the same experiments as the dataset described above (n = 25, Appendix S1, Fig. S2). This allowed a direct comparison of the magnitude of earthworm effects on plant growth in the presence and absence of herbivores.

Earthworm effects on plant growth were computed by including any measurements of plant biomass, such as aboveground biomass, belowground biomass, and/or total biomass. When fresh and dry mass were both reported, dry mass was chosen. Earthworm effects on plant resistance against herbivores were assessed by including measures of herbivore growth and development and plant damage imposed by herbivores (Karban & Baldwin 2007). Earthworm effects on plant chemical defences were assessed by including all data on plant secondary metabolites (Appendix S1).

We included multiple outcomes per study when data were reported from several independent experiments, tested on different plant species, or reported for treatments with different ecological type, species richness and density of earthworms. However, if repeated measurements of plant growth and/or resistance were available from the same experiment, only the last date of the measurements was used. If the experiments included additional treatments (e.g. manipulative drought and ambient rainfall patterns), only data of the ambient (control) condition were used. For each observation we extracted the means of the control treatment (without earthworm) and the experimental treatment (with earthworms), as well as their standard deviation (SD)

formula SD = SE * sqrt (n). If data were presented in graphical form, we extracted data points using the GetData software (http://www.getdata-graph-digitizer.com).

Following van Groenigen et al. (2014), our initial dataset included five categorical moderating variables that were used to explore additional sources of variation across the treatments: 1) herbivore feeding guild (three levels: cell-feeding herbivores including nematodes and thrips; chewing herbivores including slugs, caterpillars and rootworms; and phloem-feeding herbivores including aphids), 2) plant type (two levels: wild plants *versus* crops), 3) earthworm ecological type (four levels: epigeic alone, endogeic alone, anecic alone, and mixtures of the three ecological categories), 4) earthworm density (four levels: <100, 100-199, 200-400, >400 earthworms per m² of soil), and 5) earthworm species richness (two levels: single species *versus* multi-species) (Appendix S1).

and sample size (n). When SE was reported, we transformed it to SD by using

META-ANALYSIS

Effect sizes for earthworm effects were calculated using the natural logarithm of the response ratio (lnR) (Hedges, Gurevitch & Curtis 1999) of the mean responses in the presence (+E) and the absence (-E) of earthworm such that lnR = ln(+E/-E). For interpretation of the results, mean effects and confidence intervals were back-transformed using the formula: (EXP(lnR)-1)×100 and reported as the percentage changes between control and earthworm additions.

Because higher herbivore performance (e.g. abundance, larva mass etc.) means that plants are less resistant to herbivores whereas higher levels of plant secondary metabolites mean that plants are better defended, the effect sizes for plant resistance and plant defence had different initial signs. In order to compare resistance and defence effect sizes within the same analyses, all resistance effect sizes, beside the development time of herbivores, were calculated as inverse of lnR such as: lnR_{resistance} = ln (+E/-E)⁻¹. Therefore, for all our analyses, effect sizes with positive values indicate that earthworm presence increased plant growth, nutrient content, resistance and defences against herbivores. The variance associated with effect size was calculated from the standard deviation (SD) and sample size (n) associated with each mean value of plant growth, nutrients, resistance and chemical defences, respectively (Koricheva, Gurevitch & Mengersen 2013). Meta-analysis was performed with the 'metafor' package (Viechtbauer 2010) in R (R Development Core Team 2015). First, we estimated the overall effects of earthworms on plant growth, nutrients, resistance, and chemical defences using a random-effects model. The random-effects model was selected because of the across-studies variability and in order to partition the variance into within- and between-studies. In this analysis, individual effect sizes are weighted by the reciprocal of the sum of the variance between-study and sampling variance within study. The restricted maximum likelihood method (REML) was used to estimate between-study variance. The mean effect size was considered as significantly different from zero if

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

its 95% confidence intervals (CIs) did not include zero (Koricheva, Gurevitch & Mengersen 2013).

We assessed potential publication bias in the overall database using funnel plot and the 'trim and fill' method (Jennions et al. 2013). In order to assess the robustness of the observed overall effects of earthworm presence on plant growth, nutrients and resistance/defences, fail-safe numbers (Nfs) were calculated by using Rosenberg's weighted method ($\alpha = 0.05$) (Rosenberg 2005) (See Tables S2-S5). Rosenberg's Nfs indicates how many studies reporting zero effect size would need to be added to the meta-analysis to render the observed effect non-significantly different from zero (Rosenberg 2005).

Next, we performed meta-regressions to explore how multiple moderator variables could affect the earthworm-mediated effect size on plant resistance and defences. Meta-regressions are more effective than standard meta-analytic techniques at examining the impact of moderator variables for studying effect sizes (Benton, 2014). To avoid potential non-independence between moderators, their effects were tested hierarchically as described in Fig. S3. Moderator analyses were performed only when there were at least two levels with large enough sample size (n > 3, Fig. S3). We used mixed-effects models to estimate the effect of each moderator (herbivore type, plant type, earthworm ecological type, earthworm density, and earthworm species richness) on the magnitude of earthworm presence. This model assumes that differences among studies within a group are due to random variation, whereas variation between groups is fixed. With this model, the between-group homogeneity

 (Q_B) was used to estimate the significance of each categorical moderator (Koricheva, Gurevitch & Mengersen 2013). If the Q_B was significant, we inferred that the mean effect size differed between moderator levels, and two moderator levels were considered to be significantly different from one another if their 95% CIs did not overlap.

Finally, we computed correlations between: 1) the effect of earthworms on plant growth versus plant resistance/defences, and 2) the effect of earthworms on plant resistance versus plant nutritional parameters using Pearson's correlation analysis (Table S6-S8). Each data point of the correlation corresponded to an lnR value as calculated above. A significant positive correlation means that an increase in plant resistance in the presence of earthworms is associated with an increase in plant growth and/or plant nutritional parameters.

Results

EARTHWORM EFFECTS ON PLANT GROWTH

Overall, earthworm presence increased plant biomass by 20 % (Fig. 1a, Table S2). Specifically, earthworm presence significantly increased plant aboveground biomass by 16 %, belowground biomass by 29 % and total biomass by 22 % (Fig. 1a, Table S2). The 'trim and fill' method detected three missing studies to the left of the grant mean. The addition of three missing cases to the dataset produced a new grand mean effect size of 19 % (95% CIs: 13 % to 26 %, n = 82), suggesting a robust positive overall effect of earthworms on plant growth in the presence of herbivores (Table S2).

The Rosenberg's Nfs for the overall database is 6420, which is 15 times higher than the threshold of 405 ($5 \times 79 + 10$), also indicating a robust mean effect size (Table S2).

Additionally, by directly comparing the magnitude of earthworm effects on plant growth in the presence and absence of herbivores using a balanced subset (i.e. datapoints come from the same study, n = 25), we found that earthworm presence increased overall plant biomass by 14 % and by 11% when plants grew in the presence and absence of herbivores, respectively (Fig. S2).

EARTHWORM EFFECTS ON PLANT NUTRIENT CONTENT

Earthworm presence stimulated an overall 11 % increase in plant nutrient content in the presence of herbivores (Rosenberg's Nfs = 19035, n = 65, Fig. 1b, Table S3). The addition of 14 missing cases to the dataset by the 'trim and fill' method produced a new grand mean effect size of 21 % (95% CIs: 12 % to 31 %, n = 79), suggesting a robust positive overall effect of earthworms on plant nutrient content in response to herbivory (Table S3). This result was mainly driven by a 21 % increase in plant nitrogen content, while we detected a 20 % decrease in phosphorus and a 1% decrease in carbon content when earthworms were present (Fig. 1b, Table S3).

EARTHWORM EFFECTS ON PLANT RESISTANCE

Overall, earthworm presence decreased plant resistance to herbivores by 15% (95% CIs: -24 % to -4 %, n = 64). After 6 missing cases were added to the analysis by the

'trim and fill' method, the new grand mean effect size was -9 % (95% CIs: -19 % to 3 %, n = 70) (Table S4). Between-study variation explained 83 % of the observed variation in the magnitude of the effect. While plant cultivation type did not influence earthworm effects on plant resistance ($Q_B = 0.04$, df = 1, p = 0.844), we found a strong effect of herbivore type ($Q_B = 12.098$, df = 2, p = 0.002). Earthworm presence increased plant resistance to cell-feeders by 34 % (and by 50 % after adding two missing cases with the 'trim and fill' method; Table S4). This result was mainly driven by 80 % increase in plant resistance to thrips and 11% increase in resistance to root-feeding nematodes (Fig. 2a, Table S4). In contrast, earthworm presence had no significant effect on plant resistance to chewing herbivores (Fig. 2b, Table S4), and decreased plant resistance to phloem-feeders by 22 % (Fig. 2c, Table S4). We therefore proceeded to explore the possible causes of this heterogeneity using moderator analyses (including earthworm ecological type, species richness and density) with chewing and phloem-feeding herbivores separately (Fig. S3). Earthworm ecological type and species richness did not affect earthworm effects on plant resistance against chewing herbivores (Fig. 2b, Table S4). Plant resistance against phloem-feeders was particularly decreased when a mixture of the three earthworm ecological types or a mixture of different species of earthworms (multi-species) was added in the experiments, and when earthworm densities were high (i.e. above 400 individuals m⁻²) (Fig. 2c, Table S4).

355

356

335

336

337

338

339

340

341

342

343

344

345

346

347

348

349

350

351

352

353

354

EARTHWORM EFFECTS ON PLANT CHEMICAL DEFENCES

Overall, earthworm presence did not significantly affect plant defence compounds (Fig. 3). Between-study variation explained 81 % of the observed variation in the magnitude of the effect. Again, while plant type did not affect earthworm effects on plant chemical defences ($Q_B = 2.659$, df = 1, p = 0.103), we found a strong effect of herbivore type ($Q_B = 12.139$, df = 2, p = 0.002). Specifically, we found that earthworms had no effect on chemical defences in the presence of chewing herbivores (Table S5). However, earthworm presence increased overall chemical defences by 32 % in the presence of cell-feeding herbivores; this result was driven by a 38 % increase in defensive compounds in the presence of nematodes and a 31% increase in the presence of thrips (Table S5). Additionally, earthworm presence decreased chemical defences by 48 % in the presence of phloem-feeders (Table S5), although this result was driven by one data-point only.

Because of lack of data for phloem-feeding and chewing herbivores (Fig. S3), we proceeded to perform moderator analyses only for the cell-feeding herbivores (thrips). We found that single-species earthworm inoculations significantly increased plant chemical defences in the presence of cell-feeding thrips (Table S5). In addition, earthworm-mediated plant chemical defences against thrips were not dependent on earthworm ecological type (Table S5).

EARTHWORM-MEDIATED RELATIONSHIPS BETWEEN PLANT GROWTH, NUTRIENTS,

RESISTANCE AND DEFENCES

Effects of earthworm presence on plant resistance were negatively correlated with earthworm effects on plant growth (Fig. 4a, Table S6). However, this relationship was affected by herbivore type, plant type, earthworm ecological type, density, and species richness (Table S7). Negative correlations between earthworm effects on plant resistance and growth were strongest against phloem-feeders (r = -0.48, p = 0.008), in wild plants (r = -0.51, p = 0.009), with endogeic earthworm inoculations (r = -0.53, p = 0.008), with earthworm density below 100 individuals m^{-2} (r = -0.95, p = 0.012) and with earthworm multi-species inoculations (r = -0.52, p = 0.022) (Table S7). On the other hand, earthworm presence mediated an overall positive relationship between plant growth and chemical defences (r = 0.48, p = 0.021, Fig. 4b, Table S6). This positive earthworm-mediated relationship was strongest in crop plants (r = 0.52, p =0.025, Table S7), with earthworm single species treatment (r = 0.49, p = 0.045, Table S7), and with earthworm multi-species treatment (r = 0.97 p < 0.001, Table S7). Effects of earthworm presence on plant resistance were negatively correlated with earthworm effects on plant nutrients only when earthworms were endogeic species (r = -0.42, p = 0.032), and their density was less than 100 individuals m^{-2} (r = -0.98, p = 0.003) and 200-400 individuals m⁻² (r = -0.61, p = 0.026) (Table S8). Effects of earthworm presence on plant phosphorus content were negatively correlated with earthworm effects on plant chemical defences (Table S6). Finally, effects of earthworm presence on plant growth were positively correlated with earthworm effects on plant nutrient content, total nitrogen and carbon in particular. (Table S6).

399

398

378

379

380

381

382

383

384

385

386

387

388

389

390

391

392

393

394

395

396

Discussion

We found that earthworm presence had an overall positive effect on plant growth and nutritional content, and variable overall effects on plant resistance and chemical defences. The results were strongly dependent on the herbivore feeding guild, as well as on the ecological type, density and species richness of earthworms.

EARTHWORM EFFECTS ON PLANT GROWTH

We found an overall positive effect of earthworms on plant biomass gain against herbivores (20 % more biomass on plants inoculated with earthworms) (Fig. 1a). This is in line with previous results that extrapolated the positive effects of earthworms on plant production (e.g. van Groenigen et al. 2014). In addition to the previous studies, our subset data enabled a direct comparison of the effects of earthworms on plant growth in the presence or absence of herbivores. We found that the magnitude of the relative change in biomass of plants that experienced herbivores (14 %) was similar to that of herbivore-free plants (11 %) (Fig. S2), indicating that herbivores did not attenuate the effects of earthworms on plant growth.

Because herbivores are generally thought to decrease plant biomass, these results might be suggestive of an earthworm-mediated tolerance in plants under herbivore attack. While the meta-analysis could not tease apart the mechanisms behind plant growth enhancement in the presence of earthworms, the compensatory continuum hypothesis (CCH) asserts that plants have a greater potential for compensating for herbivore damage under resource-rich conditions (Maschinski & Whitham 1989).

Therefore, earthworms could favour tolerance responses of plants by increasing soil nutrient availability.

Plant resistance against herbivores is generally mediated by changes in nutritional

424

425

426

427

428

429

430

431

432

433

434

435

436

437

438

439

440

441

442

443

422

423

EARTHWORM EFFECTS ON PLANT RESISTANCE

quality and/or production of toxic secondary metabolites. Earthworms have been shown to affect primary and secondary metabolism in plants, as well as the expression of stress-responsive genes in both aboveground and belowground parts of plants, thus potentially impacting herbivore performance (Blouin et al. 2005; Lohmann, Scheu & Muller 2009; Jana et al. 2010). We found that earthworms increased plant susceptibility to phloem feeders, but increased resistance to cell-feeding herbivores, and had no effect on resistance to chewing herbivores. Across the studies involving the phloem feeders (aphids), we observed an overall increase in abundance of the herbivores when earthworms were present. Nonetheless, these results were context-dependent. In particular, only high densities and higher levels of species and functional diversity of earthworms decreased plant resistance against aphid herbivores. Under aphid attack, plants activate the SA pathway for stimulating chemical and physical barriers such callose deposition and the production of defensive secondary metabolites, which are transported into the phloem to increase toxicity (Elzinga & Jander 2013; Züst & Agrawal 2016). In turn, aphids could inject effector proteins to prevent callose deposition, and deal with toxic metabolites by metabolization or excretion (Kim & Jander 2007; Elzinga & Jander 2013; Züst &

444 Agrawal 2016). Earthworms, therefore, could favour plant susceptibility to aphids by 445 inhibiting the SA signalling pathway. While earthworms have been shown to affect 446 plant defence signalling pathways and gene expression (Puga-Freitas et al. 2012; 447 Puga-Freitas et al. 2016), we are not aware of studies directly linking earthworm 448 presence to plant physiological and molecular mechanisms for deterring aphid attack, 449 but this should be considered for future avenues of research. 450 In addition, accessible nutrients, such as sugars, amino acids and nitrogen are also 451 important determinants for the growth and development of herbivores including 452 aphids (Mattson 1980; Caillaud et al. 1995; Cao et al. 2016). Therefore, the positive 453 effects of earthworms on plant nutritional quality (e.g. higher nitrogen content), might 454 also cause increased plant susceptibility to aphids. This idea is corroborated by the 455 fact that in a more complex earthworm community, earthworm species act 456 synergistically to increase soil fertility (Curry & Schmidt 2007; Spurgeon et al. 2013; 457 Bertrand et al. 2015) and plant nutrient content (e.g. Laossi et al. 2009), in turn 458 increasing plant susceptibility to aphid attack. 459 Contrary to the aphids, earthworms mediated increased plant resistance against 460 cell-feeders. This was particularly true when measuring resistance against thrips (Fig. 461 2a), while the effects were more variable when measuring resistance against 462 soil-dwelling nematodes. While the effects of earthworms on nematodes could 463 partially be explained by direct interference (i.e. earthworms could ingest nematodes 464 while ingesting the surrounding substrate (Boyer et al. 2013)), the effects of 465 earthworms on thrips are likely to be mediated by changes in aboveground plant

functional traits. Our unpublished study, as described in Fig. S1, showed that earthworm-inoculated plants under thrips attack had higher concentrations of total carbon and nitrogen, lower concentrations of total phosphorus, and higher levels of jasmonic acid and total phenolic compounds (Fig. S1 d, e). Earthworm-mediated increase in resistance against thrips can thus be due to the activation of the JA signalling pathway. In addition, we speculate that stimulation of the soil microbial community by earthworms could have enhanced defence priming in plants (Blouin et al. 2005; Jana et al. 2010; Puga-Freitas et al. 2012), and ultimately increase resistance by promoting chemical defence accumulation in the plants. This however, has never been specifically tested so far.

EARTHWORM EFFECTS ON PLANT CHEMICAL DEFENCES

We found that overall, earthworm presence did not significantly affect plant chemical defences when chewing herbivores were on plants, but notably increased chemical defences when cell-content feeders, particularly thrips were present. For example, earthworm presence promoted the induction of defence compounds such as jasmonic acid and phenolics in tomato leaves when under thrips attack (Fig S1e). Similarly, earthworms significantly increased concentrations of total glucosinolates, a nitrogen-based defence compound class, in leaves of *Sinapis alba* (Lohmann, Scheu & Muller 2009). Therefore, in these cases, earthworm presence could favour plant resistance by increasing plant chemical defences. On the other hand, Wurst et al. (2006) showed that concentrations of two glucosinolates (glucoiberin and

glucoraphanin) in shoots of *Brassica oleracea* decreased when the endogeic earthworm *Octolasion tyrtaeum* was added to the system. Similarly, earthworms could induce a decline of root sesquiterpene (E)- β -caryophyllene when rootworms are present (Fig. S1 j).

488

489

490

491

492

493

494

495

496

497

498

499

500

501

502

503

504

505

506

507

508

509

The inconsistent effects of earthworms on plant chemistry might be due to the interactive effects of plant growth and nutrient uptake on plant secondary metabolism. For instance, it was shown that phytosterol concentration in *Plantago lanceolata* plants was not affected by earthworms directly, but increased with increasing nitrogen concentration of the leaves (Wurst et al. 2004), which is mediated by earthworm presence (Wurst & Jones 2003). Additionally, several studies have shown that the initial soil nutrient content and the distribution of soil organic matter could influence plant defensive secondary metabolites (Wurst et al. 2003; Wurst, Dugassa-Gobena & Scheu 2004; Ke & Scheu 2008). For instance, earthworms favoured an increase in total phytosterol content of *P. lanceolata* shoots, but only when the spatial distribution of organic residues/litter was mixed homogeneously with soil (Wurst, Dugassa-Gobena & Scheu 2004). Only few studies in our meta-analysis addressed the effects of differences in initial soil properties such as distribution of organic litter, or the changes in soil available nutrients (e.g. mineral nitrogen), driven by earthworm presence. This prevented the use of soil bio-chemical properties as a moderator in this study. Nonetheless, an increasing number of studies demonstrate that soil nutrients and microorganisms both modify the synthesis of defensive secondary metabolites (e.g. Ohkama-Ohtsu & Wasaki 2010; Badri et al. 2013), and ultimately influence

plant-herbivore interactions (Badri et al. 2013; Pineda et al. 2013). This indeed calls for a better integration of earthworms living in different soil conditions and with different ecologies into plant-herbivore interaction studies.

513

514

515

516

517

518

519

520

521

522

523

524

525

526

527

528

529

530

531

510

511

512

EARTHWORM EFFECTS ON THE TRADE-OFFS BETWEEN PLANT PERFORMANCE,

RESISTANCE AND CHEMICAL DEFENCES

We found that the effects of earthworms on growth and resistance of plants under herbivory were overall negatively correlated (Fig. 4a), as would be predicted by classic plant defence theory (Herms & Mattson 1992; Züst & Agrawal 2017). An increasing number of studies indicate that earthworms could indirectly influence the performance of herbivores such as phloem-feeders by predominantly affecting plant size, vigour, and nutrient content (Scheu, Theenhaus & Jones 1999; Eisenhauer & Scheu 2008; Trouve et al. 2014), and to a lesser extent by changes in plant secondary chemistry (Francis et al. 2001; Wurst et al. 2004; Katsanis, Rasmann & Mooney 2016). For example, Cao et al. (2016) showed that the green peach aphid (Myzus persicae) performed better on an enhanced amino acid: sugar ratio and enhanced absolute amino acid concentration in the phloem, but also activated genes responsible for glucosinolates synthesis in the leaves of Chinese cabbage. Similarly, Wurst et al. (2004) showed that A. caliginosa earthworm presence decreased the concentration of the defence compound catalpol in *P. lanceolata* leaves, but this was not positively correlated with the performance in term of development time of the aphid M. persicae.

Contrary to expectations, earthworm presence simultaneously increased both plant growth and chemical defences (Fig.4b). These effects were particularly strong on crop plants. Because of lack of data, we could not highlight a particular combination of factors explaining these results, besides the fact that bigger plants had higher level of secondary metabolites, independently of any particular plant by herbivore by earthworm combination. Several studies have shown that the assumed growth-defence trade-off might be modified (i.e. reduced or even reversed) by different levels of nutrients in the soil (Coley, Bryant & Chapin 1985; Donaldson, Kruger & Lindroth 2006; Lind et al. 2013), or not detected due to the failure to address the appropriate measure of growth-related traits (Züst et al. 2011; Züst, Rasmann & Agrawal 2015). Overall, these different patterns suggest that earthworm effects on defence allocation in plants are in part dictated by resource allocation, and are highly context dependent in terms of categories of defence compounds. However, this needs to be systematically addressed in future manipulative studies.

Acknowledgements

We thank all authors of the original studies included in our meta-analysis and Dr. Sharon Zytynska who responded to our queries on specific details on individual study systems. We are also grateful to the editor and the two anonymous referees for their constructive comments on previous versions of the manuscript. This work was funded by the National Key R&D program (2016YFD0200305), National Natural Science Foundation of China grants (31170487 and 41371263) to ML, and by a Swiss Science

554 Foundation grant (31003A_159869) to SR. ZX was supported by the China 555 Scholarship Council (201506850024) and Fundamental Research Fund for the Central 556 Universities (KYTZ2014). 557 **Data accessibility** 558 559 All data used in the meta-analysis are present in the manuscript or online supporting 560 information and deposited in the Dryad Digital Repository: 561 http://doi.org/10.5061/dryad.382k1 (Xiao et al. 2017). 562 **Authorship** 563 564 ZX, ML, FH and SR designed the study. ZX, XW and AK collected data and 565 performed the meta-analysis. JK assisted with the meta-analysis. ZX and SR wrote the 566 first draft of the manuscript and all authors contributed to revisions.

References

568

569 Aerts, R. & Chapin, F.S. (1999) The mineral nutrition of wild plants revisited: a re-evaluation of 570 processes and patterns. Advances in Ecological Research, 30, 1–67. 571 Agrawal, A.A. (2007) Macroevolution of plant defense strategies. Trends in Ecology & Evolution, 572 **22,** 103–109. 573 Andriuzzi, W.S., Schmidt, O., Brussaard, L., Faber, J.H. & Bolger, T. (2016) Earthworm 574 functional traits and interspecific interactions affect plant nitrogen acquisition and 575 primary production. Applied Soil Ecology, 104, 148–156. 576 Badri, D.V., Zolla, G., Bakker, M.G., Manter, D.K. & Vivanco, J.M. (2013) Potential impact of 577 soil microbiomes on the leaf metabolome and on herbivore feeding behavior. New 578 Phytologist, 198, 264-273. 579 Bardgett, R.D. & Chan, K.F. (1999) Experimental evidence that soil fauna enhance nutrient 580 mineralization and plant nutrient uptake in montane grassland ecosystems. Soil Biology 581 and Biochemistry, **31,** 1007–1014. 582 Benton, T., 2014. Using meta-regression to explore moderating effects in surveys of international 583 achievement. Practical Assessment Research and Evaluation 19, 3. 584 Bertrand, M., Barot, S., Blouin, M., Whalen, J., de Oliveira, T. & Roger-Estrade, J. (2015) 585 Earthworm services for cropping systems. A review. Agronomy for Sustainable 586 Development, 35, 553-567. 587 Bhadauria, T. & Saxena, K.G. (2010) Role of earthworms in soil fertility maintenance through the 588 production of biogenic structures. Applied and Environmental Soil Science, 2010, 1–7. 589 Blouin, M., Zuily-Fodil, Y., Pham-Thi, A.-T., Laffray, D., Reversat, G., Pando, A., Tondoh, J. &

590	Lavelle, P. (2005) Belowground organism activities affect plant aboveground phenotype,
591	inducing plant tolerance to parasites. Ecology letters, 8, 202–208.
592	Blouin, M., Hodson, M.E., Delgado, E.A., Baker, G, Brussard, L., Butt, K.R., Dai, J., Dendoover
593	L., Peres, G., Tondoh, J.E., Cluzeau, D., Brun, JJ. (2013). A review of earthworm impact o
594	soil function and ecosystem services. European Journal of Soil Science, 64, 161–182.
595	Bossuyt, H., Six, J. & Hendrix, P.F. (2006) Interactive effects of functionally different earthworm
596	species on aggregation and incorporation and decomposition of newly added residue
597	carbon. <i>Geoderma</i> , 130 , 14–25.
598	Bouché, M. B. (1977). Strategies lombriciennes. Ecological Bulletins, 122–132.
599	Boyer, J., Reversat, G., Lavelle, P. & Chabanne, A. (2013) Interactions between earthworms and
600	plant-parasitic nematodes. European Journal of Soil Biology, 59, 43–47.
601	Brown, G.G., Edwards, C.A. & Brussaard, L. (2004) How earthworms affect plant growth:
602	burrowing into the mechanisms. Earthworm Ecology, 2, 13–49.
603	Cao, H.H., Liu, H.R., Zhang, Z.F. & Liu, T.X. (2016) The green peach aphid Myzus persicae
604	perform better on pre-infested Chinese cabbage Brassica pekinensis by enhancing host
605	plant nutritional quality. Scientific Reports, 6, 21954.
606	Caillaud, C.M., Pierre, J., Chaubet, B. & Pietro, J. (1995) Analysis of wheat resistance to the
607	cereal aphid Sitobion avenae using electrical penetration graphs and flow charts
608	combined with correspondence analysis. Entomologia Experimentalis et Applicata, 75, 9-
609	18.
610	Coley, P.D., Bryant, J.P. & Chapin, F.S. (1985) Resource availability and plant antiherbivore
611	defense. Science, 230, 895–899.

612 Cunha, L., Brown, G.G., Stanton, D.W.G., Da Silva, E., Hansel, F.A., Jorge, G., McKey, D., 613 Vidal-Torrado, P., Macedo, R.S., Velasquez, E., James, S.W., Lavelle, P. & Kille, P. (2016) 614 Soil animals and pedogenesis: the role of earthworms in anthropogenic soils. Soil Science, 615 181, 110-125. 616 Curry, J.P. & Schmidt, O. (2007) The feeding ecology of earthworms – A review. *Pedobiologia*, **50**, 617 463-477. 618 Donaldson, J.R., Kruger, E.L. & Lindroth, R.L. (2006) Competition- and resource-mediated 619 tradeoffs between growth and defensive chemistry in trembling aspen (Populus 620 tremuloides). New Phytologist, 169, 561–570. 621 Edwards, C.A. (2004) Earthworm ecology. CRC Press. 622 Eisenhauer, N. & Scheu, S. (2008) Earthworms as drivers of the competition between grasses and 623 legumes. Soil Biology and Biochemistry, 40, 2650–2659. 624 Elzinga, D.A. & Jander, G. (2013) The role of protein effectors in plant-aphid interactions. 625 Current Opinion in Plant Biology, 16, 451-456. 626 Fine, P.V., Miller, Z.J., Mesones, I., Irazuzta, S., Appel, H.M., Stevens, M.H.H., Sääksjärvi, I., 627 Schultz, J.C. & Coley, P.D. (2006) The growth-defense trade-off and habitat 628 specialization by plants in Amazonian forests. *Ecology*, **87**, S150–S162. 629 Francis, F., Lognay, G., Wathelet, J.-P. & Haubruge, E. (2001) Effects of Allelochemicals from 630 First (Brassicaceae) and Second (Myzus persicae and Brevicoryne brassicae) Trophic 631 Levels on *Adalia bipunctata*. *Journal of Chemical Ecology*, **27**, 243–256. 632 Hedges, L.V., Gurevitch, J. & Curtis, P.S. (1999) The meta - analysis of response ratios in 633 experimental ecology. Ecology, 80, 1150–1156.

634 Herms, D.A. & Mattson, W.J. (1992) The dilemma of plants: to grow or defend. Quarterly Review 635 of Biology, 283-335. 636 Howe, G.A. & Jander, G. (2008) Plant immunity to insect herbivores. Annual Review of Plant 637 Biology, 59, 41-66. 638 Jana, U., Barot, S., Blouin, M., Lavelle, P., Laffray, D. & Repellin, A. (2010) Earthworms 639 influence the production of above- and belowground biomass and the expression of genes 640 involved in cell proliferation and stress responses in Arabidopsis thaliana. Soil Biology 641 and Biochemistry, 42, 244-252. 642 Jennions, M.D., Lortie, C.J., Rosenberg, M.S. & Rothstein, H.R. (2013) Publication and related 643 biases. Handbook of Meta-analysis in Ecology and Evolution, 207–236. 644 Johnson, M.T.J. (2011) Evolutionary ecology of plant defences against herbivores. Functional 645 Ecology, **25**, 305–311. 646 Johnson, S.N., Staley, J.T., McLeod, F.A.L. & Hartley, S.E. (2011) Plant-mediated effects of soil 647 invertebrates and summer drought on above-ground multitrophic interactions. Journal of 648 Ecology, 99, 57-65. 649 Kadir, N. (2014) Plant mediated effects of earthworms on aphid dynamics. Plymouth University 650 Press, Plymouth. 651 Karban, R., and Baldwin, I. T. (2007). Induced responses to herbivory. University of Chicago 652 Press, Chicago. 653 Katsanis, A., Rasmann, S. & Mooney, K.A. (2016) Herbivore diet breadth and host plant defense 654 mediate the tri-trophic effects of plant toxins on multiple coccinellid predators. PLoS 655 ONE, 11, e0155716.

656 Ke, X. & Scheu, S. (2008) Earthworms, Collembola and residue management change wheat 657 (*Triticum aestivum*) and herbivore pest performance (Aphidina: *Rhophalosiphum padi*). 658 Oecologia, 157, 603-617. 659 Kim, J.H. & Jander, G. (2007) Myzus persicae (green peach aphid) feeding on Arabidopsis 660 induces the formation of a deterrent indole glucosinolate. *Plant Journal*, **49**, 1008-1019. 661 Koricheva, J. (2002) Meta - analysis of sources of variation in fitness costs of plant antiherbivore 662 defenses. Ecology, 83, 176-190. 663 Koricheva, J., Gange, A.C. & Jones, T. (2009) Effects of mycorrhizal fungi on insect herbivores: a 664 meta - analysis. *Ecology*, **90**, 2088-2097. 665 Koricheva, J., Gurevitch, J. & Mengersen, K. (2013) Handbook of meta-analysis in ecology and 666 evolution. Princeton University Press. 667 Ladha, J., Khind, C., Khera, T. & Bueno, C. (2004) Effects of residue decomposition on 668 productivity and soil fertility in rice-wheat rotation. Soil Science Society of America 669 Journal, 68, 854-864. 670 Lafont, A., Risède, J.-M., Loranger-Merciris, G., Clermont-Dauphin, C., Dorel, M., Rhino, B. & 671 Lavelle, P. (2007) Effects of the earthworm *Pontoscolex corethrurus* on banana plants 672 infected or not with the plant-parasitic nematode Radopholus similis. Pedobiologia, 51, 673 311-318. 674 Laossi, K.-R., Noguera, D.C., Bartolomé-Lasa, A., Mathieu, J., Blouin, M. & Barot, S. (2009) 675 Effects of an endogeic and an anecic earthworm on the competition between four annual 676 plants and their relative fecundity. Soil Biology and Biochemistry, 41, 1668–1673. 677 Lind, E.M., Borer, E., Seabloom, E., Adler, P., Bakker, J.D., Blumenthal, D.M., Crawley, M.,

678 Davies, K., Firn, J., Gruner, D.S., Harpole, W.S., Hautier, Y., Hillebrand, H., Knops, J., 679 Melbourne, B., Mortensen, B., Risch, A.C., Schuetz, M., Stevens, C. & Wragg, P.D. 680 (2013) Life-history constraints in grassland plant species: a growth-defence trade-off is 681 the norm. *Ecology Letters*, 16, 513–521. 682 Lohmann, M., Scheu, S. & Muller, C. (2009) Decomposers and root feeders interactively affect 683 plant defence in Sinapis alba. Oecologia, 160, 289–298. 684 Loranger-Merciris, G., Cabidoche, Y.M., Deloné, B., Quénéhervé, P. & Ozier-Lafontaine, H. 685 (2012) How earthworm activities affect banana plant response to nematodes parasitism. 686 *Applied Soil Ecology*, **52**, 1–8. 687 Lyon, G.D., Newton, A.C. & Walters, D.R. (2014) Induced resistance in crop protection: the future, 688 drivers and barriers. Induced resistance for plant defense: a sustainable approach to crop 689 protection, 316-325. Oxford: Blackwell Publishing. 690 Møller, A.P. & Jennions, M.D. (2001) Testing and adjusting for publication bias. Trends in 691 Ecology & Evolution, 16, 580-586. 692 Maschinski, J. & Whitham, T.G. (1989) The continuum of plant responses to herbivory: the 693 influence of plant association, nutrient availability, and timing. The American Naturalist, 694 1-19.695 Mattson, W.J. (1980) Herbivory in relation to plant nitrogen content. Annual Review of Ecology 696 and Systematics, 11, 119–161. 697 Núñez-Farfán, J., Fornoni, J. & Valverde, P.L. (2007) The evolution of resistance and tolerance to 698 herbivores. Annual Review of Ecology, Evolution, and Systematics, 541–566. 699 Newington, J., Setälä, H., Bezemer, T. & Jones, T. (2004) Potential effects of earthworms on

- leaf chewer performance. *Functional Ecology*, **18**, 746–751.
- 701 Ohkama-Ohtsu, N. & Wasaki, J. (2010) Recent progress in plant nutrition research: cross-talk
- 702 between nutrients, plant physiology and soil microorganisms. *Plant Cell Physiology*, **51**,
- 703 1255–1264.
- Onkokesung, N., Reichelt, M., van Doorn, A., Schuurink, R.C. & Dicke, M. (2016) Differential
- 705 costs of two distinct resistance mechanisms induced by different herbivore species in
- 706 arabidopsis. *Plant Physiology*, **170**, 891–906.
- 707 Pieterse, C.M., Van der Does, D., Zamioudis, C., Leon-Reyes, A. & Van Wees, S.C. (2012)
- 708 Hormonal modulation of plant immunity. Annual Review of Cell and Developmental
- 709 Biology, **28**, 489–521.
- 710 Pineda, A., Dicke, M., Pieterse, C.M.J., Pozo, M.J. & Biere, A. (2013) Beneficial microbes in a
- 711 changing environment: are they always helping plants to deal with insects? *Functional*
- 712 *Ecology*, **27**, 574–586.
- 713 Pineda, A., Soler, R., Pozo, M.J., Rasmann, S. & Turlings, T.C. (2015) Editorial:
- Above-belowground interactions involving plants, microbes and insects. Frontiers in
- 715 *Plant Science*, **6**, 318. doi:10.3389/fpls.2015.00318.
- Pineda, A., Zheng, S.J., van Loon, J.J., Pieterse, C.M. & Dicke, M. (2010) Helping plants to deal
- 717 with insects: the role of beneficial soil-borne microbes. *Trends in Plant Science* 15:507–
- 718 514.
- 719 Prescott, C. (2005) Decomposition and mineralization of nutrients from litter and humus. In:
- Nutrient Acquisition by Plants. Springer, pp. 15–41.
- 721 Puga-Freitas, R., Barot, S., Taconnat, L., Renou, J.P. & Blouin, M. (2012) Signal molecules

722 mediate the impact of the earthworm Aporrectodea caliginosa on growth, development 723 and defence of the plant Arabidopsis thaliana. PLoS ONE, 7, e49504. 724 Puga-Freitas, R., Belkacem, L., Barot, S., Bertrand, M., Roger-Estrade, J. & Blouin, M. (2016) 725 Transcriptional profiling of wheat in response to take-all disease and mechanisms 726 involved in earthworm's biocontrol effect. European Journal of Plant Pathology, 144, 727 155–165. 728 R Development Core Team (2015) R: A language and environment for statistical computing, R 729 Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL: 730 http://www.R-project.org. 731 Rosenberg, M.S. (2005) The file-drawer problem revisited: a general weighted method for 732 calculating fail-safe numbers in meta-analysis. *Evolution*, **59**, 464–468. 733 Rosenthal, J. & Welter, S. (1995) Tolerance to herbivory by a stemboring caterpillar in 734 architecturally distinct maizes and wild relatives. *Oecologia*, **102**, 146–155. 735 Rosenthal, J.P. & Dirzo, R. (1997) Effects of life history, domestication and agronomic selection 736 on plant defence against insects: evidence from maizes and wild relatives. Evolutionary 737 Ecology, 11, 337–355. 738 Scheu, S. (2003) Effects of earthworms on plant growth: patterns and perspectives. *Pedobiologia*, 739 **47,** 846–856. 740 Scheu, S., Theenhaus, A. & Jones, T.H. (1999) Links between the detritivore and the herbivore 741 system: effects of earthworms and Collembola on plant growth and aphid development. 742 *Oecologia*, **119**, 541–551. 743 Schoonhoven, L.M., Van Loon, J.J. & Dicke, M. (2005) Insect-plant biology. Oxford University

- 744 Press.
- 745 Schwachtje, J. & Baldwin, I.T. (2008) Why does herbivore attack reconfigure primary metabolism?
- 746 *Plant Physiology,* **146,** 845–851.
- 747 Seastedt, T. (1984) The role of microarthropods in decomposition and mineralization processes.
- 748 Annual Review of Entomology, **29**, 25–46.
- 749 Spurgeon, D.J., Keith, A.M., Schmidt, O., Lammertsma, D.R. & Faber, J.H. (2013) Land-use and
- 750 land-management change: relationships with earthworm and fungi communities and soil
- structural properties. *BMC ecology,* **13,** 1–13.
- 752 Stam, J.M., Kroes, A., Li, Y., Gols, R., van Loon, J.J., Poelman, E.H. & Dicke, M. (2014) Plant
- 753 interactions with multiple insect herbivores: from community to genes. *Annual Review of*
- 754 *Plant Biology* **65**:689–713.
- 755 Strauss, S.Y. & Agrawal, A.A. (1999) The ecology and evolution of plant tolerance to herbivory.
- 756 *Trends in Ecology & Evolution,* **14,** 179–185.
- 757 Tiffin, P. (2000) Mechanisms of tolerance to herbivore damage: what do we know? *Evolutionary*
- 758 *Ecology,* **14,** 523-536.
- 759 Trouve, R., Drapela, T., Frank, T., Hadacek, F. & Zaller, J.G. (2014) Herbivory of an invasive slug
- in a model grassland community can be affected by earthworms and mycorrhizal fungi.
- 761 Biology and Fertility of Soils, **50**, 13–23.
- van Groenigen, J.W., Lubbers, I.M., Vos, H.M., Brown, G.G., De Deyn, G.B. & van Groenigen,
- 763 K.J. (2014) Earthworms increase plant production: a meta-analysis. Scientific Reports
- 764 4:6365. doi:10.1038/srep06365.
- van Loon, L.C., Bakker, P.A. & Pieterse, C.M. (1998) Systemic resistance induced by rhizosphere

- bacteria. *Annual Review of Phytopathology*, **36**, 453–483.
- 767 Viechtbauer, W. (2010) Conducting meta-analyses in R with the metafor package. *Journal of*
- 768 *Statistical Software*, **36**, 1–48.
- 769 Walling, L.L. (2000) The Myriad Plant Responses to Herbivores. Journal of Plant Growth
- 770 Regulation, **19**, 195–216.
- Whitehead, S.R., Turcotte, M.M. & Poveda, K. (2017) Domestication impacts on plant-herbivore
- interactions: a meta-analysis. Philosophical Transactions of the Royal Society of London.
- 773 *Series B: Biological Sciences*, **372**: 20160034
- 774 Wurst, S. (2010) Effects of earthworms on above- and belowground herbivores. Applied Soil
- 775 *Ecology,* **45,** 123–130.
- 776 Wurst, S. (2013) Plant-mediated links between detritivores and aboveground herbivores. Frontiers
- 777 in Plant Science, 4.
- Wurst, S., Allema, B., Duyts, H. & Van Der Putten, W.H. (2008) Earthworms counterbalance the
- 779 negative effect of microorganisms on plant diversity and enhance the tolerance of grasses
- 780 to nematodes. *Oikos*, **117**, 711–718.
- Wurst, S., Dugassa-Gobena, D., Langel, R., Bonkowski, M. & Scheu, S. (2004) Combined effects
- of earthworms and vesicular-arbuscular mycorrhizas on plant and aphid performance.
- 783 *New Phytologist,* **163,** 169–176.
- Wurst, S., Dugassa-Gobena, D. & Scheu, S. (2004) Earthworms and litter distribution affect
- plant-defensive chemistry. *Journal of Chemical Ecology*, **30**, 691–701.
- Wurst, S. & Jones, T.H. (2003) Indirect effects of earthworms (*Aporrectodea caliginosa*) on an
- above-ground tritrophic interaction. *Pedobiologia*, **47**, 91–97.

788	Wurst, S., Langel, R., Reineking, A., Bonkowski, M. & Scheu, S. (2003) Effects of earthworms
789	and organic litter distribution on plant performance and aphid reproduction. Oecologia,
790	137, 90–96.
791	Wurst, S., Langel, R., Rodger, S. & Scheu, S. (2006) Effects of belowground biota on primary and
792	secondary metabolites in <i>Brassica oleracea</i> . Chemoecology, 16 , 69–73.
793	Xiao Z., Wang X., Koricheva J., Kergunteuil A., Le Bayon R., Liu M., Hu F. & Rasmann S. (2017)
794	Data from: Earthworms affect plant growth and resistance against herbivores: a
795	meta-analysis. Dryad Digital Repository. http://doi.org/10.5061/dryad.382k1
796	Yoshitake, S., Soutome, H. & Koizumi, H. (2014) Deposition and decomposition of cattle dung
797	and its impact on soil properties and plant growth in a cool-temperate pasture. Ecological
798	Research, 29, 673–684.
799	Züst, T. & Agrawal, A.A. (2016) Mechanisms and evolution of plant resistance to aphids. <i>Nature</i>
800	Plants, 2, 15206.
801	Züst, T. & Agrawal, A.A. (2017) Trade-offs between plant growth and defense against insect
802	herbivory: an emerging mechanistic synthesis. Annual Review of Plant Biology. 68,
803	10.11–10.22.
804	
805	Supporting Information
806	Additional supporting information may be found in the online version of this article.
807	Supporting Appendix S1: Database and moderator variable
808	Supporting Tables S1-S8: Summary tables for overall effect size, moderator and
809	correlation analyses

810 Supporting Figures S1-S3: additional datapoints, moderator and hierarchical analyses 811 812 Please note: Wiley Blackwell are not responsible for the content or functionality of 813 any supporting information supplied by the authors. Any queries (other than missing 814 material) should be directed to the corresponding author for the article. 815 816 Figure legends 817 Fig. 1 The effect size of earthworms on plant growth (a) and nutrients (b) in the 818 presence of herbivores. Error bars denote 95% bias-corrected confidence intervals 819 (CIs). Sample sizes are shown in brackets. The individual effect is significant if the 95% 820 CIs does not include zero. 821 822 Fig. 2 Moderator analyses of the effect size of earthworm presence on plant resistance 823 against (a) cell-feeders, (b) chewing herbivore and (c) phloem-feeders. Error bars 824 denote 95% bias-corrected confidence intervals (CIs). Sample sizes are shown in 825 brackets. The individual effect is significant if the 95% CIs does not include zero. 826 827 Fig. 3 The effect size of earthworms on plant chemical defences in the presence of 828 different herbivores. Error bars denote 95% bias-corrected confidence intervals (CIs). 829 Sample sizes are shown in brackets. The individual effect is significant if the 95% CIs 830 does not include zero.

Fig. 4 Correlations between the effects of earthworms on (a) plant resistance and growth, (b) chemical defences and growth. Each data point of the correlation corresponded to an lnR. For instance, a significant negative correlation between growth and resistance means that positive effect of earthworm on growth is correlated with negative effect of earthworms on plant resistance, vice versa.

838 Figures:

Figure 1.

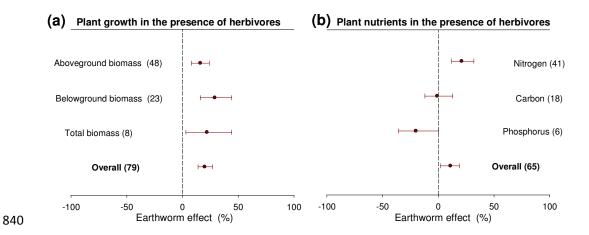


Figure 2. 842

(a) Plant resistance against cell-feeding herbivores (C) Plant resistance against phloem-feeding herbivores Earthworm species richness Cell-feeding groups Single species (25) Nematodes (5) Thrips (3) Multi-species (22) ── Overall (8) Earthworm ecological type Epigeic (9) 0 1 Earthworm effect (%) Earthworm effect (%) (b) Plant resistance against chewing herbivores Single species (5) Multi-species (4) Anecic (5) Mixture (4) Overall (9) -100 -50 0 50 100 Mixture (9) ⊢ Earthworm density (ind./m² soil) < 100 (3) 100-199 (14) 200-400 (15) >400 (15) Overall (47)

100

-100

0 Earthworm effect (%)

0 50 Earthworm effect (%)

-100

843

Figure 3.

Plant chemical defences in the presence of different herbivores

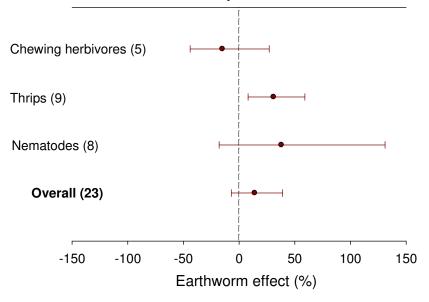


Figure 4.

