Elsevier Editorial System(tm) for Current

Opinion in Virology

Manuscript Draft

Manuscript Number: COVIRO-D-16-00038R1

Title: Impact of managed honey bee viruses on wild bees

Article Type: 19 Environmental virology 2016

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Order of Authors: Anja Tehel, MSc; Mark J Brown, PhD; Robert Paxton

Abstract: Several viruses found in the Western honey bee (Apis mellifera) have recently been detected in other bee species, raising the possibility of spill-over from managed to wild bee species. Alternatively, these viruses may be shared generalists across flower-visiting insects. Here we explore the former hypothesis, pointing out weaknesses in the current evidence, particularly in relation to deformed wing virus (DWV), and highlighting research areas that may help test it. Data so far suggest that DWV spills over from managed to wild bee species and has the potential to cause population decline. That DWV and other viruses of A. mellifera are found in other bee species needs to be considered for the sustainable management of bee populations.



## MARTIN-LUTHER-UNIVERSITÄT HALLE-WITTENBERG

Institut für Biologie Allgemeine Zoologie Prof. Dr. Robert J. Paxton

10. June 2016

June 10<sup>th</sup>, 2016

Dear Editors,

We submit our revised manuscript, entitled:

#### Impact of managed honey bee viruses on wild bees (COVIRO-D-16-00038)

for consideration for publication in Current Opinion in Virology, as one of the invited papers of the special issue on Environmental Virology. We have now revised the ms in response to reviewer comments and attach our point-by-point response to those comments as well as a version of the ms with tracked changes.

All coauthors (Anja Tehel, Mark Brown and myself) have contributed to the ms. The work described in our ms has not been published previously, it is not under consideration for publication elsewhere, its publication is approved by all authors and, if accepted, it will not be published elsewhere in the same form, in English or in any other language, including electronically. We declare no actual or potential conflict of interest including any financial, personal or other relationships with other people or organizations within three years of beginning the submitted work that could inappropriately influence, or be perceived to influence, the manuscript.

Yours faithfully,

Robert Paxton

Hausanschrift: Hoher Weg 8 06120 Halle (Saale) Prof. Dr. Robert J. Paxton Tel: (0345) 5526500 e-mail: robert.paxton@zoologie.uni-halle.de |Fax: (0345) 5527428 June 10<sup>th</sup>, 2016

Dear Editor,

Thank you for the encouraging review of our ms:

# **Impact of managed honey bee viruses on wild bees** (manuscript number COVIRO-D-16-00038)

We have attended to all of the points raised by yourself, both the specific points listed below and on the annotated ms, and upload a revised version of the ms which takes them all into consideration. Below we detail point-by-point our responses to the specific comments.

Please consider the re-submitted, revised version of our manuscript for publication in Current Opinion in Virology.

Yours sincerely,

Robert Paxton (and on behalf of coauthors Anja Tehel and Mark Brown)

Editor's comments and our responses:

#### Comment:

Thank you for an interesting and well written paper on how domestic bee viruses may be impacting wild bees. I just have a few editorial suggestions, some are listed below and some are marked directly on the manuscript.

#### Response:

Thank you for your supporting comments on the ms.

#### **Comment:**

Could you mention something in the introduction about the importance of wild bees in pollination of crops and wild plants?

### **Response:**

We have now stated that wild bees make an important ocpntribution to crop pollination on lines 41-42: Wild bee species also make a major contribution to crop pollination [6].

The relationship between wild bee species and wild plant pollination has not been so clearly defined, though undoubtedly wild bee species are important for the pollination of wild plants. We now add at lines 44-46: Wild bee species are also in decline in northern temperate regions of the world concomitant with a decline in the wild flowers they visit and pollinate [10

#### Comment:

I found the bee nomenclature quite confusing. Please choose a way to describe the bees and then use it throughout. I think defining Apis mellifera

as the Western honey bee is fine, but don't then switch back and forth, just use this designation. I corrected the first few, but please go through and correct the remainder.

#### **Response:**

We have gone through the entire ms and homogenized Apis mellifera to Western honev bee', given specific names to other honev bees (e.g. Eastern honey bee') and made consistent our naming of other bee species. Where we used 'wild bees', we now use 'wild bee species' to make is crystal clear that we do not mean 'Apis mellifera'. We thought confusion might also arise because we sometimes used 'domestic' and sometimes 'managed' to describe the Western honey bee. There is some discussion over whether Apis mellifera is 'domesticated' or merely a wild species now kept in boxes i.e. managed. We therefore replaced 'domesticated' with 'managed' throughout, including in the title, so as to avoid confusion and so as to be consistent throughout the ms.

### Comment:

Please use \* for highlighting references, rather than \*

#### **Response:**

We copied the style of other articles in Current Opinion in Virology and used a bold '\*'. We suspect there may be formatting problems when sending text via the journal's website because we could not differentiate between the two stars used above.

#### Comment:

We usually aim for around 50 references. Would it be possible to use a few reviews to replace some of the primary references, especially for areas that are indirectly related to the topic of the paper, like EIDs in other animals?

#### **Response:**

We have cut the number of references from 79 to 59 by removing many of the primary references, as you suggested, and we have reduced the number of highlighted references to 6.

#### **Comment:**

Please correct the table as noted on the manuscript.

#### **Response:**

We have given the table a short title and placed the explanation to it as a footnote.

Additional comments scored directly on the pdf by the editor.

### Comment:

Graphical Abstract: can you please define these families? Most people will have no idea what they are.

#### **Response:**

We have added additional text to the graphical abstract to make clear the bee taxa and added names in common usage as well as genus and family names of the taxa.

#### Comment:

Please add something here to define BEEHAVE.

#### **Response:**

On lines 158-9 we added the following text to define BEEHAVE: The BEEHAVE model, a software that simulates the development of a honey bee colony and its foraging for nectar and pollen in a realistic landscape,

#### **Comment:**

Figure 1 caption: Please define the other species here, most readers won't know what they are.

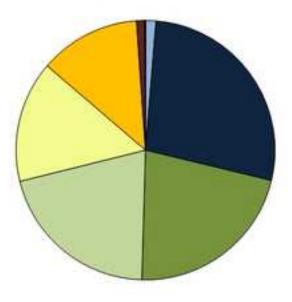
#### **Response:**

We updated the caption to Figure 1 to define the other species more thoroughly.

# Many additional, minor comments were added to the text Response:

We incorporated all the suggested changes into the text.

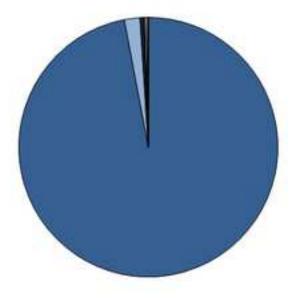
## Number of bee species per taxon



## Bee taxa (genus; family)

- honey bees (Apis; Apidae)
- bumble bees (Bombus; Apidae)
- other apid bees (Apidae)
- sweat bees (Halictidae)
- leafcutter and mason bees (Megachilidae)
- mining bees (Andrenidae)
- silk bees (Colletidae)
- oil bees (Melittidae)
- stenotritid bees (Stenotritidae)

## Number of publications on viruses per bee taxon



## Highlights

- Over 24 viruses have been associated with managed honey bees
- Many of these are also prevalent in wild bee species
- Evidence suggests spill-over of viruses from honey bees to wild bee species
- The impact of these viruses on wild bee species is poorly known

1	1	Impact of managed honey bee viruses on wild bees
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4 5 6	3	Anja Tehel <sup>1,2</sup> • Mark J.F. Brown <sup>3</sup> • Robert J. Paxton <sup>1,2</sup> *
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20	Abstract
21	Several viruses found in the Western honey bee (Apis mellifera) have recently been detected in
22	other bee species, raising the possibility of spill-over from managed to wild bee species.
23	Alternatively, these viruses may be shared generalists across flower-visiting insects. Here we explore
24	the former hypothesis, pointing out weaknesses in the current evidence, particularly in relation to
25	deformed wing virus (DWV), and highlighting research areas that may help test it. Data so far
26	suggest that DWV spills over from managed to wild bee species and has the potential to cause
27	population decline. That DWV and other viruses of A. mellifera are found in other bee species needs
28	to be considered for the sustainable management of bee populations.
29	
30	Keywords:
31	Hymenoptera, Apis mellifera, bumble bee, spill-over, Deformed wing virus
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We, our domestic animals, and wildlife are increasing challenged by a range of emerging infectious diseases (EIDs [1, 2]) that threaten respectively our health, our farming and the ecosystem services we derive from the natural world. After cattle and pigs and before poultry, the Western honey bee (Apis mellifera) has been considered the third most important domestic animal [3, 4]; pollination by insects, including by the Western honey bee, is an important ecosystem service that contributes over US \$ 200 billion to global agricultural output at today's prices and, directly and indirectly, to one in every three mouthfuls of food [5]. Wild bee species also make a major contribution to crop pollination [6]. Elevated losses of Western honey bees have been reported across northern temperate regions of the world over the past decade [7], with EIDs considered a primary cause of mortality [8], possibly because of increased viral virulence [9]. Wild bee species are also in decline in northern temperate regions of the world concomitant with a decline in the wild flowers they visit and pollinate [10], though causes of population decline, range contraction and wild bee species loss are diverse [11, 12]. Given the importance of bees for pollination, these are serious concerns that could affect food security and the functioning of terrestrial ecosystems [13].

To date, 24 viruses have been associated with the Western honey bee [14], many of which are positive sense single strand (+) RNA viruses. One of these in particular, Deformed wing virus (DWV), has risen to prominence because of its association with the honey bee exotic ectoparasitic mite Varroa destructor [15\*], a possibly synergistic association for virus and mite [16]. Varroa destructor (henceforth varroa mite) was originally an ectoparasite of the Asiatic Eastern honey bee (Apis cerana), but jumped to the Western honey bee and is nowadays widespread across most of the world with its new host, the Western honey bee, on which it vectors DWV and many other viruses [8, 14]. Introduction of varroa mites to a naïve population of Western honey bees leads to a

tremendous rise in DWV prevalence and titer per individual host [17, 18], and DWV is the leading
contender as the causal agent of colony decline [14].

Prominent among EIDs of a diverse range of animals are RNA viruses that exhibit high mutation (and substitution) rates [19] and cross species barriers to emerge (i.e. become more prevalent) in novel hosts, including humans [20] e.g. Ebola and Zika. DWV and other (+) RNA viruses of the Western honey bee have now been detected in wild bee species [21\*\*], though knowledge of their impact on wild bee hosts remains limited (Fig. 1; Table 1). That a virus is found across two or more host species, be they managed or wild bees, is important because it dictates the epidemiology and evolutionary trajectory of that pathogen [22, 23\*]. Here we bring together recent literature to explore the hypothesis that viral spill-over from managed Western honey bees negatively impacts wild bee species.

- 71 DWV prevalence and impact

Before the introduction of the varroa mite to the Western honey bee in Europe *ca*. half a century ago, serological (antibody-based) detection of DWV suggested it was at extremely low prevalence [24]. Varroa mites are now found in most, if not all, Western honey bee colonies across all of Europe (except isolated offshore islands and in extreme northern latitudes) and, using PCR-based screening, DWV is the most commonly detected virus, often found at high titer [25]. Careful screening of Western honey bee colonies on the archipelagos of Hawaii and New Zealand has clearly demonstrated that DWV rose to 100% prevalence of colonies after the introduction of varroa mites [17, 18]. It is arguably present in all Western honey bee colonies that harbor varroa mites, and phylogenetic analysis of viral gene sequences suggest it has likely followed the introduction of varroa mites to North America and elsewhere to become an EID of Western honey bees worldwide [15\*].

Australia remains the only large land mass that, to date, has not reported varroa mites in its
managed Western honey bee colonies.

Concomitant with its rise in prevalence across Western honey bee colonies and in titer in individual honey bees, DWV is the most commonly reported virus detected by PCR in a range of other bee species (Fig. 2). It has been found in three Asiatic honey bee (Apis) species in Asia [25, 26]. Across Europe and North America, DWV has also been detected in bumble bees (Bombus spp.) [27\*\*, 28, 29, 30, 31\*\*, 32, 33, 34], other wild bee species [27\*\*, 29, 35, 36, 37] and even taxonomically more distant hosts such as ants, wasps and cockroaches [28, 29, 38]. These represent an eclectic mix of host species, and the field would benefit from more systematic surveying of the prevalence of DWV, both in terms of geographic and taxonomic (host species) coverage.

Whether these increases in host range of DWV beyond Western honey bees are a consequence of
pathogen spill-over of this EID of the Western honey bee are equivocal because most studies merely
report PCR-based detection of DWV. For the Asiatic honey bees that also host varroa mites,
phylogenetic analysis of DWV isolates from these different host species supports the suggestion of
EID spill-over from introduced, managed Western honey bees harboring varroa mites and DWV [25].
Given the mechanistic link between varroa mites and DWV transmission, EID spill-over into Asiatic
honey bee species is a plausible interpretation of the phylogenetic data.

As only honey bees (*Apis* spp.) host varroa mites that can vector DWV, it is assumed that bumble
bees and other wild bee species acquire DWV though robbing of DWV-infected colonies of Western
honey bees for honey or, more likely, by collection and ingestion of DWV with pollen or nectar from
flowers that are also visited by DWV-infected Western honey bee workers [27\*\*, 39, 40]. Given this
assumption, we highlight two issues that confound the interpretation of data on DWV prevalence in
wild bee species in terms of impact on hosts.

#### Limitations to interpretation of data on DWV impact

The first issue is that many studies of wild bee species merely report the detection (or not) of DWV, often from a single site [36], making inference of DWV spill-over from Western honey bees difficult at best. We know little of the parasite community of wild bee species [41], a research area that deserves greater attention. Therefore, DWV as a natural pathogen of any given wild bee species cannot in principle be ruled out; it is a plausible hypothesis that DWV is a generalist insect pathogen, or one originally associated with a species other than the Western honey bee. 

To infer spill-over, studies need to examine the prevalence of DWV in managed Western honey bees and wild bee species across multiple sites, and couple this with proof that the pathogen in managed Western honey bees and wild bee species is the same, e.g., by analysis of genetic variants of DWV [27\*\*, 29]. The only study of which we are aware that currently fulfills both criteria has demonstrated (i) a clear correlation between DWV prevalence in Western honey bees and in wild bumble bees across multiple sites and (ii) sharing of the identical viral variant by Western honey bees and wild bees (Bombus spp.) within a site [31\*\*]. 

These data speak strongly for pathogen sharing between Western honey bees and wild bee species but still leave open the question of directionality of pathogen transmission. DWV prevalence in Western honey bees collected from flowers was approximately double that of DWV prevalence in bumble bees, strongly suggesting spill-over from Western honey bees to wild bee species [31\*\*]. Yet **131** interpretation of pathogen prevalence data is potentially conflated with virulence of that pathogen in a host; for insect populations, mathematical models suggest that highly virulent pathogens exhibit low prevalence and vice versa [42]. There is a clear need for additional research focused on understanding the directionality of transmission of DWV (and other pathogens) among host species,

especially research integrating phylogenetic methods that infer transmission over long time scales [15\*] with experimental approaches that can test for on-going transmission [27\*\*].

The second issue concerning PCR-based detection of DWV in wild bee species and other insects is that it may merely reflect contamination of the exterior of a wild bee by DWV or, in the case of wasps, ants and cockroaches, consumption of DWV-infected Western honey bees or hive products. Though such contamination might also facilitate the dispersal of DWV [28, 43], it does not necessarily represent a threat to the host, either the individual host insect, or the host colony (for social species such as bumble bees) or the host population. Amplification of the negative (-) viral strand of (+) RNA viruses like DWV can be used as an indicator of active infection [31\*\*], though we note that in principle it demonstrates viral replication and not host pathology. Similarly, the detection of DWV-infected carpenter bees (Xylocopa augusti) with deformed wings suggests that DWV can be infective and pathogenic for wild bee species [35], although other parasites can cause similar pathologies [44]. The most direct means to determine virulence is through experimental inoculation of a host species. Though laboratory experimentation is an abstraction of the real world, it controls for conflating factors when testing for virulence. In experimental inoculation assays, DWV has been shown to be a pathogen of the widespread European bumble bee Bombus terrestris [31\*\*, 33].

We have little idea of how virulent DWV is in the other wild bee species in which it has been detected. This is clearly an important area for future research if we are to understand the impact of DWV spill-over from managed Western honey bees to wild bee species. For social wild bee species, there is also a need to scale up from virulence at the level of the individual insect to virulence at the level of the colony [45] and the population. The BEEHAVE model, a software that simulates the development of a honey bee colony and its foraging for nectar and pollen in a realistic landscape,

can fulfill this propose for the Western honey bee [46], but there is a need for mathematical models incorporating other forms of social organization, such as that of the bumble bee [47].

#### Other viruses, their prevalence and impact

Ten other viruses originally isolated from the Western honey bee, mostly (+) RNA viruses, have been detected in wild bee species (Table 1). After DWV, the next most commonly reported virus in wild bee species is Black queen cell virus (BQCV), followed by Israeli acute paralysis virus (IAPV) (Fig. 2), both (+) RNA viruses.

BQCV is prevalent and widespread across managed Western honey bee populations in Europe [9, 48] and North America [49]. A structured survey of 26 sites in Great Britain showed BQCV to be more common in the Western honey bee than in bumble bee species, and prevalence of all viruses in Western honey bees was a significant predictor of viral prevalence in bumble bees [21\*\*]. These data again suggest spill-over from domestic Western honey bees to wild bee species (but see caveats above). Though BQCV is pathogenic for Western honey bee larvae following experimental inoculation [50], its impact on wild bee species is unknown, but warrants investigation.

IAPV has been rarely reported from Europe, but in North America it is prevalent in managed 45 179 Western honey bees and was initially associated with so-called colony collapse disorder [51]. Like the closely related Kashmir bee virus (KBV) and Acute bee paralysis virus (ABPV), IAPV is highly pathogenic for Western honey bees [52]. It has been detected in wild bee species near IAPV-infected **182** managed Western honey bee colonies, but not in the same wild bee species near a non-IAPVinfected apiary [27\*\*]. Furthermore, greenhouse experiments [27\*\*] have shown successful transmission of IAPV between Western honey bees and wild bee species. Spill-over of IAPV from Western honey bees to wild bee species in the field through shared use of flowers is therefore likely. 

We note, though, that the related ABPV, also highly pathogenic for Western honey bees [53], has a higher prevalence in wild bee species than Western honey bees [21\*\*], suggesting spill-over from wild bee species to domestic Western honey bees (but see caveats above).

Though studies on BQCV and IAPV in wild bee species are few (Fig. 2), they highlight the potentially wide distribution of viruses across multiple bee species, likely promoted by shared use of flowers. Experimental demonstration of IAPV cross-species transmission [27\*\*] is exemplary. Similar experiments with other viruses would help our understanding of the threat posed to wild bee species by viruses of managed Western honey bees.

#### **Future directions**

Evidence is mounting that viruses originally detected in the managed Western honey bee are widely distributed across wild bee species. For DWV, data support the idea that virus spills over from Western honey bees to wild bee species, where it potentially reduces individual wild bee host fitness [31\*\*]. Yet viral prevalence varies considerably across wild bee species [21\*\*], and some viruses have higher prevalence in wild bees, suggesting spill-over to managed Western honey bees. A key future aim is to determine the epidemiology and transmission dynamics of these viruses in the field. Do they show source-sink dynamics, or active back-and-forth transmission? Understanding this will give us tools to manipulate transmission in the field, for the benefit of both wild bee species and managed Western honey bees. From the direct perspective of impact, an interesting question is whether some wild bee species, particularly those that are rare or declining, are less tolerant of viral **208** pathogens than others; analysis of their anti-viral innate immune defence mechanisms [54], whose complements vary among Dipteran species [55\*], may be one productive means of investigation. Similarly, genetic analyses of population size and bottlenecks in coupled wild bee-virus systems might elucidate past impacts. Whether viral emergence in managed Western honey bees leads to

host switching through the mass action principle [56, 57], i.e. through increasing density of infective virions in the environment, versus viral adaptation is another question that would benefit from greater attention, not only in understanding the threat posed to wild bee species by managed Western honey bee viruses but also as a model of EID impacts on wildlife. The impact of viruses may also vary with co-infection by two or more viruses [58] and through interactions with other so-called 'stressors' of wild bee species such as pesticide exposure [8], as has been demonstrated at the molecular level for the Western honey bee [59]. With such information to hand, we may be able to understand how pathogen spill-over from Western honey bees impacts communities of wild bee species, their populations, and the ecosystem service of pollination, from which terrestrial life and human well-being benefit. Acknowledgements We thank the editor for helpful comments on the manuscript. For funding, AT thanks the DBU, MJFB thanks BB/N000668/1, and RJP thanks the DFG's Priority Programme 1596 (Ecology and species barriers in emerging viral diseases), project Pa 632/11. **References and recommended reading** Papers of particular interest, published within the period of review, have been highlighted as: \* of special interest \*\* of outstanding interest 1. Daszak P, Cunningham AA, Hyatt AD: Emerging infectious diseases of wildlife - threats to biodiversity and human health. Science 2000, 287:443-449. 2. Woolhouse ME, Haydon DT, Antia R: Emerging pathogens: the epidemiology and evolution of species jumps. Trends Ecol Evol 2005, 20:238-44. 3. Tautz J: The Buzz about Bees: Biology of a Superorganism. Springer; 2008. 4. Aizen MA, Harder LD: The global stock of domesticated honey bees is growing slower than agricultural demand for pollination. Curr Biol 2009, 19:915-8. 5. Klein AM, Vaissiere BE, Cane JH, Steffan-Dewenter I, Cunningham SA, Kremen C, Tscharntke T: Importance of pollinators in changing landscapes for world crops. P Roy Soc Lond B 2007, 274:303-313. 

-	241	6. Garibaldi LA, Carvalheiro LG, Vaissière BE, Gemmill-Herren B, Hipólito J, Freitas BM, Ngo HT,
1 2	242	Azzu N, Sáez A, Åström J <i>et al</i> .: Mutually beneficial pollinator diversity and crop yield
3	243	outcomes in small and large farms. Science 2016, 351:388-91.
4	244	7. Neumann P, Carreck NL: Honey bee colony losses. J Apic Res 2010, 49:1-6.
5	245	8. McMenamin AJ, Brutscher LM, Glenny W, Flenniken ML: Abiotic and biotic factors affecting
6 7	246	the replication and pathogenicity of bee viruses. Curr Opin Insect Sci 2016, 16:14-21.
8	247	9. McMahon DP, Natsopoulou M, Doublet V, Fürst MA, Weging S, Brown MJF, Gogol-Döring A,
9	248	Paxton RJ: Elevated virulence of an emerging viral genotype as a driver of honeybee loss.
10 11	249	Proc R Soc Biol Sci Ser B in press.
$12^{11}$	250	10. Biesmeijer JC, Roberts SP, Reemer M, Ohlemüller R, Edwards M, Peeters T, Schaffers AP,
13	251	Potts SG, Kleukers R, Thomas CD <i>et al.</i> : <b>Parallel declines in pollinators and insect-pollinated</b>
14 15	251	plants in Britain and the Netherlands. Science 2006, <b>313</b> :351-4.
15 16	252	11. Vanbergen AJ, the Insect Pollinators Initiative: <b>Threats to an ecosystem service: pressures</b>
17		-
18	254	on pollinators. Front Ecol Environ 2013, <b>11:</b> 251-259.
19 20	255 256	12. Goulson D, Nicholls E, Botías C, Rotheray EL: <b>Bee declines driven by combined stress from</b>
21		parasites, pesticides, and lack of flowers. Science 2015, <b>347</b> :1255957.
22	257	13. Potts SG, Biesmeijer JC, Kremen C, Neumann P, Schweiger O, Kunin WE: Global pollinator
23 24	258	declines: trends, impacts and drivers. Trends Ecol Evol 2010, 25:345-53.
25	259	14. McMenamin AJ, Genersch E 2015: Honey bee colony losses and associated viruses. Curr
26	260	<i>Opin Insect Sci</i> 2015, <b>8:</b> 121-129.
27 28	261	15. * Wilfert L, Long G, Leggett HC, Schmid-Hempel P, Butlin R, Martin SJM Boots M: Deformed
28 29	262	wing virus is a recent global epidemic in honeybees driven by Varroa mites. Science 2016,
30	263	<b>351:</b> 594-597.
31		
32 33	264	This study applies Bayesian statistical methods to the analysis of three gene sequences of DWV
34	265	from isolates collected across the world. There is strong support for the emergence of DWV in
35	266	managed Western honey bees (Apis mellifera) as a consequence of varroa mite (Varroa
36 37	267	destructor) parasitism of honey bees, and its likely spread from Europe to North America and
38	268	elsewhere.
39 40	269	16. Di Prisco G, Annoscia D, Margiotta M, Ferrara R, Varricchio P, Zanni V, Caprio E, Nazzi F,
40 41	205	Pennacchio F: A mutualistic symbiosis between a parasitic mite and a pathogenic virus
42		
43	271	
1 1	271 272	undermines honey bee immunity and health. PNAS 2016, 113: 3203-3208.
44 45	272	undermines honey bee immunity and health. PNAS 2016, <b>113:</b> 3203-3208. 17. Martin SJ, Highfield AC, Brettell L, Villalobos EM, Budge GE, Powell M, Nikaido S, Schroeder
44 45 46	272 273	<ul> <li>undermines honey bee immunity and health. PNAS 2016, 113: 3203-3208.</li> <li>17. Martin SJ, Highfield AC, Brettell L, Villalobos EM, Budge GE, Powell M, Nikaido S, Schroeder DC: Global honey bee viral landscape altered by a parasitic mite. Science 2012, 336:1304-</li> </ul>
45 46 47	272 273 274	<ul> <li>undermines honey bee immunity and health. PNAS 2016, 113: 3203-3208.</li> <li>17. Martin SJ, Highfield AC, Brettell L, Villalobos EM, Budge GE, Powell M, Nikaido S, Schroeder DC: Global honey bee viral landscape altered by a parasitic mite. Science 2012, 336:1304-1306.</li> </ul>
45 46 47 48	272 273 274 275	<ul> <li>undermines honey bee immunity and health. PNAS 2016, 113: 3203-3208.</li> <li>17. Martin SJ, Highfield AC, Brettell L, Villalobos EM, Budge GE, Powell M, Nikaido S, Schroeder DC: Global honey bee viral landscape altered by a parasitic mite. Science 2012, 336:1304-1306.</li> <li>18. Mondet F, de Miranda JR, Kretzschmar A, Le Conte Y, Mercer AR: Colonies along a New</li> </ul>
45 46 47	272 273 274 275 276	<ul> <li>undermines honey bee immunity and health. PNAS 2016, 113: 3203-3208.</li> <li>17. Martin SJ, Highfield AC, Brettell L, Villalobos EM, Budge GE, Powell M, Nikaido S, Schroeder DC: Global honey bee viral landscape altered by a parasitic mite. Science 2012, 336:1304-1306.</li> <li>18. Mondet F, de Miranda JR, Kretzschmar A, Le Conte Y, Mercer AR: Colonies along a New Expansion front of the parasite Varroa destructor. PLoS Pathog 2014, 10:e1004323.</li> </ul>
45 46 47 48 49 50 51	272 273 274 275 276 277	<ul> <li>undermines honey bee immunity and health. PNAS 2016, 113: 3203-3208.</li> <li>17. Martin SJ, Highfield AC, Brettell L, Villalobos EM, Budge GE, Powell M, Nikaido S, Schroeder DC: Global honey bee viral landscape altered by a parasitic mite. Science 2012, 336:1304-1306.</li> <li>18. Mondet F, de Miranda JR, Kretzschmar A, Le Conte Y, Mercer AR: Colonies along a New Expansion front of the parasite Varroa destructor. PLoS Pathog 2014, 10:e1004323.</li> <li>19. Moya A. Holmes EC, Gonzalez-Candelas F: The population genetics and evolutionary</li> </ul>
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	284	This study examines the prevalence and viral load of multiple putative Western honey bee (Apis
1	285	mellifera) viruses in wild bumble bees (Bombus spp.) in a taxonomically and geographically well
2 3	286	replicated and structured sampling design. It re-affirms results of [31**] for DWV and shows that
4	287	multiple other 'honey bee' viruses are present in bumble bees, some at high titer per bumble bee.
5		······································
б	288	22. Brown MJF: Unravelling the complexities of host-parasite interactions. Bee World 2013,
7 8	289	<b>90:</b> 36-37.
8 9	290	23. * Manley R, Boots M, Wilfert L: Emerging viral disease risk to pollinating insects: ecological,
10	291	evolutionary and anthropogenic factors. J App Ecol 2015, <b>52</b> :331-340.
11		
12 13	292	This is a thorough and up-to-date review of the risks posed by viruses of managed bees for wild
14	293	bees and other flower-visiting insects.
15		
16	294	24. Bailey L, Ball BV, Perry JN: The prevalence of viruses of honey bees in Britain. Ann Appl Biol
17 18	295	1981, <b>97:</b> 109-118.
19	296	25. Zhang X, He SY, Evans JD, Pettis SE, Yin GF, Chen YP: New evidence that Deformed wing
20	297	virus and Black queen cell virus are multi-host pathogens. J Invertebr Pathol 2012, 109:156-
21 22	298	159.
22	299	26. Forsgren E, Wei S, Guiling D, Zhiguang L, Tran TV, Tang PT, Truong TA, Dinh TQ, Fries I:
24	300	Preliminary observations on possible pathogen spill-over from Apis mellifera to Apis
25	301	cerana. Apidologie 2015, <b>46:</b> 265-275.
26 27	302	27. ** Singh R, Levitt AL, Rajotte EG, Holmes EC, Ostiguy N, vanEngelsdorp S, Lipkin WI,
28	303	
29		dePamphillis CW, Toth AL, Cox-Foster DL: <b>RNA viruses in hymenopteran pollinators:</b>
30	304	Evidence of inter-taxa virus transmission via pollen and potential impact on non- <i>Apis</i>
31 32	305	hymenopteran species. PLoS ONE 2010, 5:e14357.
33	306	This is a significant study because it employed several approaches to test for the transmission of
34	307	numerous viruses within and between bee species. Most notably, in a semi-field experimental
35 36	308	setting, IAPV was shown to be transferred from managed Western honey bees ( <i>Apis mellifera</i> ) to
37		
38	309	bumble bees (Bombus sp.) and vice versa through shared use of flowers.
39 40	310	28. Evison SEF, Roberts KE, Laurenson L, Pietravalle S, Hui J, Biesmeijer JC, Smith JE, Budge G,
40 41	311	Hughes WHO: <b>Pervasiveness of parasites in pollinators.</b> <i>PLoS ONE</i> 2012, <b>7</b> :e30641.
42	312	29. Levitt AL, Singh R, Cox-Foster DL, Rajotte E, Hoover K, Ostiguy N, Holmes EC: <b>Cross-species</b>
43	313	transmission of honey bee viruses in associated arthropods. Virus Res 2013, <b>176</b> :232-240.
44 45	313	30. Reynaldi FJ, Sguazza GH, Albicoro FJ, Pecoraro MR, Galosi CM: First molecular detection of
46		
47	315	co-infection of honey bee viruses in asymptomatic <i>Bombus atratus</i> in South America. <i>Braz</i>
48 49	316	<i>J Biol.</i> 2013, <b>73</b> :797-800.
49 50	317	31. ** Fürst M A, McMahon DP, Osborne JL, Paxton RJ, Brown MJF: Disease associations
51	318	between honeybees and bumblebees as a threat to wild pollinators. <i>Nature</i> 2014, 506:364-
52	319	366.
53 54	220	and the second
55	320	Using a highly replicated and structured sampling design, this study provided strong evidence for
56	321	DWV transmission in the field between managed Western honey bees ( <i>Apis mellifera</i> ) and wild
57 50	322	bumble bees (Bombus spp.). Sequencing of DWV variants gave unequivocal support for the on-
58 59	323	going sharing of DWV between bee taxa while lab experiments demonstrated the pathogenicity of
60	324	DWV for wild bumble bees when ingested with pollen.
61		
62 63		
63 64		12
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	325	32.	Gamboa V, Ravoet J, Brunain M, Smagghe G, Meeus I, Figueroa J, Riano D, de Graaf DC: Bee
1	326		pathogens found in Bombus atratus from Colombia: A case study. J Invertebr Pathol 2015,
2 3	327		<b>129:</b> 36-39.
4	328	33.	Graystock P, Meeus I, Smagghe G, Goulson D, Hughes WHO: The effects of single and mixed
5	329		infections of Apicystis bombi and Deformed wing virus in Bombus terrestris Parasitol
6 7	330		2015b, <b>143:</b> 358-65.
8	331	34.	Sachman-Ruiz B, Narva´ez-Padilla V, Reynaud E: Commercial Bombus impatiens as
9	332		reservoirs of emerging infectious diseases in central Mexico. Biol Invasions 2015, 17:2043-
10 11	333		2053.
12	334	35.	Lucia M, Reynaldi FJ, H Sguazza GH, Abrahamovich AH: First detection of deformed wing
13	335		virus in Xylocopa augusti larvae (Hymenoptera: Apidae) in Argentina. J Apic Res 2015,
14 15	336		<b>53:</b> 466-468.
16	337	36.	Ravoet J, De Smet L, Meeus I, Smagghe G, Wenseleers T, de Graaf DC: Widespread
17	338		occurrence of honey bee pathogens in solitary bees. J Invertebr Pathol 2014, <b>122</b> :55-58.
18 19	339	37	Guzman-Novoa E, Hamiduzzaman MM, Anguiano-Baez R, Correa-Benitez A, Castaneda-
20	340		Cervantes E, Arnold NI: First detection of honey bee viruses in stingless bees in North
21	341		America. J Apic Res 2015, <b>54:</b> 93-95.
22 23	342	38	Sebastien A, Lester PJ, Hall RJ, Wang J, Moore NE, Gruber MAM: Invasive ants carry novel
24	343	50.	viruses in their new range and form reservoirs for a honeybee pathogen. <i>Biol Lett</i> 2015,
25	344		<b>11</b> :20150610.
26 27	345	20	Mazzei M, Carrozza ML, Luisi E, Forzan M, Giusti M, Sagona S, Tolari F, Felicioli A: Infectivity
28	346	55.	of DWV associated to flower pollen: experimental evidence of a horizontal transmission
29	340 347		route. PLoS ONE 2014, 9:e113448.
30 31	348	40	McArt SH, Koch H, Irwin RE, Adler LS: Arranging the bouquet of disease: floral traits and the
32	349	40.	transmission of plant and animal pathogens. Ecol Lett 2014, 17:624-636.
33 34	350	/1	Engel P, Kwong WK, McFrederick Q, Anderson KE, Barribeau SM, Chandler JA, Cornman RS,
35	351	41.	Dainat J, de Miranda JR, Doublet V:The Bee Microbiome: <b>Impact on bee health and model</b>
36	352		for evolution and ecology of host-microbe interactions. <i>mBio</i> 2016, <b>7</b> :e02164-15.
37 38		10	
39		42.	Anderson RM, May RM: The population dynamics of microparasites and their invertebrate
40	354 255	10	hosts. Phil Trans R Soc B 1981, <b>291:</b> 451-524.
41 42	355	45.	Graystock P, Goulson D, Hughes WHO: <b>Parasites in bloom: flowers aid dispersal and</b>
43	356		transmission of pollinator parasites within and between bee species. Proc R Soc Lond B
44	357		2015, <b>282</b> :20151371.
45 46	358	44.	Rutrecht ST, Brown MJF: <b>Differential virulence in a multiple-host parasite of bumble bees:</b>
47	359	45	resolving the paradox of parasite survival? <i>Oikos</i> 2009, <b>118</b> :941-949.
48	360	45.	Barron AB: Death of the bee hive: understanding the failure of an insect society. Curr Opin
49 50	361		Insect Sci 2015, <b>10:</b> 45-50.
51	362	46.	Becher MA, Grimm V, Thorbek P, Horn J, Kennedy PJ, Osborne JL: <b>BEEHAVE: a systems</b>
52	363		model of honeybee colony dynamics and foraging to explore multifactorial causes of
53 54	364		colony failure. J Appl Ecol 2014, 51:470-482.
55	365	47.	Bryden J, Gill RJ, Mitton RA, Raine NE, Jansen VA: Chronic sublethal stress causes bee colony
56	366		failure. Ecol Lett 2013, 16:1463-9.
57 58	367	48.	. Tentcheva D, Gauthier L, Zappulla N, Dainat B, Cousserans F, Colin ME, Bergoin M:
58 59	368		Prevalence and seasonal variations of six bee viruses in Apis mellifera L. and Varroa
60	369		destructor mite populations in France. Appl Environ Microbiol 2004, 70:7185-7191.
61 62			
63			10
64			13

	370	49. Desaia SD, Kumara S, Currie RW: Occurrence, detection, and quantification of economically
1	371	important viruses in healthy and unhealthy honey bee (Hymenoptera: Apidae) colonies in
2 3	372	<b>Canada.</b> Behav Ecol 2016, <b>148:</b> 22-35.
4	373	50. Doublet V, Labarussias M, de Miranda JR, Moritz RF, Paxton RJ: Bees under stress: sublethal
5	374	doses of a neonicotinoid pesticide and pathogens interact to elevate honey bee mortality
6 7	375	across the life cycle. Environ Microbiol 2015, 17:969-983.
8	376	51. Cox-Foster DL, Conlan S, Holmes EC, Palacios G, Evans J, Moran NA: A metagenomic survey
9	377	of microbes in honey bee colony collapse disorder. Science 2007, <b>318:</b> 283-287.
10 11	378	52. de Miranda JR, Cordoni G, Budge G: The Acute bee paralysis virus-Kashmir bee virus-Israeli
12	379	acute paralysis virus complex. J Invertebr Pathol 2010, <b>103</b> :S30-47.
13	380	53. Sumpter DJT, Martin SJ: <b>The dynamics of virus epidemics in <i>Varroa</i>-infested honey bee</b>
14 15	381	colonies. J Anim Ecol 2004, 73:51-63.
16	382	54. Brutscher LM, Daughenbaugh KF, Flenniken ML: Antiviral defense mechanisms in honey
17	383	bees. Curr Opin Insect Sci 2015, <b>10</b> :71-82.
18	384	55. * Bronkhorst AW, van Rij RP: <b>The long and short of antiviral defense: small RNA-based</b>
19 20		
21	385	immunity in insects. Curr Opin Virol 2014, 7:19-28.
22	386	This is an excellent introduction to the diversity of RNAi defenses in insects.
23 24		····
25	387	56. Anderson RM, May RM: Population biology of infectious diseases: Part I. Nature 1979,
26	388	<b>280:</b> 361-367.
27 28	389	57. May RM, Anderson RM: Population biology of infectious diseases: Part II. Nature 1979,
29	390	<b>280:</b> 455-461.
30	391	58. Rigaud T, Perrot-Minnot MJ, Brown MJ: Parasite and host assemblages: embracing the
31 32	392	reality will improve our knowledge of parasite transmission and virulence. Proc Biol Sci
33	393	2010, <b>277:</b> 693-702.
34	394	59. Di Prisco G, Cavaliere V, Annoscia D, Varricchio P, Caprio E, Nazzi F, Gargiulo G, Pennacchio F:
35 36	395	Neonicotinoid clothianidin adversely affects insect immunity and promotes replication of a
37	396	viral pathogen in honey bees. PNAS 2013, <b>110:</b> 18466–18471.
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	DWV	BQCV	IAPV	SBPV	ABPV	SBV	KBV	LSV	CBPV	AmFV	VdM
Andrena vaga ª		17	20					17		17	
Andrena ventralis <sup>a</sup>								17		17	
Augochlora pura <sup>b</sup>	20										
Bombus agrorum <sup>c</sup>					1						
Bombus atratus <sup>c</sup>	4, 18	4, 18		4	4	4, 18					
Bombus hortorum <sup>c</sup>		12		12	1, 12						
Bombus huntii <sup>c</sup>	9	15									
Bombus impatiens <sup>c</sup>	9, 19, 20	8, 20	8, 19	8	19		19		19		
Bombus lapidarius <sup>c</sup>	3	12		12, 14	12			14			14
Bombus lucorum <sup>c</sup>	3	12		12	1, 12						
Bombus monticola <sup>c</sup>	3										
Bombus pascuorum <sup>c</sup>	2, 5	12		12	12			14			14
Bombus pratorum <sup>c</sup>								14			
Bombus ruderarius <sup>c</sup>					1						
Bombus ternarius <sup>c</sup>	20	20	20								
Bombus terrestris <sup>c</sup>	2, 3, 5, 6	12	13, 16	12	1, 12	12, 20	13				
Bombus vagans <sup>c</sup>	8, 20	8, 20	20	8		20					
Ceratina dupla <sup>d</sup>	20										
Heriades truncorum <sup>e</sup>		17									
Osmia bicornis <sup>e</sup>	17							17		17	17
Osmia cornuta <sup>e</sup>	11	17						17		17	17
Scaptotrigona mexicana <sup>f</sup>	7	7									
Xylocopa latreille <sup>d</sup>	10										
Xylocopa virginica <sup>d</sup>	20	8, 20		8							1

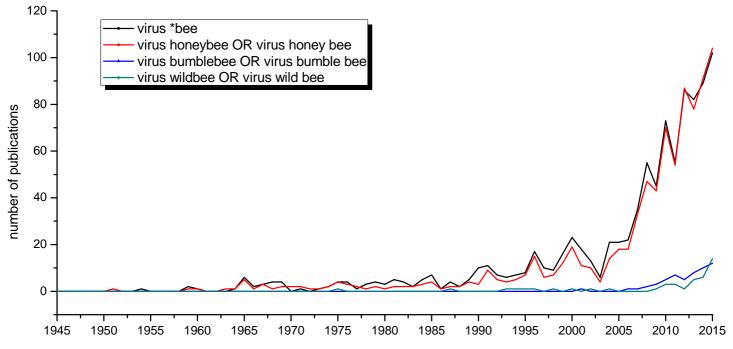
Table 1: Reported taxonomic distribution in wild bee species of viruses initially detected in managed Western honey bees (Apis mellifera)

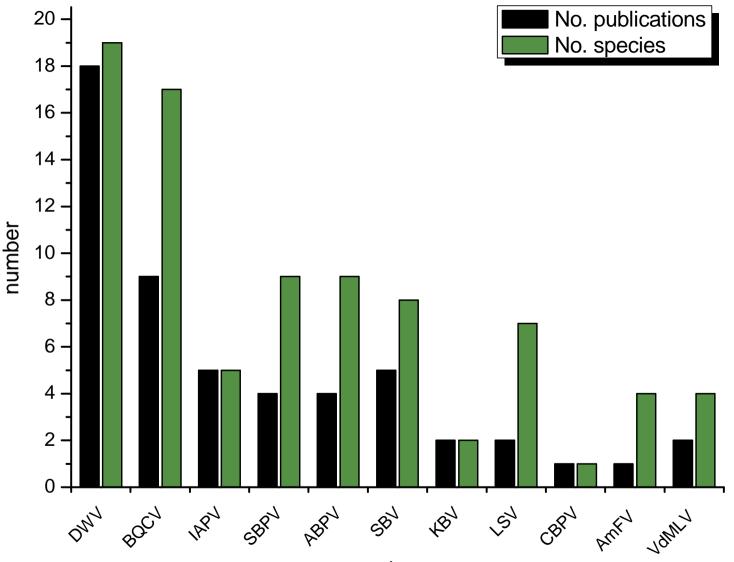
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5 6 <b>400</b>	For numbered references, see supplementary material Appendix A; cell shading in yellow: detection of a virus in a host species; in orange: detection of (-)
7 <b>401</b> 8 <b>402</b>	strand of virus in a host species, indicative of RNA replication; in red: demonstration of pathogenicity in a host species; DWV (Deformed wing virus), BQCV (Black queen cell virus), IAPV (Israeli acute paralysis virus), SBPV (Slow bee paralysis virus), ABPV (Acute bee paralysis virus), SBV (Sacbrood virus), KBV
10 405	(Kashmir bee virus), LSV (Lake Sinai virus), CBPV (Chronic bee paralysis virus), AmFV (Apis mellifera filamentous virus), VdMLV (Varroa destructor macula-
10 <b>404</b>	like virus); <sup>a</sup> mining bees (family Andrenidae); <sup>b</sup> sweat bees (family Halictidae); <sup>c</sup> bumble bees (genus Bombus, family Apidae); <sup>d</sup> carpenter bees (members of
12 <b>405</b>	the family Apidae); <sup>e</sup> leafcutter and mason bees (family Megachilidae); <sup>f</sup> stingless bees (members of the family Apidae)
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**Figure captions:** 

442	Figure 1: Increase in the number of publications on viruses across bee species over the past 70
443	years, showing that most relate to managed honey bees (the Western honey bee, Apis mellifera,
444	or other Asiatic honey bee species that all belong to the genus Apis), and relatively few to
445	bumble bees (genus Bombus) or other wild bees (species in the family Apidae other than honey
446	bees and bumble bees, plus all species in the families: Halictidae, Megachilidae, Andrenidae,
447	Colletidae, Melittidae and Stenotritidae); Web of Science search (accessed on March 18 <sup>th</sup> , 2016)
448	using terms in the box
449	
450	Figure 2: The frequency with which putative honey bee (the Western honey bee, Apis mellifera,
450 451	<b>Figure 2:</b> The frequency with which putative honey bee (the Western honey bee, <i>Apis mellifera</i> , or other Asiatic honey bee species that all belong to the genus <i>Apis</i> ) viruses have been reported
451	or other Asiatic honey bee species that all belong to the genus <i>Apis</i> ) viruses have been reported
451 452	or other Asiatic honey bee species that all belong to the genus <i>Apis</i> ) viruses have been reported in other bee species, both in terms of the total number of papers per virus and the number of
451 452 453	or other Asiatic honey bee species that all belong to the genus <i>Apis</i> ) viruses have been reported in other bee species, both in terms of the total number of papers per virus and the number of host species (excluding <i>Apis</i> spp.) per virus; DWV (Deformed wing virus), BQCV (Black queen cell
451 452 453 454	or other Asiatic honey bee species that all belong to the genus <i>Apis</i> ) viruses have been reported in other bee species, both in terms of the total number of papers per virus and the number of host species (excluding <i>Apis</i> spp.) per virus; DWV (Deformed wing virus), BQCV (Black queen cell virus), IAPV (Israeli acute paralysis virus), SBPV (Slow bee paralysis virus), ABPV (Acute bee

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#### Supplementary Material

#### Appendix A: numbered references embedded within Table 1

- 1. Bailey L, Gibbs AJ: Acute infection of bees with paralysis virus. *J Insect Pathol* 1964, **6**:395-407.
- Evison SEF, Roberts KE, Laurenson L, Pietravalle S, Hui J, Biesmeijer JC, Smith JE, Budge G, Hughes WHO: Pervasiveness of parasites in pollinators. *PLoS ONE* 2012, 7:e30641.
- 3. Fürst M A, McMahon DP, Osborne JL, Paxton RJ, Brown MJF: **Disease associations between honeybees and bumblebees as a threat to wild pollinators.** *Nature* 2014, **506**:364-366.
- 4. Gamboa V, Ravoet J, Brunain M, Smagghe G, Meeus I, Figueroa J, Riano D, de Graaf DC: **Bee pathogens found in** *Bombus atratus* from Colombia: a case study. *J Invertebr Pathol* 2015, **129**:36-39.
- 5. Genersch E, Yue C, Fries I, De Miranda JR: **Detection of Deformed Wing Virus, a honey bee viral pathogen, in bumble bees (***Bombus terrestris* and *Bombus pascuorum***) with wing deformities.** *J Invertebr Pathol* 2006, **91**:61-63.
- 6. Graystock P, Meeus I, Smagghe G, Goulson D, Hughes WHO: **The effects of single** and mixed infections of *Apicystis bombi* and deformed wing virus in *Bombus terrestris. Parasitol* 2015, **143**:358-65.
- 7. Guzman-Novoa E, Hamiduzzaman MM, Anguiano-Baez R, Correa-Benitez A, Castaneda-Cervantes E, Arnold NI: **First detection of honey bee viruses in stingless bees in North America**. *J Apic Res* 2015, **54**:93-95.
- 8. Levitt AL, Singh R, Cox-Foster DL, Rajotte E, Hoover K, Ostiguy N, Holmes EC: **Cross-species transmission of honey bee viruses in associated arthropods**. *Virus Res* 2013, **176**:232-240.
- 9. Li J, Peng W, Wu J, Strange JP, Boncristiani H, Chen Y: **Cross-species infection of deformed wing virus poses a new threat to pollinator conservation.** *J Econ Entomol* 2011, **104**:732-739.
- 10. Lucia M, Reynaldi FJ, H Sguazza GH, Abrahamovich AH: **First detection of deformed wing virus in** *Xylocopa augusti* **larvae (Hymenoptera: Apidae) in Argentina**. J Apic Res 2015, **53:**466-468.
- 11. Mazzei M, Carrozza ML, Luisi E, Forzan M, Giusti M, Sagona S, Tolari F, Felicioli A: Infectivity of DWV associated to flower pollen: experimental evidence of a horizontal transmission route. *PLoS ONE* 2014, **9**:e113448.
- 12. McMahon DP, Fürst MA, Caspar J, Theodorou P, Brown MJF, Paxton RJ: A sting in the spit: widespread cross-infection of multiple RNA viruses across wild and managed bees. *J Anim Ecol* 2015, **84**:615-624.
- 13. Meeus I, de Miranda JR, de Graaf DC, Wäckers F, Smagghe G: **Effect of oral infection with Kashmir bee virus and Israeli acute paralysis virus on bumblebee** *(Bombus terrestris)* **reproductive success**. *J Invertebr Pathol* 2014, **121**:64-69.
- 14. Parmentier L, Smagghe G, de Graaf DC, Meeus I: **Varroa destructor Macula-like virus, Lake Sinai virus and other new RNA viruses in wild bumblebee hosts**

(Bombus pascuorum, Bombus lapidaries and Bombus pratorum). J Invertebr Pathol 2016, **134**:6–11.

- 15. Peng W, Li J, Boncristiani Jr HF, Strange JP, Hamilton M, Chen YP: **Host range** expansion of honey bee Black Queen Cell Virus in the bumble bee, *Bombus huntii*. *Apidologie* 2011, **42**:650-658.
- 16. Piot N, Snoeck S, Vanlede M, Smagghe G, Meeus I: **The effect of oral administration of** dsRNA on viral replication and mortality in *Bombus terrestris Viruses* 2015, **7**:3172-8.
- 17. Ravoet J, De Smet L, Meeus I, Smagghe G, Wenseleers T, de Graaf DC: **Widespread** occurrence of honey bee pathogens in solitary bees. *J Invertebr Pathol* 2014, 122:55-58.
- 18. Reynaldi FJ, Sguazza GH, Albicoro FJ, Pecoraro MR, Galosi CM: **First molecular detection of co-infection of honey bee viruses in asymptomatic** *Bombus atratus* **in South America**. *Braz J Biol.* 2013, **73**:797-800.
- 19. Sachman-Ruiz B, Narváez-Padilla V, Reynaud E: **Commercial Bombus impatiens as** reservoirs of emerging infectious diseases in central Mexico. *Biol Invasions* 2015, **17**:2043-2053.
- 20. Singh R, Levitt AL, Rajotte EG, Holmes EC, Ostiguy N, van Engelsdorp S, Lipkin WI, dePamphillis CW, Toth AL, Cox-Foster DL: **RNA viruses in hymenopteran pollinators: evidence of inter-taxa virus transmission via pollen and potential impact on non** *Apis* **hymenopteran species**. *PLoS ONE* 2010, **5**:e14357.