

Dystrophin ∆78 Cter Dystrophin +78 Cter Cter Nte O Cter PEPFOLD PEPFOLD s TPG к P MR R % of 0.1 0 secondary structure 8.0 -9.0 80 Coil 4 0.4 Sheet 0.2 Helix 20 0 0 SRGRNTPGKPMRE С Dystrophin +78 Cter Dystrophin ∆78 Cter polar non-polar total % non-polar polar non-polar total % non-polar model1 1119,1 941,4 177,7 15,9 1705,0 786,7 2491,7 31,6 model2 984,4 194,1 1178,5 16,5 1427,7 821,1 2248,8 36,5 208,5 957,3 1423,3 649,9 2073,3 model3 1165,9 17.9 31,3 model4 1050,0 187,9 1237,9 15,2 1443,1 746,6 2189,7 34,1 1003,8 195,1 1198,9 1223,8 704,9 1928,7 36,5 model5 16,3 average 987,4 192,7 1180,0 16,3 1444.6 741,9 2186,4 34,0

Supplementary Figure 1: a) Schematic representation of DMD exon 78 splicing and C-ter tail amino acid sequence in absence or presence of exon 78. b) Five models provided by PEP-fold program for the structure of dystrophin +78 and dystrophin Δ 78 Cter tails. c) Molecular surface hydrophobicity of all PEP-fold models.

1,0

171,2

67,3

209,9

2,5

b

std

42,5

11,2

43,6

а



Supplementary Figure 2: a) qRT-PCR analysis of *dmd* mRNA levels in control embryos and *dmd*-exon 78 morphants at 48 hpf (from 3 independent experiments). **b)** RT-PCR analysis of μ Dys-CTL and μ Dys- Δ ex78 mRNA levels in TA injected muscles (n=5). **c)** qRT-PCR analysis of μ Dys-CTL and μ Dys- Δ ex78 mRNA level in TA injected muscles (n=8). **d)** Percentage of dystrophin positive fibers in μ Dys-CTL and μ Dys- Δ ex78 injected muscles (n=5). **c)** and (n=5). Bars indicate s.e.m and "ns" indicates not significant; Student *t-test*.



Supplementary Figure 3: a) gRT-PCR analysis of total Dmd mRNA level in U7-Dmd exon 78 injected muscles compared to saline-injected muscles, 6 months after injection (n=10). b) Tandem mass Spectrum (MS/MS) of the precursor ion at m/z 581.25 determining oxidized peptide ³⁶⁶⁵GHNVGSLFHMoxADDLGR³⁶⁸⁰. The fragmentation Spectrum shown is a trypsinderived peptide. The inset show the peptide sequence and the observed ions obtained in the MS/MS Spectrum labeled to show singly and doubly charged b and y ions, as well as ion corresponding to neutral losses of water (circles) and NH3 (asteriks); M, parent ion mass. c) Dystrophin N-terminal domain (ex10-11; Manex1011B antibody), Dystrophin C-terminus domain (ex78-79; Dys2 antibody), α -syntrophin and α -dystrobrevin immunostaining of TA muscles injected with U7-Dmd exon 78 compared to saline-injected TA muscles (scale bar, 50µm). d) Absolute maximal muscle force (P0) of U7-Dmd exon 78 TA injected muscles compared to saline-injected muscles (n=10). e) TA muscles weight of U7-Dmd exon 78 injected muscles compared to saline-injected muscles (n=10). f) Specific maximal force (sP0) of U7-Dmd exon 78 TA injected muscles compared to saline-injected muscles (n=10). g) Maximal cross-section area of U7-Dmd exon 78-injected TA muscles compared to saline injected-TA muscles (n=10). Bars indicate s.e.m and "ns" indicates not significant; * indicates p<0.05; ** indicates p<0.01; paired *t*-test.



Supplementary Figure 4: Upper panel: Another example of disorientated myofibrils (pseudocolored in green) within an oxidative myofiber: a band of myofibrils running at right angle to the fiber major direction traverse the whole fiber. **Lower panel:** Dilated sarcoplasmic reticulum in a fast twitch fiber. Arrows point triads with swollen reticular terminal cisternae Scale bar, 10µm in the right panels and scale bar, 1µm in the left panels.

Primers for splicing		
	primer FW	primer Rev
human DMD ex78	TTAGAGGAGGTGATGGAGCA	GATACTAAGGACTCCATCGC
mouse <i>Dmd</i> ex 78	TGGTTGGCAGTCAAACTTCA	TCATCTGCCATGTGGAAAAG
zebrafish dmd ex78	CCCAGGATGCAAGCACTGGATTAG	TTACATGAACCAGCGACTCC
Primers for RT-PCR		
	primer FW	primer Rev
mouse µDys-CTL	CTCTCAGACCAGCGAGAGCA	CTCCCGCATGGGCTTGCCGG
mouse μDys-Δ78	CTCTCAGACCAGCGAGAGCA	ACTCCATCGCTCTGCCCAAA
Primers for qRT-PCR		
	primer FW	primer Rev
zebrafish <i>elfa</i>	CTTCTCAGGCTGACTGTGC	CCGCTAGCATTACCCTCC
zebrafish dmd	GGAGCTGACGTCTCACCAG	TGCTCTGTCGCTCCATACTG
mouse <i>Rlp0</i>	GAGGACCTCACTGAGATTCGG	TTCTGAGCTGGCACAGTGAC
mouse <i>Dmd</i>	TGGATCTGACATCTCATCAAGGAC	CCATGCTAGCTACCCTGAGAC
mouse μDys	AGGCAGAGCACCAGAAACTACC	CTGGCACTTGGCGATGTTGAAG
mouse Myh1	GCGAGGTTCACACCAAAATCA	TGGTCACTTTCCTGCTTTGGA
mouse Myh2	AAGCGAAGAGTAAGGCTGTC	GTGATTGCTTGCAAAGGAAC
mouse Myh4	ACAAGCTGCGGGTGAAGAGC	CAGGACAGTGACAAAGAACG

Supplementary Table S1: Primer sequences used for RT-PCR and qRT-PCR analysis.



Figure 1e (right panel)



Figure 2a



Supplementary Figure 2b



Supplementary Figure 5: Uncropped PCR gels

Figure 1e (left panel)



Figure 1f



Figure 4a



Supplementary Figure 2b

